

Effect of lymphocyte-to-monocyte ratio on survival in septic patients: an observational cohort study

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Abstract

Introduction: The purpose of the present study was to evaluate the potential relationship of lymphocyte-to-monocyte ratio (LMR) with outcomes of septic patients at intensive care unit (ICU) admission.

Material and methods: 3087 septic patients were included in the final cohort by using the Medical Information Mart for Intensive Care (MIMIC) database. We evaluated the association of different groups of LMR_{max} with 28-day survival and 1-year survival via Kaplan-Meier (K-M) analysis and Cox regression analysis. Subgroups analysis of LMR_{max} was performed to further explore the effect of LMR_{max} on survival.

Results: According to the optimal cut-off value, the cohort was divided into low-LMR_{max} and high-LMR_{max} groups. The 28-day and 1-year survival rates were 47.9% and 19.9%, respectively, in the low-LMR_{max} group, and 60.4% and 25.9%, respectively, in the high-LMR_{max} group. Univariate logistic regression and K-M analyses revealed that the 28-day and 1-year survival rates of the high-LMR_{max} group were higher than those of the low-LMR_{max} group (both $p < 0.001$). A subgroup analysis of LMR_{max} identified a significant stepwise decrease in the risk of death at 28 days and 1 year from group 1 to group 4 (LMR_{max} increased gradually) after adjustment for multiple variables.

Conclusions: We report for the first time that a lower LMR_{max} value is independently predictive of a poor prognosis in septic patients. Therefore, as an inexpensive and readily available indicator, LMR_{max} may facilitate stratification of prognosis in septic patients.

Key words: critical illness, intensive care unit, lymphocyte-to-monocyte ratio, sepsis, infection.

Introduction

Sepsis, defined as severe organ dysfunction caused by a dysregulated host response to infection, has a high incidence and high mortality rate worldwide [1–4]. Several conventional indicators, such as procalcitonin (PCT) and C-reactive protein (CRP), may reflect the severity of infection [5, 6]. The Simplified Acute Physiology Score (SAPS) and the Sequential Organ Failure Assessment (SOFA) score play important roles in evaluat-

ing the severity and prognosis of critical illnesses [7, 8]. Notably, the neutrophil-to-lymphocyte ratio (NLR) and the platelet-to-lymphocyte ratio (PLR) are novel predictors of the prognosis of sepsis [9–13].

Similarly, the lymphocyte-to-monocyte ratio (LMR), which can be easily derived from the complete blood count, is closely correlated with the severity and prognosis of several clinical diseases. A decreased LMR is reportedly predictive of a poor prognosis of acute ischaemic stroke (AIS) [14, 15]. Also, the inflammatory response participates in the pathophysiology of AIS [16]. However, no previous study has explored the relationship between LMR and the prognosis of sepsis in a large population.

The SAPS and SOFA scores allow assessment of the prognosis of septic patients; however, the subitems concerning white blood cell (WBC) components, which change significantly during an inflammatory response, are incorporated into neither. Therefore, we evaluated the relationship of the LMR with the short- and long-term outcomes of sepsis in critically ill patients at the time of admission to the intensive care unit (ICU).

Material and methods

Patients

Data for this study were extracted from the Medical Information Mart for Intensive Care (MIMIC) clinical database (v. 1.4), which contains comprehensive deidentified clinical data of patients admitted to the Beth Israel Deaconess Medical Center in Boston, Massachusetts [17]. MIMIC III contains data on more than 58,000 distinct adult and neonate admissions to critical care units from 2001 to 2012. The Institutional Review Board of the Beth Israel Deaconess Medical Center and Massachusetts Institute of Technology approved the use of the MIMIC III database for authorised users who have completed the required training (Wei Zhou; ID: 25222342). The requirement for individual patient consent was waived for this study because it did not impact clinical care and all health information was deidentified [17].

Patients diagnosed with sepsis according to the criteria of Angus *et al.* were enrolled [18]. We selected patients with severe sepsis, ICD-9 codes for bacterial or fungal infection, and a diagnosis of acute organ dysfunction. Only patients aged 18 years or older with lymphocyte and monocyte counts obtained at ICU admission were included. The exclusion criteria were as follows: (1) date of death unclear, (2) definite haematological malignancies, (3) repeat ICU admissions, and (4) incomplete patient data for multivariate analysis.

Data collection

The data extracted from the MIMIC III database comprised gender, age, results of laboratory tests, comorbidities, special treatments during ICU stay, and ICU length of stay (LOS). The SAPS and SOFA scores on admission were calculated from the database [7, 8]. The baseline laboratory data and illness scores were evaluated during the first 24 h of the ICU stay.

Target and outcome variables

We recorded the complete blood count of each patient during the first 24 h of ICU admission. The LMR was calculated as the absolute lymphocyte count to absolute monocyte count ratio. The maximum value of the LMR (LMR_{max}) was the target variable.

