

Dichloroacetate ameliorates myocardial ischemia-reperfusion injury via regulating autophagy and glucose homeostasis

Shuo Li^{1,2}, Boqia Xie³, Wenqian Zhang¹, Tong Li¹

¹Department of Thoracic Surgery, Beijing Chao-Yang Hospital, Capital Medical University, Beijing, China

²Department of Thoracic Surgery, The Affiliated Hospital of QingDao University, Qingdao, China

³Department of Cardiology, Beijing Chao-Yang Hospital, Capital Medical University, Beijing, China

Submitted: 13 January 2019; **Accepted:** 21 July 2019

Online publication: 27 August 2019

Arch Med Sci 2023; 19 (2): 420–429

DOI: <https://doi.org/10.5114/aoms.2019.87503>

Copyright © 2019 Termedia & Banach

Corresponding author:

Dr. Bo-qia Xie

Department of Cardiology
Beijing Chao-Yang
Hospital

Capital University
of Medical Science

8 Gongtinan Road

Beijing 100020, China

Phone: +86 10-85231000

Fax: +86 10-85231000

E-mail:

dr.boqiaxie@hotmail.com

Abstract

Introduction: Autophagy occurs in response to myocardial ischemia-reperfusion (I/R) injury and plays both a protective and detrimental role in this pathological process. Dichloroacetate (DCA) has a protective role during myocardial I/R injury, but its effects on autophagy induction are less well understood. We established a rat myocardial I/R injury model and investigated the effects of DCA on autophagy induction.

Material and methods: Healthy Sprague-Dawley (SD) rats were used to establish a myocardial I/R model. The effect of DCA on cardiac infarct size, cardiac function, and serum levels of serum creatine kinase (CK), lactic dehydrogenase (LDH) and cardiac troponin I (cTnI) during I/R injury were assessed. Using cultured rat cardiac myocytes, we investigated the functional implications of DCA and explored the relevant mechanism(s) of DCA protection during I/R injury.

Results: DCA significantly decreases infarct size and arrhythmia scores, increases heart rate (HR), left ventricular developed pressure (LVDP), the maximal rate of pressure rise (+dp/dtmax) and coronary flow (CF), and reduces the levels of CK, LDH and cTnI after I/R. DCA treatment enhanced myocardial cell viability and alleviated apoptosis assessed through TUNEL assays and Bcl-2/Bax expression. Notably, the levels of the autophagy biomarkers Beclin-1, LC3II and reactive oxygen species (ROS) significantly decreased following DCA treatment. When Bcl-2 was silenced, Beclin-1 expression significantly decreased following DCA treatment, suggesting that DCA confers a protective role during I/R injury through autophagy regulation. Moreover, DCA was found to regulate glucose homeostasis through increasing the expression of Glut-1 and Glut-4.

Conclusions: DCA contributes to I/R injury protection through the regulation of autophagy and glucose homeostasis.

Key words: ischemia-reperfusion injury, dichloroacetate, autophagy, glucose metabolism.

Introduction

Ischemic heart disease is characterized by high morbidity and mortality and is a major cause of death worldwide [1]. Although rapid reperfusion is considered the most effective therapy for myocardial infarction

[2], it can induce injury during the early stages, termed myocardial ischemia/reperfusion (I/R) injury. It was reported that I/R injury disturbs ionic homeostasis, increases the production of reactive oxygen species (ROS) and inflammatory responses, induces mitochondrial disorders, and causes calcium overload [3].

Autophagy is a crucial house-keeping process that controls the degradation of long-lived proteins and damaged organelles, playing a vital role in I/R in the myocardium [4]. Autophagy is induced in response to stress conditions in the heart, including cardiac muscle hypertrophy and heart failure, and is regarded as a prominent feature of cardiovascular dysfunction [5–11]. It has been shown that autophagy exerts a dual function during I/R injury in the myocardium. On one hand, autophagy compensates for mitochondrial injury, providing support to proteostasis and the survival of damaged cardiomyocytes following I/R injury [12–14]. However, sustained autophagy during reperfusion can induce cell death through the digestion of essential proteins and organelles. Matsui *et al.* indicated that autophagy plays a protective role during ischemia, but can detrimentally affect the heart during reperfusion [15]. Regarding the early reperfusion of ischemic myocardium, a modest increase in autophagy protects against necrocytosis due to the degradation of damaged organelles and the provision of free fatty acids (FFA) and amino acids, but sustained up-regulation of autophagy drives cardiac cell apoptosis and/or injury, leading to cell death. In addition, autophagy is an important regulator of glucose homeostasis. He *et al.* revealed that autophagy controlled by exercise-induced Bcl-2 is essential for muscle glucose homeostasis, and that Bcl-2-knockout mice fail to increase GLUT4 localization at the plasma membrane, reducing glucose uptake [16]. Roy *et al.* revealed that TBC1D5 shuttling is dependent on autophagy as it controls the plasma membrane translocation of GLUT1 [17].

It has been reported that dichloroacetate (DCA) activates pyruvate dehydrogenase and stimulates glucose aerobic metabolism in perfused hearts [18, 19]. DCA also reduces oxygen consumption and lactate production [20]. These data demonstrate that DCA ameliorates cardiac dysfunction by depleting lactic acidosis [21]. Stacpoole *et al.* [22] reported that DCA elevates systolic pressure and cardiac output for those suffering from hypotension and lactic acidosis. Moreover, Burns *et al.* [23] suggested that DCA stimulates pyruvate dehydrogenase and restores myocardial ATP levels to augment myocardial protection in rat hearts. However, the role of DCA in I/R through its ability to regulate autophagy has not been reported.

In this study, we hypothesized that DCA may alleviate I/R injury through the inhibition of autophagy.

We further investigated the potential mechanism(s) involved in the regulation of autophagy by DCA.

