

Evaluation of protective effects of GnRH agonist or antagonist on ovarian reserve with anti-Müllerian hormone and histological analysis in a rat model using cisplatin

Mustafa Tas¹, Gokalp Oner², Pasa Ulug³, Adem Yavuz⁴, Bulent Ozcelik⁵

¹Department of Obstetrics and Gynecology, Acibadem Mehmet Ali Aydinlar University, Acibadem Kayseri Hospital, Kayseri, Turkey

²Department of Obstetrics and Gynecology, Magnet Hospital, Kayseri, Turkey

³Department of Obstetrics and Gynecology, Ömer Halisdemir University, Niğde, Turkey

⁴Department of Obstetrics and Gynecology, Erzincan University, Erzincan, Turkey

⁵Department of Obstetrics and Gynecology, Erciyes University, Kayseri, Turkey

Submitted: 19 February 2019; Accepted: 1 August 2019

Online publication: 27 August 2019

Corresponding author:

Mustafa Tas

Department

of Obstetrics and

Gynecology

Acibadem University

Turkey

E-mail: drmustafatas@yahoo.com

Arch Med Sci 2023; 19 (2): 448–451

DOI: <https://doi.org/10.5114/aoms.2019.87540>

Copyright © 2019 Termedia & Banach

Abstract

Introduction: The aim of this prospective trial was to evaluate the ovarian reserve with anti-Müllerian hormone (AMH), which is the best predictor of ovarian reserve, and perform histological analysis after exposure to cisplatin with a GnRH agonist or antagonist.

Material and methods: Twenty-four Wistar albino rats were randomly divided into three groups, each consisting of eight rats. In the GnRH agonist group (group 1), rats received a single dose of 50 mg/m² cisplatin with 1 mg/kg triptorelin. In the GnRH antagonist group (group 2), rats received a single dose of 50 mg/m² cisplatin with 1 mg/kg cetrorelix. In the control group (group 3), rats received 50 mg/m² cisplatin. Ovarian reserve was assessed by AMH and histology.

Results: Primary follicle counts were higher in group 2 (4.50 ± 1.47 vs. 3.50 ± 1.70 vs. 3.00 ± 3.54) and secondary follicle counts were higher in group 1 (2.96 ± 1.11 vs. 1.74 ± 1.03 vs. 1.37 ± 3.11). Numbers of tertiary follicles were higher both in groups 1 and 2 than the control group (1.36 ± 0.83 vs. 0.84 ± 0.99 vs. 0.50 ± 0.75). The total follicle count of the study groups were significantly higher compared with the control group (14.32 ± 5.96 vs. 12.48 ± 4.12 vs. 10.63 ± 6.80). AMH was significantly higher in groups 1 and 2 compared with the control group (18.56 ± 25.33 vs. 16.48 ± 24.66 vs. 9.37 ± 26.54).

Conclusions: This is the first prospective randomized controlled study showing the protective effects of GnRH agonist and antagonist on ovarian reserve after cisplatin exposure in an animal model.

Key words: ovarian reserve, cisplatin, anti-Müllerian hormone.

Introduction

Chemotherapy is a pivotal part of cancer treatment and chemotherapy usage has been gradually increased in the world for this reason. It is known that chemotherapeutic agents damage ovaries and may cause ovarian failure and infertility [1]. Due to these damaging effects of chemotherapy on ovarian reserve, many prophylactic agents have been used to try to protect against destructive effects of chemotherapies on ovarian reserve [2].



Cisplatin was the most commonly used alkylating agent in gynecological cancers. Most of the studies revealed that GnRH agonist or antagonist usage with chemotherapeutic agents may protect ovarian reserve [3, 4]. Alkylating agents may affect ovaries with cortical fibrosis, blood vessel damage, and severe reduction in primordial follicles and ovarian cell proliferation. Also arrest and apoptosis may start in granulosa cells.

The fertility of women may be determined by the ovarian reserve tests including counting the number of ovarian follicles and measuring the level of anti-Müllerian hormone (AMH). The tests are the most widely used parameters in infertility clinics [5].