The primary end point was 28-day survival and the secondary end point was 1-year survival. Mortality was calculated based on dates of admission and death, obtained from the MIMIC III database.

Statistical analysis

Numerical variables were evaluated for a normal distribution using the Kolmogorov-Smirnov normality test. Non-normally distributed data are presented as medians with interquartile ranges. Categorical variables are presented as frequency with percentage. Differences in the non-normally distributed variables were evaluated using the Wilcoxon rank-sum test. The Pearson χ^2 test and the Fisher exact test were used for analysing categorical variables. By receiver operating characteristic (ROC) curve analysis, the cut-off value of 3.0 (taking the integer of the cut-off value in two ROC curves), which was confirmed using Youden's index, was regarded as the optimal point for distinguishing low- LMR_{max} and high- LMR_{max} groups for further statistical analysis.

We assessed the association of LMR_{max} with the risk of death at 28 days and 1 year by univariate logistic regression. The results are expressed as odds ratios (ORs) and 95% confidence intervals (CIs). We generated Kaplan-Meier (K-M) curves to analyse the probability of survival and compared the results among the LMR_{max} groups using the log-rank test.

A Cox regression analysis was performed to identify whether LMR_{max} was independently associated with the prognosis of septic patients. The hazard ratio (HR) and 95% CI were calculated for the final model. After removing collinear factors, the following variables were adjusted for in the multivariable analysis: gender, age, laboratory test results (haemoglobin and lactate levels), SOFA and SAPS scores, alcohol abuse, comorbidities (congestive heart failure, cardiac arrhythmias, hy-

pertension, chronic pulmonary, renal failure, liver disease, solid tumour, and diabetes), and LOS in the ICU.

Two-sided p -values of < 0.05 were considered indicative of statistical significance. All statistical analyses were performed using SPSS software (ver. 20.0; IBM Corp., Armonk, NY).

Results

Baseline characteristics of participants

A total of 3,087 patients were included in this study. The baseline characteristics of the low- and high-LMR_{max} groups are listed in Table I. The

two groups were significantly different in terms of gender ratio and age. Compared to the low-LMR_{max} group, the high-LMR_{max} group had a lower WBC count, creatinine level, and SAPS score (all $p < 0.01$), and a longer duration of mechanical ventilation and ICU LOS (both $p < 0.01$). With regard to comorbidities, the high-LMR_{max} group exhibited lower frequencies of cardiac arrhythmias ($p < 0.01$), chronic pulmonary ($p < 0.05$), renal failure ($p < 0.01$) and solid tumour ($p < 0.05$) than the low-LMR_{max} group. Additionally, comparison of baseline characteristics of the study cohort vs. the missing data cohort is presented in Table II.

Table I. Baseline characteristics of the study participants

Characteristics	Total (N = 3087)	Low-LMR _{max} (≤ 3) (N = 1409)	High-LMR _{max} (> 3) (N = 1678)
Gender (men/women)	1664/1423	792/617	872/806 ^a
Age [years], n (%):			
> 20, ≤ 40	88 (2.9)	20 (1.4)	68 (4.0) ^b
> 40, ≤ 60	603 (19.5)	251 (17.8)	352 (21.0) ^a
> 60, ≤ 80	1322 (42.8)	617 (43.8)	705 (42.0)
> 80	1074 (34.8)	521 (37.0)	553 (33.0) ^a
Alcohol abuse, n (%)	231 (7.5)	108 (7.7)	123 (7.3)
Lab items:			
WBC [$\times 10^9/l$]	12.90 (8.60–18.30)	14.20 (10.30–20.10)	11.80 (7.50–16.80) ^b
Hemoglobin [g/dl]	10.90 (9.50–12.50)	10.80 (9.50–12.50)	10.90 (9.50–12.50)
Lactate [mmol/l]	2.20 (1.50–3.60)	2.10 (1.40–3.50)	2.20 (1.50–3.80)
Creatinine [g/dl]	1.40 (0.90–2.40)	1.50 (0.90–2.60)	1.40 (0.90–2.20) ^b
SOFA score	6 (4–9)	6 (4–9)	6 (4–9)
SAPS score	47 (38–57)	48 (39–58)	46 (38–56) ^b
Mechanical ventilation (first 24 h), n (%)	1811 (58.7)	812 (57.6)	999 (59.5)
Duration of mechanical ventilation [h]	24.00 (0–134.67)	18.03 (0–111.50)	31.00 (0–158.26) ^b
Vasopressor, n (%)	1657 (53.7)	750 (53.2)	907 (54.1)
Duration of vasopressor [h]	3.13 (0–38.97)	2.52 (0–38.79)	3.43 (0–39.10)
Comorbidities, n (%):			
Congestive heart failure	1250 (40.5)	578 (41.0)	672 (40.0)
Cardiac arrhythmias	1178 (38.2)	573 (40.7)	605 (36.1) ^b
Hypertension	1648 (53.4)	776 (55.1)	872 (52.0)
Chronic pulmonary	755 (24.5)	374 (26.5)	381 (22.7) ^a
Renal failure	707 (22.9)	361 (25.6)	346 (20.6) ^b
Liver disease	403 (13.1)	193 (13.7)	210 (12.5)
Solid tumor	184 (6.0)	98 (7.0)	86 (5.1) ^a
Diabetes	911 (29.5)	399 (28.3)	512 (30.5)
ICU LOS [days]	4.13 (1.97–8.91)	3.77 (1.89–7.70)	4.53 (2.02–10.17) ^b