Material and methods

Myocardial ischemia-reperfusion model *in vivo*

Healthy Sprague-Dawley (SD) rats weighing 250–300 g were purchased from the Institute of Laboratory Animal Sciences, Peking Union Medical College (Beijing, China). Rats were raised individually in plastic cages under conditions of a 12 h light/dark cycle (temperature at $23 \pm 1^\circ\text{C}$; 12 h light/dark cycle, light on at 7 a.m.). Thirty SD rats were randomly divided into three groups: the control group (normal saline), the I/R group (normal saline and I/R) and the I/R + DCA (3 mM) group. Each group consisted of 12 rats. The study group was anesthetized with 3% sodium pentobarbital (i.p.) (60 mg/kg) and treated with heparin (i.p.) (150 U/kg). The rats in the control group underwent a sham operation. Myocardial I/R was produced in rats of the I/R group according to previous studies [24]. Briefly, the BI-OPAC multi-guide biometric device was used to test the ECG with needle electrodes installed under the skin. A catheter was inserted into the carotid artery to take blood samples upon completion of the experiment. Following intubation, rats were ventilated mechanically using a rodent ventilator (SAR-830, USA). The chest was opened, the pericardium was incised, and the heart was visible. Subsequently, the left anterior descending coronary artery was ligated to perform coronary artery occlusion, which was confirmed by ST segment elevation, an increase in R-wave amplitude and epicardial cyanosis. At 30 min post-occlusion, a snare was used to release the coronary artery occlusion, and the heart was reperfused for 120 min. Additionally, reperfusion was confirmed by the recovery from cyanosis and the inversion of ECG changes [25]. After reperfusion, hearts were separated and stained with 1% (w/v) triphenyl tetrazolium chloride (TTC) to assess infarct size. All procedures were strictly conducted in accordance with the national (D.Ln.26, March 4, 2014) and international regulations and policies (directive 2010/63/EU). This protocol was permitted by the Animal Experimental Ethics Committee, Beijing Chao-Yang Hospital, Capital University of Medical Science.

Evaluation of cardiac function

Arrhythmia was defined and determined based on the criteria of the Lambeth Conventions [26]. ECG recordings were analyzed during the first 30 min of reperfusion for ventricular arrhythmias. Arrhythmias were categorized into 6 groups and their severity scored [27]: 0 = no arrhythmia; 1 = less than 10 s of ventricular tachycardia or oth-

er arrhythmias, with no ventricular fibrillation (VF); 2 = 11 s to 30 s of ventricular tachycardia or other arrhythmias, with no VF; 3 = 31 s to 90 s of ventricular tachycardia or other arrhythmias, with no VF; 4 = 91 s to 180 s of ventricular tachycardia or other arrhythmias, and/or less than 10 s of reversible VF; 5 = greater than 180 s of ventricular tachycardia or other arrhythmias, and/or greater than 10 s of reversible VF; 6 = irreversible VF. In addition, the heart rate (HR), the left ventricular developed pressure (LVDP), and the maximal rate of pressure rise (+dp/dtmax) were recorded, and the coronary flow (CF) was measured to evaluate cardiac function.

Serum determination

After 120 min of reperfusion, 3 ml of blood was collected through the common carotid artery to detect the levels of serum creatine kinase (CK) (Abcam), lactic dehydrogenase (LDH) (Abcam) and cardiac troponin I (cTnI) (Abcam) by enzyme-linked immunosorbent assay (ELISA). Assays were performed according to the manufacturer's instructions.

Cardiac myocyte culture

Cardiac myocytes in neonatal rats were isolated and maintained as previously described [28]. Briefly, isolated cells were cultured in serum-free, glucose-free Dulbecco's modified Eagle's medium (DMEM) (Thermo Fisher Scientific, Shanghai, China) in the anoxia chamber (95% N₂, 5% CO₂) at 37°C for 24 h. Reperfusion was then initiated through the replacement of normal culture media containing 10% fetal bovine serum (FBS) (Gibco) in 5% CO₂ and 95% O₂ for 2 h. At the beginning of the reperfusion, different concentrations of DCA were used to treat cells and were maintained for 12 h. Cells were then harvested for specific assays.

Myocardial cell viability and apoptosis

To gain insight into the role of DCA in cardiac myocytes, cell viability and apoptosis assays were performed. Treated myocardial cells were seeded into 96-well plates at a density of 10,000 cells per hole and viability was assessed via MTT assay. For the assessment of apoptosis, TUNEL staining was performed according to the manufacturer's instructions. Apoptotic cells were counted in five random visual fields. The percentage of cells undergoing apoptosis was calculated using the following equation: Apoptosis % = number of apoptotic cells/total number of apoptotic cells × 100%.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR)

RNA isolation from myocardial cells was performed to analyze the expression of Bcl-2 and Bax

at the mRNA level using TaqMan assays (Life Technologies, CA, US). Tests were performed as previously described [29].

Immunofluorescence

For immunofluorescence, cardiac myocytes were first fixed with 4% paraformaldehyde (PFA) for 10 min. Cardiac myocytes were then permeabilized with 0.1% Triton X-100 for 10 min, blocked for 1 h with 1% bovine serum albumin (BSA), and incubated with anti-LC3 antibody (Abcam, dilution 1 : 100) and anti-Beclin-1 antibody (Abcam, dilution 1 : 100) for 12 h at 4°C. After incubation of primary antibodies, cardiac myocytes were incubated with anti-mouse IgG labeled with Alexa Fluor-488 for 1 h at 37°C and then stained with DAPI for 5 min. The immunofluorescence results were assessed under Laser confocal microscopy (Carl ZEISS, Oberkochen, Germany).

Intracellular ROS detection

The myocardium of the left ventricle was sectioned and fixed in 10% formalin, and frozen in liquid nitrogen prior to use. Assays were performed as previously described [30]. ROS positive cells were detected by DCFH-DA (Beyotime, China). Using fluorescence microscopy, cell images were obtained and ROS fluorescence intensity was quantified using Image J software (NIH, Bethesda, Maryland, USA).

siRNA treatment

Hairpin siRNA was used to silence Bcl-2 expression in myocardial cells. siRNA was synthesized by Thermo Fisher. Bcl-2 siRNA: 5'-GGGAGAACAG-GGTACGATA-3', scrambled control siRNA: 5'-GG-GAAACGGTGCGAGAGATA-3'. Following the transient transfection of myocardial cells with siRNA, cells were incubated under normal conditions. Samples were then prepared for further assays.

Western blot

Total proteins from cultured cells and the left ventricle were extracted according to our protocols. Western blot assays were conducted as previously described. Obtained proteins were incubated with anti-Beclin-1 (1 : 1000, Cell Signaling Technology), anti-Bcl-2 (1 : 1000, Cell Signaling Technology), anti-LC3 (1 : 1000, Cell Signaling Technology), anti-Glut1 (1 : 1000, Abcam) and anti-Glut4 primary antibodies (1 : 100, Abcam).

Statistical analysis

All data analysis was performed using SAS 9.3 software (SAS Institute, USA) and a *p*-value less than 0.05 was considered statistically significant.

Normally distributed data with homogeneous variance were expressed as the mean \pm standard deviation (SD). A one-way analysis of variance was performed for multiple group comparisons, followed by post-hoc analysis for group-wise comparisons.