To date, there has been no study clearly investigating ovarian reserve with anti-Müllerian hormone (AMH) and histological analysis after cisplatin exposure with a GnRH agonist or antagonist in an animal model. In this prospective randomized controlled trial, we evaluated the effects of a GnRH agonist or antagonist on ovarian reserve of rats exposed to cisplatin, which was the most commonly used chemotherapeutic agent in gynecological cancers.

Material and methods

Twenty-five- to six-month-old female Wistar-Albino rats weighing 180–210 g were used in this study. All procedures were approved by Ethics Committee of Kobay Experimental Animal Laboratory. All procedures were carried out at Kobay Experimental Animal Laboratory. All rats were housed under controlled temperatures ($22 \pm 2^\circ\text{C}$) and 12/12 h light/dark cycle with food and water ad libitum.

Twenty-four Wistar albino rats were randomly divided into three groups, each consisting of eight rats. In the GnRH agonist group (group 1), rats received a single dose of 50 mg/m² cisplatin with 1 mg/kg Triptorelin. In the GnRH antagonist group (group 2), rats were received single dose 50 mg/m² cisplatin with 1 mg/kg Cetrorelix. In the control group (group 3), rats received 50 mg/m² cisplatin.

Blood samples (2 ml) were collected from the heart of the rats before laparotomy. The serum of the blood was separated from cells by centrifugation, and samples were frozen at -20°C until assayed. AMH levels (ng/ml) were determined by using ELISA (Cusabio Rat AMH kit; Biotek Synergy HT Microplate Reader, US) in Duzen Laboratory by a researcher blinded to the groups. All samples were tested in the same assay. The coefficients of intra- and inter-assay variations were 4.8% and 7.5%, respectively.

Ovarian samples were randomly prepared in 5-μm slices to assess follicles [6]. Hematoxylin-eosin was used for staining of the samples. The same

pathologist blindly examined the ovarian follicles and noted the counts of the follicles. Primordial, primary, secondary and tertiary follicles were defined according to the usual standard textbooks of microscopic anatomy [6].

Statistical analysis

The Statistical Package for the Social Sciences version 15.0 was used for the statistical analyses (SPSS Inc., Chicago, IL, USA) and groups were compared with the *t* test. $P < 0.05$ was considered statistically significant. Values were expressed as mean \pm standard deviation.