ICU – intensive care unit, LMR – lymphocyte-to-monocyte ratio, LOS – length of stay, SAPS – Simplified Acute Physiology Score, SOFA – Sequential Organ Failure Assessment, WBC – white blood cells. Data were expressed as median (interquartile range) or frequency (percentage). ^a $P < 0.05$; ^b $p < 0.01$.

Table II. Baseline characteristics of study cohort and missing data cohort

Characteristics	Study cohort (N = 3087)	Missing data (N = 7627)	P-value
Gender (men/women)	1664/1423	4001/3626	0.175
Age [years], n (%):			
≤ 40	88 (2.9)	840 (11.0)	< 0.001
> 40, ≤ 60	603 (19.5)	2052 (26.9)	< 0.001
> 60, ≤ 80	1322 (42.8)	3118 (40.9)	0.065
> 80	1074 (34.8)	1617 (21.2)	< 0.001
Alcohol abuse, n (%)	231 (7.5)	679 (8.9)	0.017
SOFA score	6 (4–9)	5 (3–7)	< 0.001
SAPS II score	47 (38–57)	37 (29–47)	< 0.001
Mechanical ventilation (first 24 h), n (%)	1811 (58.7)	4206 (55.1)	0.001
Duration of mechanical ventilation [h]	24.00 (0–134.67)	19.50 (0–140.00)	0.162
Vasopressor, n (%)	1657 (53.7)	3261 (42.8)	< 0.001
Duration of vasopressor [h]	3.13 (0–38.97)	0 (0–21.75)	< 0.001
Comorbidities, n (%):			
Congestive heart failure	1250 (40.5)	2353 (30.9)	< 0.001
Cardiac arrhythmias	1178 (38.2)	2320 (30.4)	< 0.001
Hypertension	1648 (53.4)	3888 (51.0)	0.024
Chronic pulmonary	755 (24.5)	1564 (20.5)	< 0.001
Renal failure	707 (22.9)	1200 (15.7)	< 0.001
Liver disease	403 (13.1)	734 (9.6)	< 0.001
Solid tumor	184 (6.0)	349 (4.6)	0.003
Diabetes	911 (29.5)	2077 (27.7)	0.017
ICU LOS [days]	4.13 (1.97–8.91)	4.14 (1.99–10.19)	0.028

ICU – intensive care unit, LOS – length of stay, SAPS II – Simplified Acute Physiology Score II, SOFA – Sequential Organ Failure Assessment. Data were expressed as median (interquartile range) or frequency (percentage). Significant p-values are indicated in bold.

Association of LMR_{max} with 28-day and 1-year survival

The 28-day and 1-year survival rates were 47.9% and 19.9%, respectively, in the low- LMR_{max} group, and 60.4% and 25.9%, respectively, in the high- LMR_{max} group. Univariate logistic regression analyses revealed that the high- LMR_{max} group had higher 28-day and 1-year survival rates than the low- LMR_{max} group (both $p < 0.001$).

Furthermore, in the K-M analysis of 28-day and 1-year survival, patients with a high LMR_{max} had a more favourable prognosis than those with a low LMR_{max} (both $p < 0.001$) (Figure 1 A, B).

Subgroup analysis of LMR_{max}

Based on the values of LMR_{max} , the subjects were divided into four groups: group 1 ($LMR_{max} \leq 1$, $n = 203$), group 2 ($LMR_{max} > 1$ and ≤ 2 , $n = 572$), group 3 ($LMR_{max} > 2$ and ≤ 3 , $n = 634$), and group 4 ($LMR_{max} > 3$, $n = 1,678$). In univariate logistic regression analyses, there was a significant stepwise decrease in the risk of death at both 28 days and 1 year from group 1 to group 4 (Figures 2 A, B).

The K-M 28-day and 1-year survival curves suggested significant differences in survival probability among the four groups (both $p < 0.001$) (Figures 3 A, B).