Results

Effects of DCA on the myocardial I/R model *in vivo*

To assess whether DCA can alleviate myocardial I/R injury, we compared the cardiac infarct size,

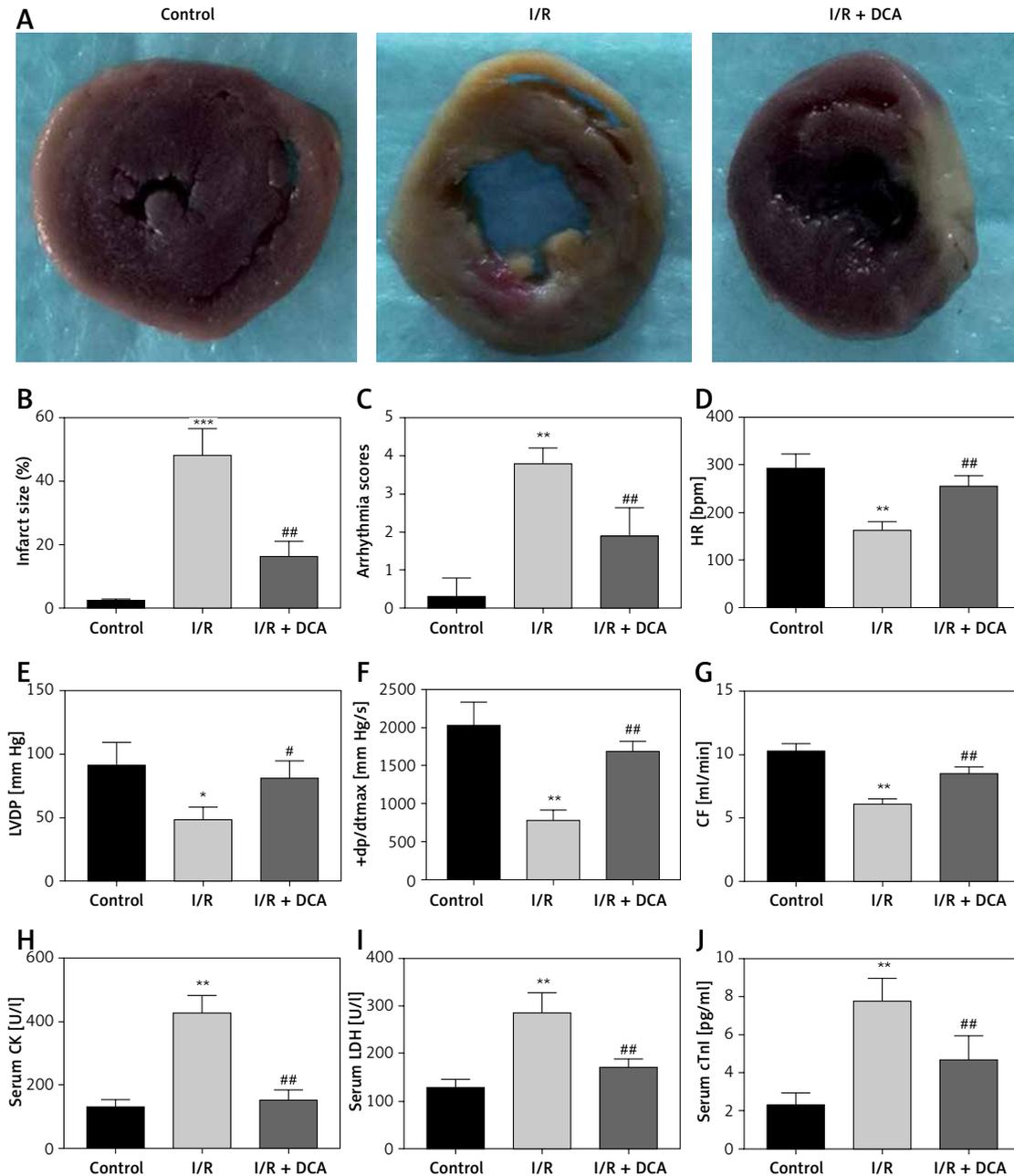


Figure 1. Evaluation of arrhythmia and myocardial damage indicators *in vivo*. **A** – TTC staining results of hearts in control, ischemia/reperfusion (I/R) and I/R+ dichloroacetate (DCA) groups ($n = 4$); **B** – infarct size of hearts in the three groups ($n = 4$); **C** – arrhythmia score changes in the three groups ($n = 4$); **D** – heart rate (HR) changes in the three groups ($n = 4$); **E** – left ventricular developed pressure (LVDP) changes in the three groups ($n = 4$); **F** – the maximal rate of pressure rise (+dp/dtmax) changes in the three groups ($n = 4$); **G** – coronary flow (CF) changes in the three groups ($n = 4$); **H** – I/R groups display higher creatine kinase (CK) levels compared to the control group ($n = 4$); **I** – serum lactic dehydrogenase (LDH) ($n = 4$); **J** – serum cardiac troponin I (cTnI) ($n = 4$). I/R represents ischemia/reperfusion group. Rats in the I/R + DCA group had ischemia/reperfusion and were treated with DCA

*** $P < 0.01$ compared to the control group. ** $P < 0.01$ compared to the I/R group.

cardiac arrhythmia scores, HR, LVDP, +dp/dtmax, CF and serum myocardial injury markers during reperfusion in the three experimental groups. The infarct size in the I/R group was significantly higher than the control group, while DCA significantly reduced the infarct size compared to the I/R group (Figures 1 A, B). Notably, arrhythmia scores in the I/R groups were dramatically higher in comparison to the control group, but treatment with DCA strongly reduced the arrhythmia score, suggesting that it reverses the effects of IR (Figure 1 C). The HR, LVDP, +dp/dtmax, and CF were significantly lower in the I/R group compared to the control group, while they were significantly higher after treatment with DCA compared to the I/R group (Figures 1 D–G). In addition, we also observed an apparent increase in the serum levels of CK, LDH and cTnI in the I/R group, which was reversed by DCA treatment (Figures 1 H–J). Collectively, these results confirmed that DCA exerts a protective role in myocardial I/R models *in vivo*.