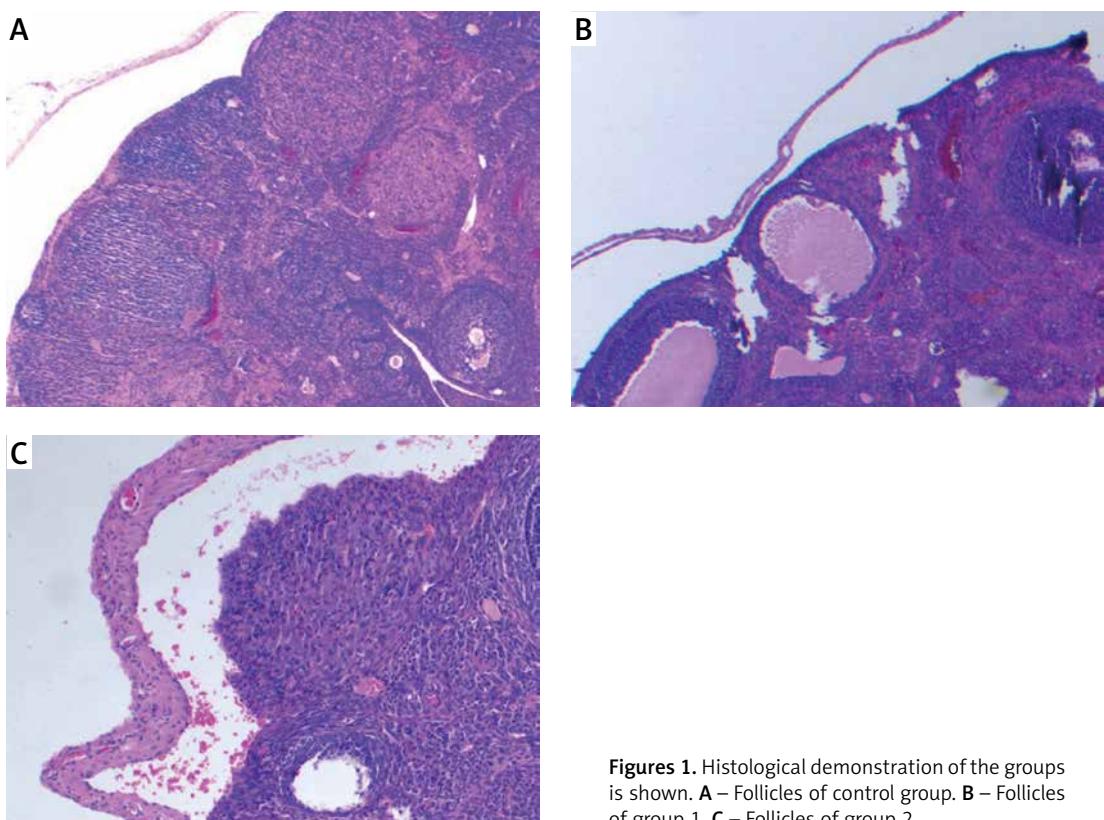
Results

Ovarian reserve was assessed by AMH and histology. There was no difference in the primordial follicle count between the groups. Primary follicle counts were higher in group 2 (4.50 ± 1.47 vs. 3.50 ± 1.70 vs. 3.00 ± 3.54) and secondary follicle counts were higher in group 1 (2.96 ± 1.11 vs. 1.74 ± 1.03 vs. 1.37 ± 3.11). Tertiary follicle counts were higher both in group 2 and 3 compared to the control group (1.36 ± 0.83 vs. 0.84 ± 0.99 vs. 0.50 ± 0.75). The total follicle count of the study groups was significantly higher compared with the control group (14.32 ± 5.96 vs. 12.48 ± 4.12 vs. 10.63 ± 6.80). AMH was found to be significantly higher in groups 1 and 2 compared with the control group (18.56 ± 25.33 vs. 16.48 ± 24.66 vs. 9.37 ± 26.54). Histological examination of the groups is demonstrated in Figure 1. There is no difference between the protective effect of GnRH agonist and GnRH antagonist (Table I).

Discussion

Fertility and premature menopause are major concerns for young women who are undergoing treatment for cancer. Premature ovarian failure caused by chemotherapy depends on type and dosage of the agents. Alkylating agents such as cisplatin are widely used chemotherapeutic agents and cause ovarian damage. Cisplatin has a heavy metal combination and antineoplastic effects on rapidly divided cells [7]. In this prospective randomized study, we evaluated how to decrease the detrimental effect of cisplatin on ovaries.

There are lots of experimental and clinical studies on protective agents such as gonadotropin-releasing hormone (GnRH) agonists and GnRH antagonists [8–10]. Our previous study showed that a GnRH agonist with cisplatin may protect ovarian reserve compared to solely cisplatin [11]. Controversially after this study meta-analyses showed that GnRH agonists had no effect to protect ovarian reserve against chemotherapy [12]. Then a committee opinion suggested that the data on



Figures 1. Histological demonstration of the groups is shown. **A** – Follicles of control group. **B** – Follicles of group 1. **C** – Follicles of group 2

Table I. Results of ovarian follicle tests and endometrium thickness of the groups

Parameter	Control group 3	Group 1 (GnRH agonist)	Group 2 (GnRH antagonist)
Primordial follicles	5.76 ±6.97 ^a	6.50 ±3.32 ^a	5.50 ±4.33 ^a
Primary follicles	3.00 ±3.54 ^a	3.50 ±1.70 ^{a,b}	4.50 ±1.47 ^b
Secondary follicles	1.37 ±3.11 ^a	2.96 ±1.11 ^b	1.74 ±1.03 ^{a,b}
Tertiary follicles	0.50 ±0.75 ^a	1.36 ±0.83 ^b	0.84 ±0.99 ^b
Total follicles	10.63 ±6.80 ^a	14.32 ±5.96 ^b	12.48 ±4.12 ^{a,b}
AMH [ng/ml]	9.37 ±26.54 ^a	18.56 ±25.33 ^b	16.48 ±24.66 ^b