Association of LMR_{max} with survival after multivariable adjustments

The association of LMR_{max} with survival after multivariable adjustments was evaluated by Cox regression analysis. As presented in Table III, after adjustment for gender, age, haemoglobin and lactate levels, SOFA and SAPS scores, alcohol abuse, comorbidities, and ICU LOS, the risk of death at 28 days (group 1: reference; group 2: $p = 0.001$, HR = 0.709; group 3: $p < 0.001$, HR = 0.556; group 4: $p < 0.001$, HR = 0.479) and 1 year (group 1: reference; group 2: $p = 0.001$, HR = 0.741; group 3: $p < 0.001$, HR = 0.599; group 4: $p < 0.001$, HR = 0.580) showed significant stepwise decreases from group 1 to group 4.

Discussion

In this study, septic patients with a high LMR_{max} were more likely to survive, and had a bet-

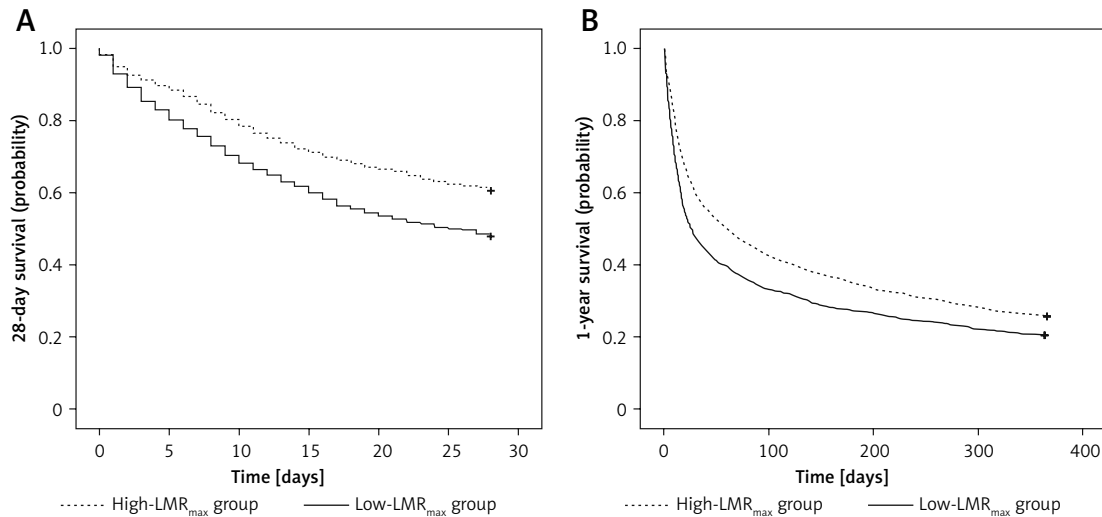


Figure 1. Kaplan-Meier (K-M) survival analysis for low-LMR_{max} and high-LMR_{max} group. **A** – 28-day survival curve; **B** – 1-year survival curve. The curves demonstrated that the patients with high LMR_{max} had higher probability of survival than those with low LMR_{max} at 28 days and 1 year (both $p < 0.001$)

LMR – lymphocyte-to-monocyte ratio.

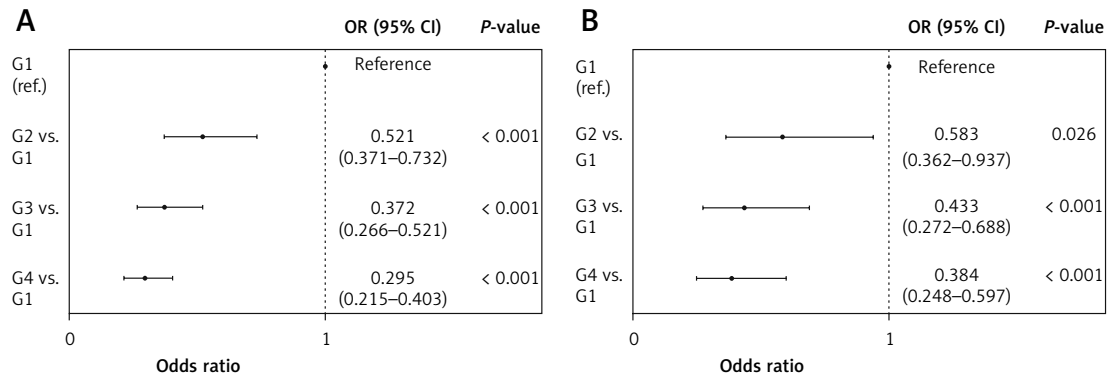


Figure 2. Univariate logistic regression analysis for subgroups. **A** – 28-day survival; **B** – 1-year survival. There was a statistically significant stepwise decrease in the risk of death at 28 days and 1 year from group 1 to group 4 (G1: LMR_{max} ≤ 1; G2: LMR_{max} > 1, ≤ 2; G3: LMR_{max} > 2, ≤ 3; G4: LMR_{max} > 3)

CI – confidence interval, G – group, LMR – lymphocyte-to-monocyte ratio, OR – odds ratio.