In vitro effects of DCA on myocardial cell viability and apoptosis

We next determined the viability of myocardial cells *in vitro* through MTT assays. Treated cells were divided into control, I/R, and I/R + DCA groups. The results showed that cell viability in

the I/R group was lower than the control group ($p < 0.01$) (Figure 2 A). Notably this effect was reversed by DCA treatment. When the concentration of DCA was 30 μM , cell viability improved (Figure 2 A). When myocardial cell apoptosis was determined by TUNEL assay, the percentage of apoptotic cells in the I/R group was strongly elevated compared to the control group. It was noteworthy that DCA treatment reversed these effects and 30 μM DCA significantly alleviated cell apoptosis (Figure 2 B). In addition, when the expression of Bcl-2 and Bax was assessed at the mRNA level, qRT-PCR assays indicated that the Bcl-2/Bax ratio in the I/R group was strongly down-regulated, whilst DCA recovered the Bcl-2/Bax ratio (Figure 2 C). Taken together, these observations demonstrate that DCA promotes cell viability and inhibits apoptosis following I/R.

DCA attenuates I/R injury by suppressing reperfusion-induced autophagy

To explore the role of DCA in the regulation of autophagy, we analyzed its effects on Beclin-1 and LC3-II expression, considered reliable autophagy biomarkers. Western blot analysis indicated that Beclin-1 level ($p < 0.05$, Figures 3 A, B) and the ratio of LC3-II/LC3-I ($p < 0.05$, Figures 3 A, C) were significantly higher in the I/R group in comparison to

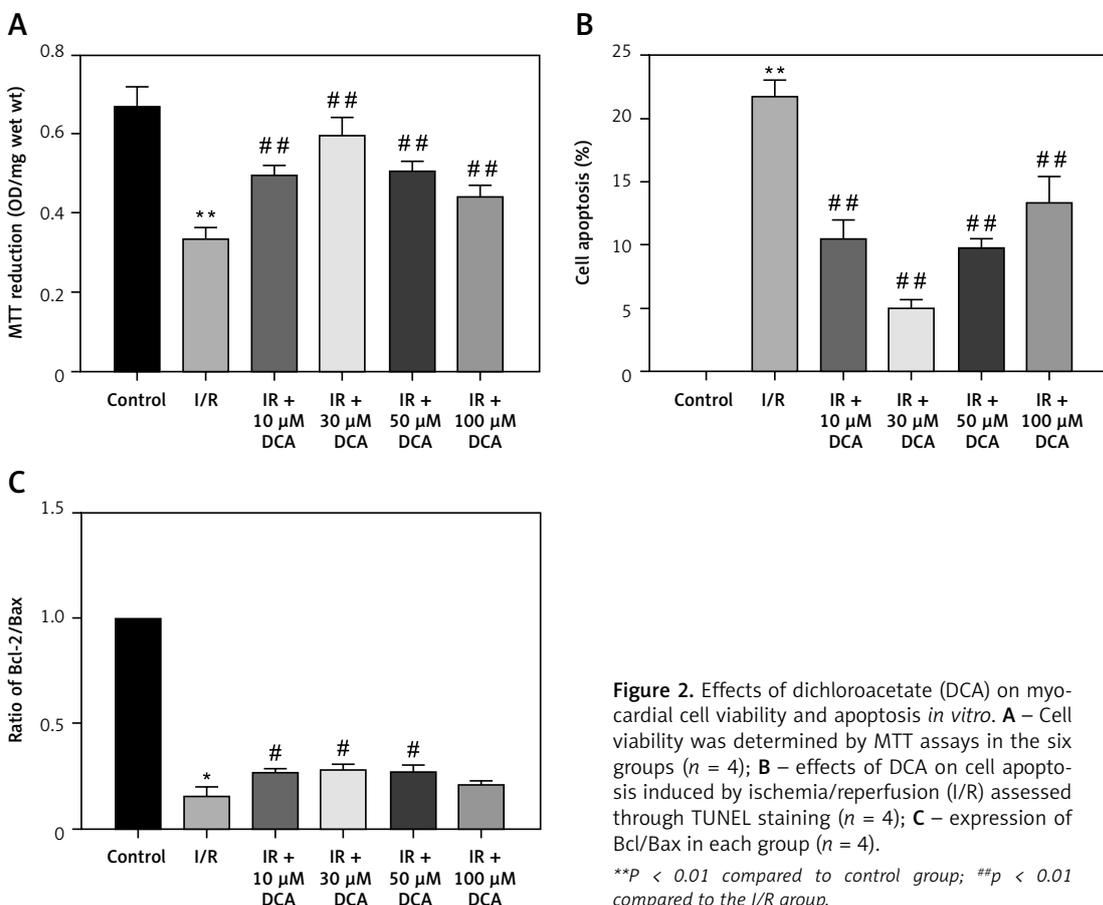


Figure 2. Effects of dichloroacetate (DCA) on myocardial cell viability and apoptosis *in vitro*. **A** – Cell viability was determined by MTT assays in the six groups ($n = 4$); **B** – effects of DCA on cell apoptosis induced by ischemia/reperfusion (I/R) assessed through TUNEL staining ($n = 4$); **C** – expression of Bcl/Bax in each group ($n = 4$).