There is no statistically significant difference between groups sharing the same letter. All data sets of power of performed test with $\alpha = 0.050$ (power of the study): 0.976–1.000.

the use of GnRH analogs for ovarian suppression have been conflicting in terms of the efficacy, so other fertility preservation options should be offered in addition to GnRH analog treatment [13]. But the studies and meta-analyses evaluated different types and dosages of chemotherapeutic agents. Additionally application of GnRH analogs was different. In our study we added a GnRH analog on the same day as cisplatin.

The protective mechanism of the GnRH agonists on the ovaries after exposure to cytotoxic effects on ovaries involves the GnRH receptors at the gonadal and pituitary levels. A GnRH agonist may downregulate the receptors of gonads and suppress primordial follicle development [14]. Additionally, the hypoestrogenic state decreases

ovarian perfusion and delivery of chemotherapy to the ovaries and a direct effect of the GnRH agonist on the ovary occurs independently of the gonadotropin level may be discussed for the protection of ovarian germ cells [15, 16]. Our study showed that GnRH agonist administration has a protective effect on ovarian reserve compared with the control group.

Recently, GnRH antagonists have been used to decrease the effects of chemotherapy on ovaries [17, 18]. A GnRH antagonist shows higher competitive blockade of the receptors than a GnRH agonist. Additionally a GnRH antagonist inhibits apoptosis triggered by chemotherapy [17]. To our knowledge, there has been no study comparing the protective effects of a GnRH agonist and antagonist on ova-

ries exposed to chemotherapy. Our study was the first to show the effects of a GnRH agonist and antagonist on ovaries after application of cisplatin, which was the most commonly used chemotherapeutic agent in gynecological malignancies.

Various ovarian reserve tests have been used to predict the ovarian response and pregnancy outcomes in couples. Two of most widely used tests are AMH levels and AFC [19, 20]. AMH is an increasingly used reliable marker of present ovarian reserve and the number of follicles remaining in ovaries [21]. In this study, ovarian reserve tests such as AMH and histological follicles count were used.

This study clearly demonstrated that cisplatin had a cytotoxic effect on the ovaries and combination with a GnRH agonist and antagonist had statistically significant protective effects on the total follicle count but not the primordial follicles. Although primordial follicles did not significantly change, primary, secondary and tertiary follicles might be protected with a GnRH agonist and antagonist. Additionally, AMH levels supported this situation and increased with the GnRH agonist and antagonist.

In conclusion, ovarian reserve was first evaluated after exposure to cisplatin with a GnRH agonist and antagonist in an animal randomized prospective study using most common ovarian reserve tests including AMH and histological examination of ovarian follicles. These data represent an important benefit for understanding the effect of GnRH agonists and antagonists on ovarian reserve. Also, both GnRH agonists and antagonists have limited protective effects on ovarian reserve.

Conflict of interest

The authors declare no conflict of interest.