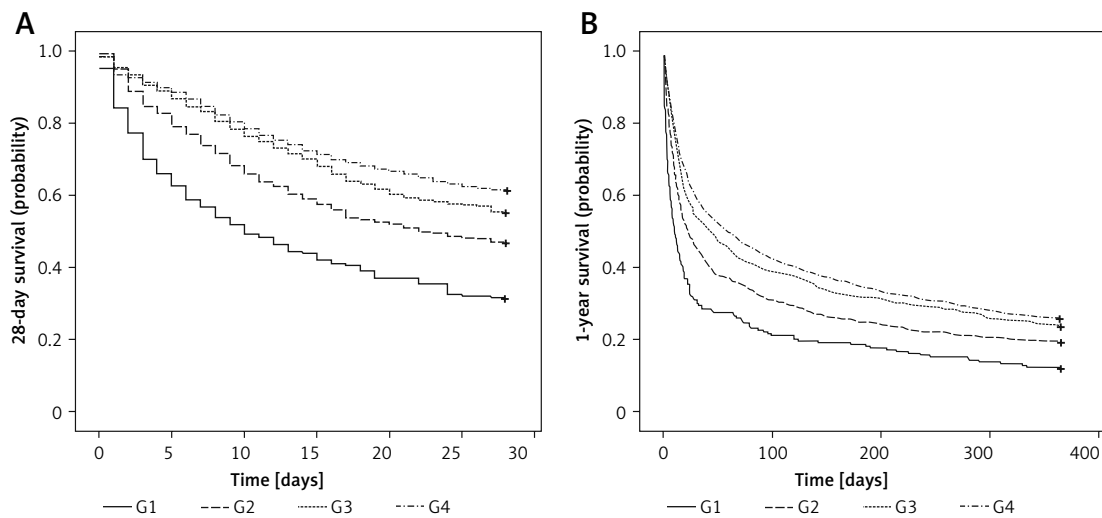


Figure 3. Kaplan-Meier (K-M) survival analysis for subgroups. **A** – 28-day survival curve; **B** – 1-year survival curve. The curves demonstrated that the patients of group 4 (maximum LMR_{max}) had the highest probability of survival at 28 days and 1 year, and the patients of group 1 (minimum LMR_{max}) had the lowest probability of survival at 28 days and 1 year (both $p < 0.001$)

G – group, LMR – lymphocyte-to-monocyte ratio.

Table III. Cox regression analysis for 28-day survival and 1-year survival

Research variable	28-day survival			1-year survival		
	HR	95% CI	P-value	HR	95% CI	P-value
Group 1: $LMR_{max} \leq 1$	Reference			Reference		
Group 2: $LMR_{max} > 1, \leq 2$	0.709	0.580–0.867	0.001	0.741	0.623–0.881	0.001
Group 3: $LMR_{max} > 2, \leq 3$	0.556	0.454–0.682	< 0.001	0.599	0.504–0.712	< 0.001
Group 4: $LMR_{max} > 3$	0.479	0.398–0.575	< 0.001	0.580	0.495–0.680	< 0.001

CI – confidence interval, ICU – intensive care unit, LMR – lymphocyte-to-monocyte ratio, LOS – length of stay, HR – hazard ratio, SAPS – Simplified Acute Physiology Score, SOFA – Sequential Organ Failure Assessment. The variables for multivariable adjustments: gender, age, lab items (hemoglobin and lactate), scores of SOFA and SAPS, state of alcohol abuse and comorbidities (congestive heart failure, cardiac arrhythmias, hypertension, chronic pulmonary, renal failure, liver disease, solid tumor and diabetes), and ICU LOS.

ter prognosis at 28 days and 1 year after hospital admission. The 28-day and 1-year survival rates increased significantly as the LMR_{max} increased. There was an independent and positive association between LMR_{max} and the 28-day and 1-year survival rates.

Sepsis, a leading cause of mortality worldwide, is defined as a syndrome of physiological, pathological, and biochemical abnormalities induced by infection [19–22]. The incidence of sepsis is increasing [23, 24]. Moreover, a considerable proportion of septic patients require treatment at the time of ICU admission [18]. Severe sepsis occurs when infection causes acute organ dysfunction, mediated by the inflammatory response [1, 2, 25]. Thus, it is appropriate to extract septic patients in the MIMIC III database using the criteria of Angus *et al.* [18].

Lymphocytes are involved in the pathophysiological inflammatory response of septic patients [26]. Sepsis-induced lymphocyte apoptosis, characterised by a decrease in quantity of lymphocytes and suppression of the immune response, is associated with a protracted course, infectious complications, and poor prognosis in septic patients [27–29]. Pro-inflammatory mediators produced by monocytes play a pivotal role in the pathogenesis of sepsis [30]. Based on their surface expression of CD14 and CD16, human monocytes can be subdivided into three subtypes: (1) classical monocytes, phagocytes with no inflammatory attributes; (2) non-classical monocytes, a subpopulation with inflammatory features; and (3) intermediate monocytes, a subpopulation with both phagocytic and inflammatory functions [31]. Therefore, it is confirmed that lymphocytes are a protective factor, while monocytes are a harmful factor in the process of inflammation.