** $P < 0.01$ compared to control group; ## $p < 0.01$ compared to the I/R group.

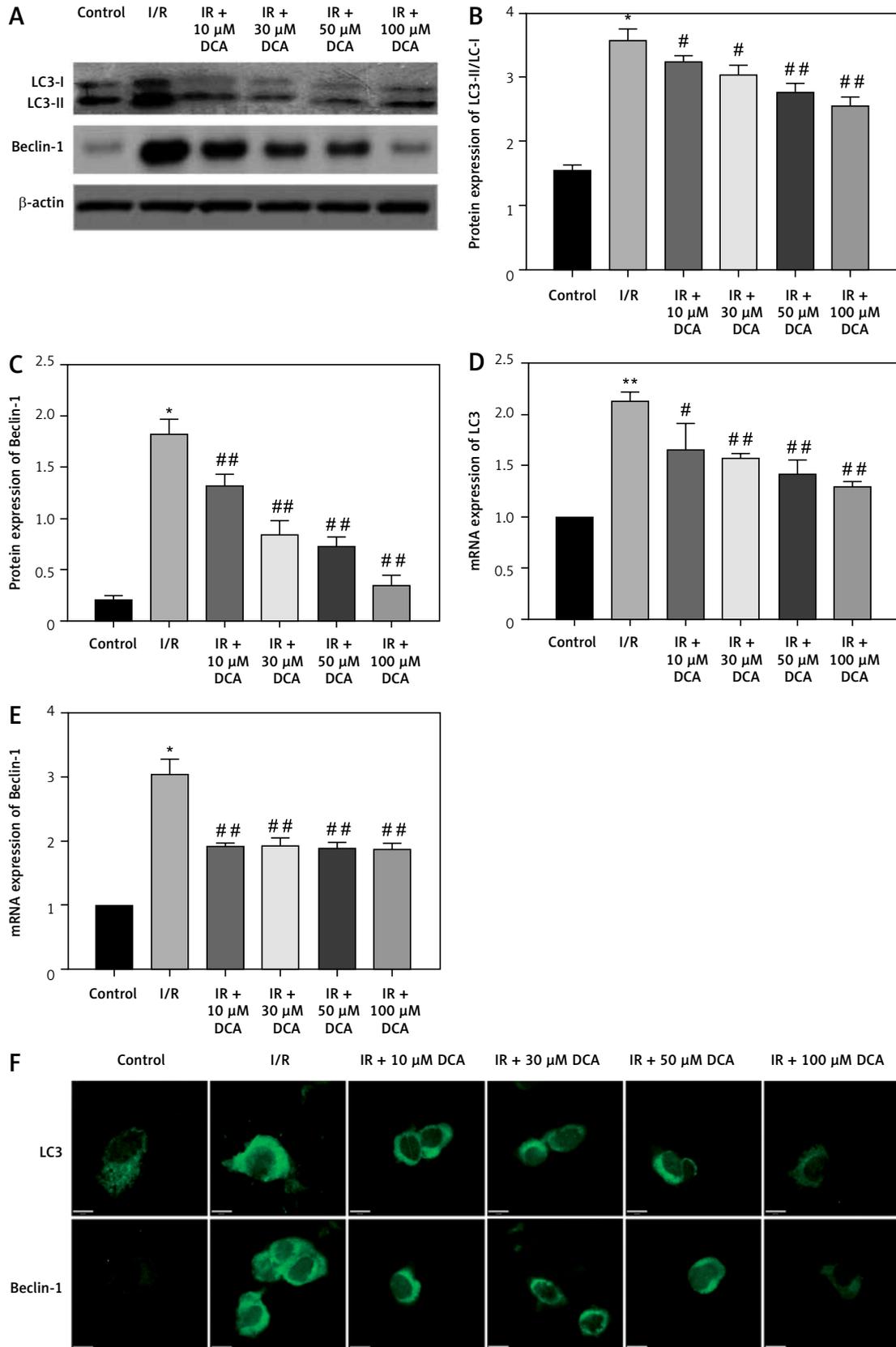


Figure 3. Dichloroacetate (DCA) attenuates ischemia/reperfusion (I/R) injury by suppressing reperfusion-induced autophagy. **A** – Western blot analysis of LC3-II/LC3-I and Beclin-1 following DCA treatment ($n = 4$); **B** – relative expression of LC3-II/LC3-I ($n = 4$); **C** – Beclin-1 expression was determined by western blot ($n = 4$); **D** – LC3, **E** – Beclin-1 mRNA levels in the six groups ($n = 4$); **F** – immunofluorescence results of LC3 and Beclin-1 in the six groups ($n = 4$)

* $P < 0.05$ compared to control group. # $P < 0.05$ compared to I/R group. ## $P < 0.01$ compared to I/R group.

the control group. Conversely, this effect was significantly inhibited in a concentration-dependent manner by DCA treatment (Figures 3 A–C). These findings were confirmed at the mRNA level through qRT-PCR (Figures 3 D, E). In addition, the immunofluorescence results also showed that the expression of LC3 and Beclin-1 increased in the I/R group and decreased in the DCA treated group (Figure 3 F). Collectively, these results revealed that DCA inhibits reperfusion-induced autophagy through the down-regulation of potential indicators.

Effects of DCA on autophagy through ROS and Bcl-2

It has been reported that ROS accumulation drives the oxidative destruction of DNA, proteins and lipids, triggering cell damage and apoptosis [24]. ROS accumulation is also essential for autophagy induction under conditions of starvation or stress [25]. To assess the effects of ROS in the DCA-mediated inhibition of autophagy, we assessed the levels of ROS in myocardial cells using DCFH-DA. The results showed that ROS production increased after I/R, which was significantly reduced by DCA (Figure 4 A). These effects were

lost following treatment with N-acetyl-L-cysteine (NAC), a classical ROS inhibitor.

Hairpin siRNA was used to silence Bcl-2 and western blot analysis revealed that Beclin-1 expression decreased following DCA treatment. A significant difference in the LC3-II/LC3-I levels following Bcl-2 knock-down was not observed (Figures 4 B, C). These observations suggest that ROS and Bcl-2 contribute to DCA-mediated autophagy inhibition.

Effects of DCA on myocardial glucose homeostasis *in vivo*

We finally assessed the effects of DCA on myocardial glucose homeostasis after reperfusion *in vivo*. Data from both western blot and qRT-PCR assays revealed upregulation of Glut-1 and Glut-4 at both the protein and mRNA level in left ventricular tissue following DCA treatment Figures 5 A–E. This suggested that DCA promotes glucose uptake.

Discussion

Ischemic heart disease involving myocardial infarction leads to serious health issues. Rapid reper-