References

1. Gadducci A, Cosio S, Genazzani AR. Ovarian function and childbearing issues in breast cancer survivors. *Gynecol Endocrinol* 2007; 23: 625-31.
2. Yeh J, Kim BS, Liang YJ, Peresie J. Gonadotropin stimulation as a challenge to calibrate cisplatin induced ovarian damage in the female rat. *Reprod Toxicol* 2009; 28: 556-62.
3. Ozcelik B, Turkyilmaz C, Ozgun MT, et al. Prevention of paclitaxel and cisplatin induced ovarian damage in rats by a gonadotropin-releasing hormone agonist. *Fertil Steril* 2010; 93: 1609-14.
4. Bedaiwy MA, Abou-Setta AM, Desai N, et al. Gonadotropin-releasing hormone analog cotreatment for preservation of ovarian function during gonadotoxic chemotherapy: a systematic review and meta-analysis. *Fertil Steril* 2011; 95: 906-14.
5. Yeh J, Kim B, Peresie J, Liang YJ, Arroyo A. Serum and ovarian Müllerian inhibiting substance, and their decline in reproductive aging. *Fertil Steril* 2007; 87: 1227-30.
6. Ulug P, Oner G. Evaluation of the effects of single or multiple döse methotrexate administration, salpingec-
- tom on ovarian reserve of rat with the measurement of anti-Müllerian hormone (AMH) levels and histological analysis. *Eur J Obstet Gynecol Reprod Biol* 2014; 181: 205-9.
7. Kail NG, McGuire WP. Chemotherapy for advanced epithelial ovarian carcinoma. *Best Pract Res Clin Obstet Gynaecol* 2002; 16: 553-71.
8. Blumenfeld Z. Endocrine prevention of chemotherapy-induced ovarian failure. *Future Oncol* 2016; 12: 1671-4.
9. Blumenfeld Z. How to preserve fertility in young women exposed to chemotherapy? The role of GnRH agonist cotreatment in addition to cryopreservation of embryos, oocytes, or ovaries. *Oncologist* 2007; 12: 1044-54.
10. Lemos CN, Reis FM, Pena GN, Silveira LC, Camargos AF. Assessment of fertility protection and ovarian reserve with GnRH antagonist in rats undergoing chemotherapy with cyclophosphamide. *Reprod Biol Endocrinol* 2010; 8: 51.
11. Ataya KM, McKenna JA, Weintraub AM, Clark MR, Le-Maire WJ. A luteinizing hormone-releasing hormone agonist for the prevention of chemotherapy-induced ovarian follicular loss in rats. *Cancer Res* 1985; 45: 3651-6.
12. Elgindy E, Sibai H, Abdelghani A, Mostafa M. Protecting ovaries during chemotherapy through gonad suppression: a systematic review and meta-analysis. *Obstet Gynecol* 2015; 126: 187-95.
13. Ethics Committee of the American Society for Reproductive Medicine. Fertility preservation and reproduction in cancer patients. *Fertil Steril* 2005; 83: 1622-8.
14. Blumenfeld Z, Avivi I, Eckman A, Epelbaum R, Rowe JM, Dann EJ. Gonadotropin-releasing hormone agonist decreases chemotherapy-induced gonadotoxicity and premature ovarian failure in young female patients with Hodgkin lymphoma. *Fertil Steril* 2008; 89: 166-73.
15. Sutcliffe SB. Cytotoxic chemotherapy and gonadal function in patients with Hodgkin's disease. *JAMA* 1979; 242: 1898-901.
16. Shenns RJ. Gonadal dysfunction. In: *Cancer – Principles & Practice of Oncology*. DeVita VT Jr, Hellman S, Rosenberg SA (eds.). J.B. Lippincott Co., Philadelphia 1993; 2395-406.
17. Zhao XJ, Huang YH, Yu YC, Xin XY. GnRH antagonist cetrorelix inhibits mitochondria-dependent apoptosis triggered by chemotherapy in granulosa cells of rats. *Gynecol Oncol* 2010; 118: 69-75.
18. Meirav D, Assad G, Dor J, Rabinovici J. The GnRH antagonist cetrorelix reduces cyclophosphamide-induced ovarian follicular destruction in mice. *Hum Reprod* 2004; 19: 1294-9.
19. Cook CL, Siow Y, Taylor S, Fallat ME. Serum müllerian-inhibiting substance levels during normal menstrual cycles *Fertil Steril* 2000; 73: 859-61.
20. Maheshwari A, Fowler P, Bhattacharya S. Assessment of ovarian reserve – should we perform tests of ovarian reserve routinely? *Hum Reprod* 2006; 21: 2729-35.
21. Rajpert-De Meyts E, Jorgensen N, Graem N, Müller J, Cate RL, Skakkebaek NE. Expression of anti-Müllerian hormone during normal and pathological gonadal development: association with differentiation of Sertoli and granulosa cells. *J Clin Endocrinol Metabol* 1999; 84: 3836-44.