Silva *et al.* reported that a lower LMR was independently associated with a higher risk of 6-month mortality after discharge among patients with acute heart failure [32]. Also, a lower LMR was independently associated with mortality in patients with hepatitis B virus-related liver cirrhosis [33],

malignant haematological disorders [34, 35], and peripheral arterial diseases [36]. Therefore, the LMR, which comprises the effects of both lymphocytes and monocytes, could be predictive of the prognosis of critically ill patients. In line with previous reports, we found that a lower LMR_{max} was associated with increased risk of short- and long-term mortality in septic patients, suggesting that the responses of lymphocytes and monocytes to inflammation, and the interaction between these two subtypes of leukocyte, are involved in the inflammatory cascade and subsequent recovery. The LMR_{max} value better reflects the continuous changes that occur during ICU admission. However, no study of the LMR_{max} has been reported, to our knowledge.

The LMR is determined from the counts of lymphocytes and monocytes, the activation, apoptosis, necrosis, and cytokine production of which are involved in the pathogenesis of sepsis [37]. Naess *et al.* demonstrated that, compared to patients with fever of non-infectious causes, patients with fever due to bacterial infection had a lower LMR, whereas those with viral infection had a higher LMR; this may be useful in the diagnosis of bacterial infection and selection of antibiotics [38]. Monocyte to lymphocyte ratios in the extreme percentiles (< 9% or > 25%) are reportedly associated with active tuberculosis [39]. Moreover, Djordjevic *et al.* suggested that the LMR value was not predictive of the outcomes of critically ill patients with peritonitis and pancreatitis, but was predictive of the severity of bacteraemia [40]. We found that the correlation between the LMR_{max} and survival was independent of sepsis-related scores; thus, an inflammation-related reaction may be involved in the recovery from sepsis.

The molecular mechanism underlying the role of LMR in survival is unclear. In septic lymphocytes, decreased expression of T-bet, GATA3, and ROR- γ t, which regulate Th1, Th2, and Th17 effector CD4 T cells, was observed [29, 41]. Furthermore, the mediation of microvesicular caspase-1 may induce apoptosis of lymphocytes after sepsis [42]. Also,

lower HLA-DR and fractalkine receptor (CX3CR1) expression on monocytes was correlated with a poor immunosuppression outcome in patients with clinical sepsis [43, 44]. Also, the number of CD16⁺ monocytes, which after stimulation produce the pro-inflammatory cytokine tumour necrosis factor but not anti-inflammatory factors, was markedly increased in response to sepsis [45, 46]. Moreover, Giamarellos-Bourboulis *et al.* reported that early apoptosis of blood monocytes was a mechanism of protection in sepsis, which in part supports our findings [47].

This study had several limitations. First, the retrospective design of the study renders it vulnerable to selection bias, where participants were drawn from only a single centre, patients with missing data were excluded and causality could not be determined. Second, we did not compare the effect of fungal and bacterial sepsis on prognosis. Third, we merely analysed the maximum value of LMR at ICU admission without dynamic observation, which perhaps affects the association between LMR and prognosis in septic patients. Thus, further prospective studies with continuous monitoring of the LMR are warranted to validate our findings.

Conclusions

We report for the first time that a lower LMR_{max} value was an independent predictor of a poor prognosis in septic patients, which provides clinical evidence regarding the mechanism underlying sepsis. Therefore, as an inexpensive and readily available indicator, LMR_{max} may facilitate stratification of prognosis in septic patients.

Acknowledgments

Xiang Hu and Xiaoyi Qin contributed equally to this work.

Conflict of interest

The authors declare no conflict of interest.