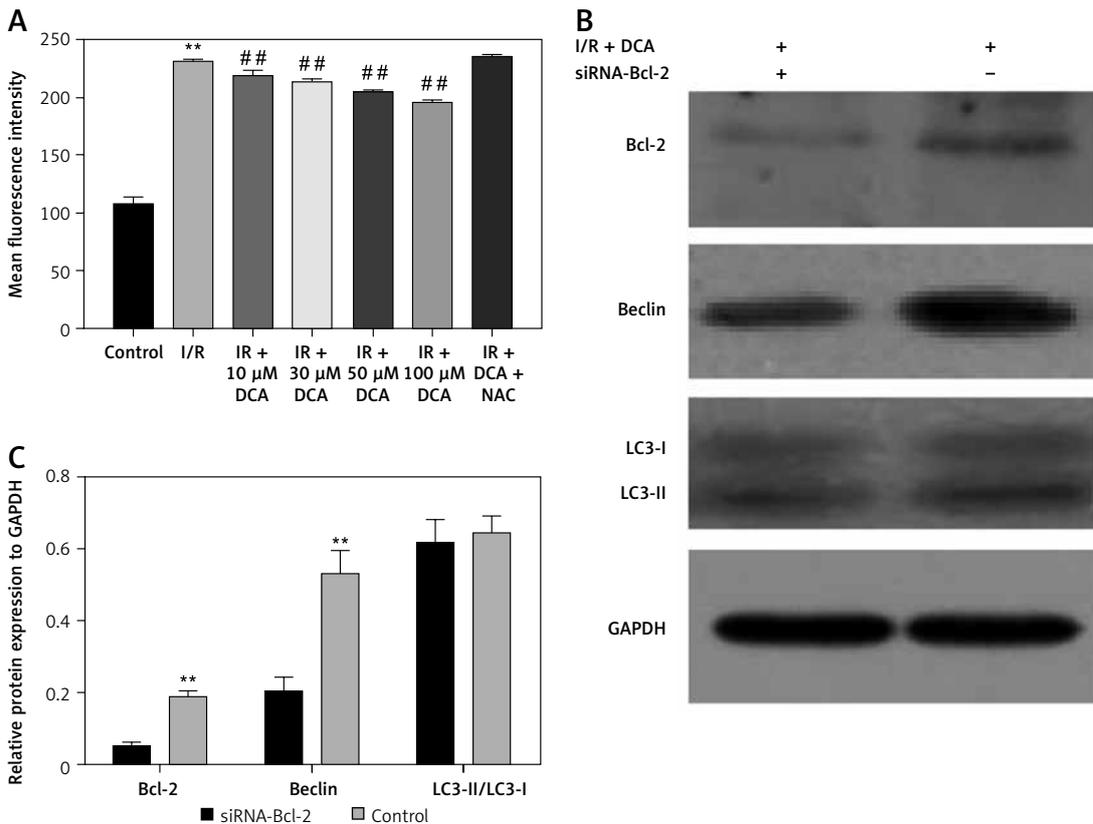


Figure 4. Effects of dichloroacetate (DCA) on autophagy through reactive oxygen species (ROS) and Bcl-2. **A** – ROS generation was detected by DCFH-DA in myocardial cells ($n = 4$); **B** – expression levels of Bcl-2, LC3-II/LC3-I and Beclin-1 after Bcl-2 siRNA silencing in ischemia/reperfusion (I/R) + DCA groups ($n = 4$); **C** – band intensities from western blots were calculated and normalized to GAPDH levels ($n = 4$). Protein expression is expressed as the fold change relative to experimental controls

* $P < 0.1$, ** $p < 0.01$.

fusion of the ischemic myocardium remains the fundamental therapy for myocardial infarction [2]. Importantly, an additional injury, termed myocardial I/R, occurs during the initial period. DCA can activate pyruvate dehydrogenase and improves cardiac function [31]. In this study, we explored the functional implications of DCA after I/R injury *in vitro* and *in vivo* and identified a protective role of DCA against reperfusion injury through the modulation of autophagy and glucose homeostasis.

Autophagy is an intracellular bulk degradation process that has the capacity to degrade and recycle cytosolic and long-lived proteins and organelles [4, 32, 33]. Autophagy occurs at low levels and is enhanced by stressors such as nutrient depletion [34]. Recent studies have shown that autophagy has both beneficial and adaptive roles depending

on the context and its magnitude of induction [35]. Autophagy is also implicated in I/R injury. At low levels, autophagy can prevent cell injury following I/R. In high concentrations or through sustained up-regulation, autophagy can promote cardiac cell apoptosis or cell injury through Beclin-1 [36]. This study indicated that DCA improves cardiac function, increases cell viability and inhibits apoptosis. Furthermore, the first evidence that DCA can inhibit Beclin-1 expression and influence the LC3-II/LC3-I ratio is provided, suggesting that DCA reduces autophagy induction during the reperfusion period.

Growing evidence demonstrates that ROS-activated autophagy occurs during myocardial reperfusion [37, 38]. ROS functions as an essential signaling molecule that modulates a series of biological processes, including gene expression,

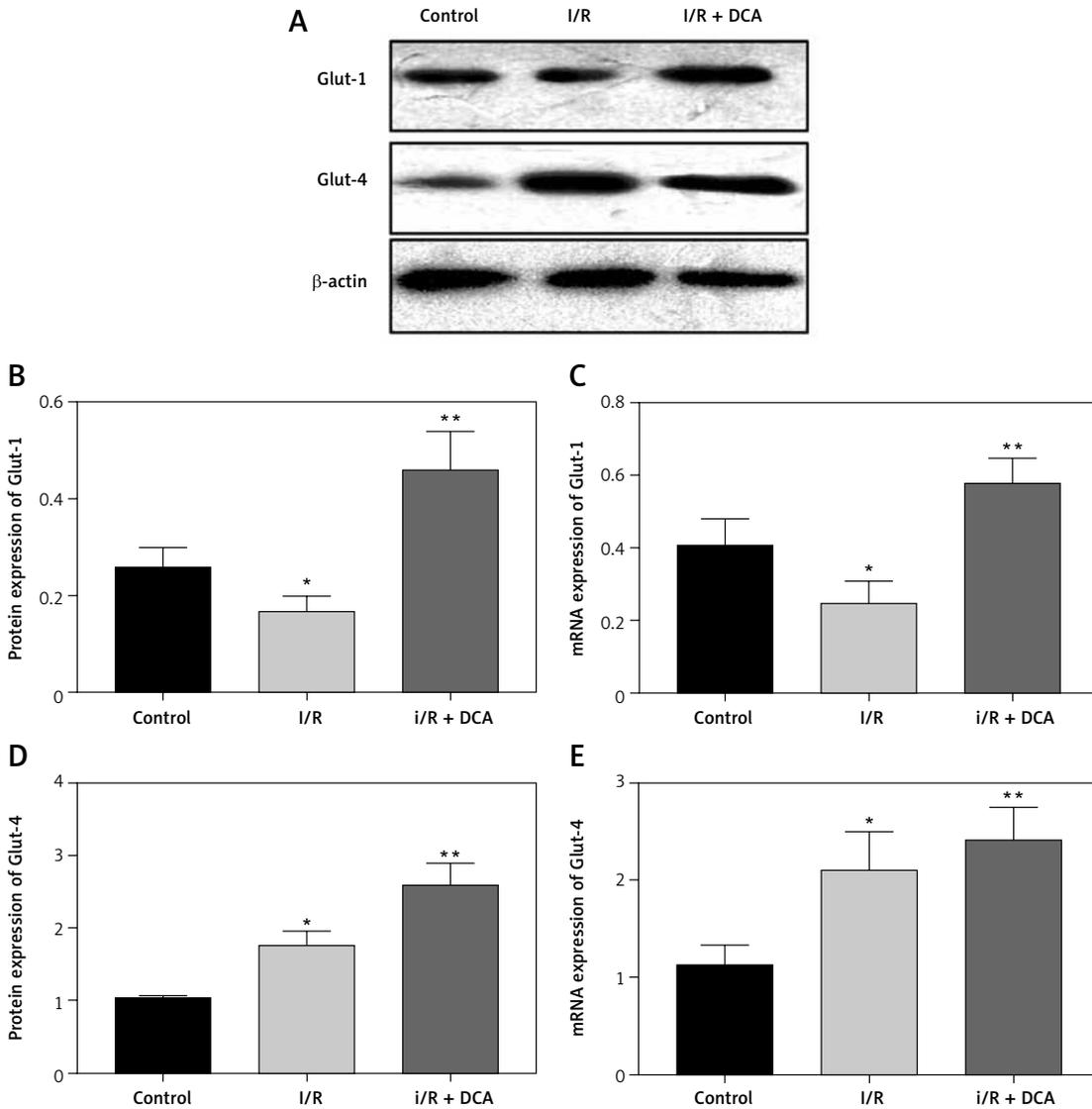


Figure 5. Effects of dichloroacetate (DCA) on myocardial glucose homeostasis *in vivo*. Glut-1 (A, B) and Glut-4 (A, C) expression levels were detected by western blot analysis in the three groups. Glut-1 (D) and Glut-4 (E) mRNA levels were also determined by RT-PCR ($n = 4$)

* $P < 0.05$ compared to the control group. ** $P < 0.01$ compared to the control group.

cell metabolism, growth and survival. However, excessive ROS is detrimental to cellular homeostasis, leading to cell damage and apoptosis [24, 39]. It has been reported that autophagy is up-regulated through ROS accumulation during myocardial reperfusion. In this study, we found that DCA treatment strongly inhibited ROS production during myocardial reperfusion, but NAC (an ROS inhibitor) had the capacity to abolish this effect. Combined with the aforementioned data, these results indicate that ROS contributes to DCA-mediated autophagy inhibition, promoting myocardial cell protection during the reperfusion period.

It has been demonstrated that Bcl-2 is involved in apoptotic signaling. Interestingly, Bcl-2 family members regulate a variety of biological effects, including morphogenesis, glucose homeostasis, and autophagy [16, 40, 41]. Moreover, previous studies have shown that Bcl-2 decreases the levels of starvation-induced autophagy *in vitro* and *in vivo*, indicating its key role in autophagy regulation [42, 43]. In this *in-vitro* study, DCA significantly alleviated apoptosis by up-regulation of the Bcl-2/Bax ratio. Following Bcl-2 silencing, Beclin-1 was inhibited by DCA treatment, suggesting that it contributes to the ability of DCA to regulate autophagy. We also found that Bcl-2 silencing does not influence the LC3-II/LC3-I ratio, a finding that warrants further investigation.

It was recently reported that myocardial efficiency is reduced when the levels of FFA are elevated [44, 45]. We therefore predict that myocardial glucose abundance decreases oxygen consumption to improve myocardial efficiency. In addition, autophagy regulates glucose homeostasis [16]. We found that DCA dramatically enhanced Glut-1 and Glut-4 expression, raising the possibility that DCA represses autophagy to improve myocardial energy metabolism.

In conclusion, DCA regulates autophagy and glucose homeostasis to improve I/R injury. Importantly, we revealed the utility of DCA as a therapeutic strategy for I/R injury.

Acknowledgments

Shuo Li and Boqia Xie contributed equally to this article.

This study was supported by the National Natural Science Foundation of China (grant number 81400268).

Conflict of interest

The authors declare no conflict of interest.

References

- Fang L, Fan X, Yu Z, et al. Cardioprotection by combination of three compounds from ShengMai preparations in mice with myocardial ischemia/reperfusion injury through AMPK activation-mediated mitochondrial fission. *Sci Rep* 2016; 6: 37114.
- Guo J, Wang SB, Yuan TY, et al. Coptisine protects rat heart against myocardial ischemia/reperfusion injury by suppressing myocardial apoptosis and inflammation. *Atherosclerosis* 2013; 231: 384-91.
- Bystrom P, Foley N, Toledo-Pereyra L, et al. Ischemic preconditioning modulates ROS to confer protection in liver ischemia and reperfusion. *EXCLI J* 2017; 16: 483.
- Thapalia BA, Zhou Z, Lin X. Autophagy, a process within reperfusion injury: an update. *Int J Clin Exp Pathol* 2014; 7: 8322-41.
- Jensen BC, Bultman SJ, Holley D, et al. Upregulation of autophagy genes and the unfolded protein response in human heart failure. *Int J Clin Exp Med* 2017; 10: 1051-8.
- Ren J, Sowers J R, Zhang Y. Metabolic stress, autophagy, and cardiovascular aging: from pathophysiology to therapeutics. *Trends Endocrinol Metab* 2018; 29: 699-711.
- Miyata S, Takemura G, Kawase Y, et al. Autophagic cardiomyocyte death in cardiomyopathic hamsters and its prevention by granulocyte colony-stimulating factor. *Am J Pathol* 2006; 168: 386-97.
- Li Z, Wang J, Yang X. Functions of autophagy in pathological cardiac hypertrophy. *Int J Biol Sci* 2015; 11: 672-8.
- Shimomura H, Terasaki F, Hayashi T, Kitaura Y, Isomura T, Suma H. Autophagic degeneration as a possible mechanism of myocardial cell death in dilated cardiomyopathy. *Jpn Circ J* 2001; 65: 965-8.
- Zhu H, Tannous P, Johnstone JL, et al. Cardiac autophagy is a maladaptive response to hemodynamic stress. *J Clin Invest* 2007; 117: 1782-93.
- Ma S, Wang Y, Chen Y, Cao F. The role of the autophagy in myocardial ischemia/reperfusion injury. *Biochim Biophys Acta* 2015; 1852: 271-6.
- Tannous P, Zhu H, Nemchenko A, et al. Intracellular protein aggregation is a proximal trigger of cardiomyocyte autophagy. *Circulation* 2008; 117: 3070-8.
- Quinsay MN, Thomas RL, Lee Y, Gustafsson AB. Bnip3-mediated mitochondrial autophagy is independent of the mitochondrial permeability transition pore. *Autophagy* 2010; 6: 855-62.
- Ma X, Liu H, Foyil SR, Godar RJ, Weinheimer CJ, Diwan A. Autophagy is impaired in cardiac ischemia-reperfusion injury. *Autophagy* 2012; 8: 1394-6.
- Matsui Y, Takagi H, Qu X, et al. Distinct roles of autophagy in the heart during ischemia and reperfusion roles of AMP-activated protein kinase and beclin 1 in mediating autophagy. *Circ Res* 2007; 100: 914-22.
- He C, Bassik MC, Moresi V, et al. Exercise-induced BCL2-regulated autophagy is required for muscle glucose homeostasis. *Nature* 2012; 481: 511-5.
- Roy S, Leidal AM, Ye J, Ronen SM, Debnath J. Autophagy-dependent shuttling of TBC1D5 controls plasma membrane translocation of GLUT1 and glucose uptake. *Mol Cell* 2017; 67: 84-95.e5.
- Jaimes R, Kuzmiak-Glancy S, Brooks DM, et al. Functional response of the isolated, perfused normoxic heart to pyruvate dehydrogenase activation by dichloroacetate and pyruvate. *Pflügers Archiv* 2016; 468: 131-42.
- Kato T, Niizuma S, Inuzuka Y, et al. Analysis of metabolic remodeling in compensated left ventricular hypertrophy and heart failure. *Circ Heart Fail* 2010; 3: 420-30.
- Niewisch MR, Kuçi Z, Wolburg H, et al. Influence of dichloroacetate (DCA) on lactate production and oxygen consumption in neuroblastoma cells: is DCA a suitable