References

1. Singer M, Deutschman CS, Seymour CW, et al. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016; 315: 801-10.
2. Angus DC, van der Poll T. Severe sepsis and septic shock. *N Engl J Med* 2013; 369: 840-51.
3. Kaukonen KM, Bailey M, Suzuki S, Pilcher D, Bellomo R. Mortality related to severe sepsis and septic shock among critically ill patients in Australia and New Zealand, 2000-2012. *JAMA* 2014; 311: 1308-16.
4. Jedynak M, Siemiatkowski A, Milewski R, Mroczko B, Szmitkowski M. Diagnostic effectiveness of soluble triggering receptor expressed on myeloid cells-1 in sepsis, severe sepsis and septic shock. *Arch Med Sci* 2019; 15: 713-21.
5. Jain S, Sinha S, Sharma SK, et al. Procalcitonin as a prognostic marker for sepsis: a prospective observational study. *BMC Res Notes* 2014; 7: 458.
6. Będzichowska A, Przekora J, Stapińska-Syniec A, et al. Frequency of infections caused by ESBL-producing bacteria in a pediatric ward – single-center five-year observation. *Arch Med Sci* 2019; 15: 688-93.
7. Auriant I, Vinatier I, Thaler F, Tourneur M, Loirat P. Simplified acute physiology score II for measuring severity of illness in intermediate care units. *Crit Care Med* 1998; 26: 1368-71.
8. Ferreira FL, Bota DP, Bross A, Mélot C, Vincent JL. Serial evaluation of the SOFA score to predict outcome in critically ill patients. *JAMA* 2001; 286: 1754-8.
9. Pantzaris ND, Platanaki C, Pierrako C, Karamouzou V, Velissaris D. Neutrophil-to-lymphocyte ratio relation to sepsis severity scores and inflammatory biomarkers in patients with community-acquired pneumonia: a case series. *J Transl Int Med* 2018; 6: 43-6.
10. Orak M, Karakoç Y, Ustundag M, Yildirim Y, Celen MK, Güloğlu C. An investigation of the effects of the mean platelet volume, platelet distribution width, platelet/lymphocyte ratio, and platelet counts on mortality in patients with sepsis who applied to the emergency department. *Niger J Clin Pract* 2018; 21: 667-71.
11. Saliccioli JD, Marshall DC, Pimentel MA, et al. The association between the neutrophil-to-lymphocyte ratio and mortality in critical illness: an observational cohort study. *Crit Care* 2015; 19: 13.
12. Biyikli E, Kayipmaz AE, Kavalci C. Effect of platelet-lymphocyte ratio and lactate levels obtained on mortality with sepsis and septic shock. *Am J Emerg Med* 2018; 36: 647-50.
13. Terradas R, Grau S, Blanch J, et al. Eosinophil count and neutrophil-lymphocyte count ratio as prognostic markers in patients with bacteremia: a retrospective cohort study. *PLoS One* 2012; 7: e42860.
14. Ren H, Han L, Liu H, Wang L, Liu X, Gao Y. Decreased lymphocyte-to-monocyte ratio predicts poor prognosis of acute ischemic stroke treated with thrombolysis. *Med Sci Monit* 2017; 23: 5826-33.
15. Ren H, Liu X, Wang L, Gao Y. Lymphocyte-to-monocyte ratio: a novel predictor of the prognosis of acute ischemic stroke. *J Stroke Cerebrovasc Dis* 2017; 26: 2595-602.
16. Kim JY, Kawabori M, Yenari MA. Innate inflammatory responses in stroke: mechanisms and potential therapeutic targets. *Curr Med Chem* 2014; 21: 2076-97.
17. Johnson AE, Pollard TJ, Shen L, et al. MIMIC-III, a freely accessible critical care database. *Sci Data* 2016; 3: 160035.
18. Angus DC, Linde-Zwirble WT, Lidicker J, Clermont G, Carcillo J, Pinsky MR. Epidemiology of severe sepsis in the United States: analysis of incidence, outcome, and associated costs of care. *Crit Care Med* 2001; 29: 1303-10.
19. Vincent JL, Marshall JC, Namendys-Silva SA, et al. Assessment of the worldwide burden of critical illness: the intensive care over nations (ICON) audit. *Lancet Respir Med* 2014; 2: 380-6.
20. Fleischmann C, Scherag A, Adhikari NK, et al. International forum of acute care trialists: assessment of global incidence and mortality of hospital-treated sepsis. current estimates and limitations. *Am J Respir Crit Care Med* 2016; 193: 259-72.
21. Soylu L, Aydin OU, Atli M, et al. Does early removal of double J stents reduce urinary infection in living donor renal transplantation?. *Arch Med Sci* 2019; 15: 402-7.

22. Sahin M. The role of topical Genta Fleece HD and gentamicin spray in prevention of sternum wound infections after open heart surgery: a comparative study. *Arch Med Sci Atheroscler Dis* 2018; 3: e29-e34.
23. Iwashyna TJ, Cooke CR, Wunsch H, Kahn JM. Population burden of long-term survivorship after severe sepsis in older Americans. *J Am Geriatr Soc* 2012; 60: 1070-7.
24. Gaieski DF, Edwards JM, Kallan MJ, Carr BG. Benchmarking the incidence and mortality of severe sepsis in the United States. *Crit Care Med* 2013; 41: 1167-74.
25. Fan H, Zhao Y, Sun M, Zhu JH. Urinary neutrophil gelatinase-associated lipocalin, kidney injury molecule-1, N-acetyl-beta-D-glucosaminidase levels and mortality risk in septic patients with acute kidney injury. *Arch Med Sci* 2018; 14: 1381-6.
26. Monneret G, Venet F. A rapidly progressing lymphocyte exhaustion after severe sepsis. *Crit Care* 2012; 16: 140.
27. Castellino DJ, McNair P, Kay TW. Lymphocytopenia in a hospital population: what does it signify? *Aust N Z J Med* 1997; 27: 170-4.
28. Abraham E. Physiologic stress and cellular ischemia: relationship to immunosuppression and susceptibility to sepsis. *Crit Care Med* 1991; 19: 613-8.
29. Cabrera-Perez J, Condotta SA, Badovinac VP, Griffith TS. Impact of sepsis on CD4 T cell immunity. *J Leukoc Biol* 2014; 96: 767-77.
30. Tsaganos T, Giamarellos-Bourboulis EJ, Kollias S, et al. Kinetics of progenitor hemopoietic stem cells in sepsis: correlation with patients survival? *BMC Infect Dis* 2006; 6: 142.
31. Ziegler-Heitbrock L, Ancuta P, Crowe S, et al. Nomenclature of monocytes and dendritic cells in blood. *Blood* 2010; 116: e74-80.
32. Silva N, Bettencourt P, Guimarães JT. The lymphocyte-to-monocyte ratio: an added value for death prediction in heart failure. *Nutr Metab Cardiovasc Dis* 2015; 25: 1033-40.
33. Zhang J, Feng G, Zhao Y, Zhang J, Feng L, Yang J. Association between lymphocyte-to-monocyte ratio (LMR) and the mortality of HBV-related liver cirrhosis: a retrospective cohort study. *BMJ Open* 2015; 5: e008033.
34. Porrata LF, Ristow K, Colgan JP, et al. Peripheral blood lymphocyte/monocyte ratio at diagnosis and survival in classical Hodgkin's lymphoma. *Haematologica* 2012; 97: 262-9.
35. Watanabe R, Tomita N, Itabashi M, et al. Peripheral blood absolute lymphocyte/monocyte ratio as a useful prognostic factor in diffuse large B-cell lymphoma in the rituximab era. *Eur J Haematol* 2014; 92: 204-10.
36. Gary T, Pichler M, Belaj K, et al. Lymphocyte-to-monocyte ratio: a novel marker for critical limb ischemia in PAOD patients. *Int J Clin Pract* 2014; 68: 1483-7.
37. Stearns-Kurosawa DJ, Osuchowski MF, Valentine C, Kurosawa S, Remick DG. The pathogenesis of sepsis. *Annu Rev Pathol* 2011; 6: 19-48.
38. Naess A, Nilssen SS, Mo R, Eide GE, Sjursen H. Role of neutrophil to lymphocyte and monocyte to lymphocyte ratios in the diagnosis of bacterial infection in patients with fever. *Infection* 2017; 45: 299-307.
39. Wang J, Yin Y, Wang X, et al. Ratio of monocytes to lymphocytes in peripheral blood in patients diagnosed with active tuberculosis. *Braz J Infect Dis* 2015; 19: 125-31.
40. Djordjevic D, Rondovic G, Surbatovic M, et al. Neutrophil-to-lymphocyte ratio, monocyte-to-lymphocyte ratio, platelet-to-lymphocyte ratio, and mean platelet volume-to-platelet count ratio as biomarkers in critically ill and injured patients: which ratio to choose to predict outcome and nature of bacteremia? *Mediators Inflamm* 2018; 2018: 3758068.
41. Pachot A, Monneret G, Voirin N, et al. Longitudinal study of cytokine and immune transcription factor mRNA expression in septic shock. *Clin Immunol* 2005; 114: 61-9.
42. Exline MC, Justiniano S, Hollyfield JL, et al. Microvesicular caspase-1 mediates lymphocyte apoptosis in sepsis. *PLoS One* 2014; 9: e90968.
43. Flohé S, Scholz M. HLA-DR monitoring in the intensive care unit: more than a tool for the scientist in the laboratory? *Crit Care Med* 2009; 37: 2849-50.
44. Pachot A, Cazalis MA, Venet F, et al. Decreased expression of the fractalkine receptor CX3CR1 on circulating monocytes as new feature of sepsis-induced immunosuppression. *J Immunol* 2008; 180: 6421-9.
45. Munn DH, Bree AG, Beall AC, et al. Recombinant human macrophage colony-stimulating factor in nonhuman primates: selective expansion of a CD16+ monocyte subset with phenotypic similarity to primate natural killer cells. *Blood* 1996; 88: 1215-24.
46. Frankenberger M, Sternsdorf T, Pechumer H, Pforte A, Ziegler-Heitbrock HW. Differential cytokine expression in human blood monocyte subpopulations: a polymerase chain reaction analysis. *Blood* 1996; 87: 373-7.
47. Giamarellos-Bourboulis EJ, Routsis C, Plachouras D, et al. Early apoptosis of blood monocytes in the septic host: is it a mechanism of protection in the event of septic shock? *Crit Care* 2006; 10: R76.