- drug for neuroblastoma therapy?. *Cell Physiol Biochem* 2012; 29: 373-80.
21. Barbee RW, Kline JA, Watts JA. Depletion of lactate by dichloroacetate reduces cardiac efficiency after hemorrhagic shock. *Shock* 2000; 14: 208-14.
 22. Stacpoole PW, Barnes CL, Hurbanis MD, Cannon SL, Kerr DS. Treatment of congenital lactic acidosis with dichloroacetate. *Arch Dis Child* 1997; 77: 535-41.
 23. Burns AH, Giaimo ME, Summer WR. Dichloroacetic acid improves in vitro myocardial function following in vivo endotoxin administration. *J Crit Care* 1986; 1: 11-7.
 24. Ran X, Diao JX, Sun XG, et al. Huangzhi oral liquid prevents arrhythmias by upregulating caspase-3 and apoptosis network proteins in myocardial ischemia-reperfusion injury in rats. *Evid Based Complement Alternat Med* 2015; 2015: 518926.
 25. Himori N, Matsuura A. A simple technique for occlusion and reperfusion of coronary artery in conscious rats. *Am J Physiol* 1989; 256: 1719-25.
 26. Walker MJA, Curtis MJ, Hearse DJ, et al. The Lambeth conventions: guidelines for the study of arrhythmias in ischaemia, infarction, and reperfusion. *Cardiovasc Res* 1988; 22: 447-55.
 27. Cai HB, Sun XG, Liu ZF, et al. Effects of dahuangzhechong pills on cytokines and mitogen activated protein kinase activation in rats with hepatic fibrosis. *J Ethnopharmacol* 2010; 132: 157-64.
 28. Kang JX, Xiao YF, Leaf A. Free, long-chain, polyunsaturated fatty acids reduce membrane electrical excitability in neonatal rat cardiac myocytes. *Proc Natl Acad Sci USA* 1995; 92: 3997-4001.
 29. Stather PW, Sylvius N, Wild JB, Choke E, Sayers RD, Bown MJ. Differential microRNA expression profiles in peripheral arterial disease. *Cir Cardiovasc Genet* 2013; 6: 490-7.
 30. Tao R, Kim SH, Honbo N, Karliner JS, Alano CC. Minocycline protects cardiac myocytes against simulated ischemia-reperfusion injury by inhibiting poly(ADP-ribose) polymerase-1. *J Cardiovasc Pharmacol* 2010; 56: 659-68.
 31. Piao L, Fang YH, Kubler MM, et al. Enhanced pyruvate dehydrogenase activity improves cardiac outcomes in a murine model of cardiac arrest. *PLoS One* 2017; 12: e0185046.
 32. Bai Y, Chen Y, Chen X, et al. Trichostatin A activates FOXO1 and induces autophagy in osteosarcoma. *Arch Med Sci* 2019; 15: 204-13.
 33. Pandurangan AK, Ismail S, Esa NM, et al. Inositol-6 phosphate inhibits the mTOR pathway and induces autophagy-mediated death in HT-29 colon cancer cells. *Arch Med Sci* 2018; 14: 1281-8.
 34. Lum JJ, Deberardinis RJ, Thompson CB. Autophagy in metazoans: cell survival in the land of plenty. *Nat Rev Mol Cell Biol* 2005; 6: 439-48.
 35. Schiattarella GG, Hill JA. Therapeutic targeting of autophagy in cardiovascular disease. *J Mol Cell Cardiol* 2015; 95: 86-93.
 36. Munasinghe PE, Riu F, Dixit P, et al. Type-2 diabetes increases autophagy in the human heart through promotion of Beclin-1 mediated pathway. *Int J Cardiol* 2016; 202: 13-20.
 37. Xu A, Szczepanek K, Maceyka MW, et al. Transient complex I inhibition at the onset of reperfusion by extracellular acidification decreases cardiac injury. *Am J Physiol Cell Physiol* 2014; 306: 1142-53.
 38. Godar RJ, Ma X, Liu H, et al. Repetitive stimulation of autophagy-lysosome machinery by intermittent fasting preconditions the myocardium to ischemia-reperfusion injury. *Autophagy* 2015; 11: 1537-60.
 39. Dunn JD, Alvarez LA, Zhang X, Soldati T. Reactive oxygen species and mitochondria: a nexus of cellular homeostasis. *Redox Biol* 2015; 6: 472-85.
 40. Danial NN, Korsmeyer SJ. Cell death: critical control points. *Cell* 2004; 116: 205-19.
 41. Cleland MM, Norris KL, Karbowski M, et al. Bcl-2 family interaction with the mitochondrial morphogenesis machinery. *Cell Death Differ* 2011; 18: 235-47.
 42. Levine B, Yuan J. Autophagy in cell death: an innocent convict? *J Clin Invest* 2005; 115: 2679-88.
 43. Pattingre S, Tassa A, Qu X, et al. Bcl-2 antiapoptotic proteins inhibit beclin 1-dependent autophagy. *Cell* 2005; 122: 927-39.
 44. Ma P, Han L, Lv Z, et al. In-hospital free fatty acids levels predict the severity of myocardial ischemia of acute coronary syndrome. *BMC Cardiovasc Disord* 2016; 16: 29.
 45. Wang Y, Zhang HW, Guo YL, et al. Free fatty acids as a marker for predicting periprocedural myocardial injury after coronary intervention. *Postgrad Med J* 2019; 95: 18-22.