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Type
Research paper

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Material and methods
The levels of NEAT1 and microRNA-320b (miR-320b) were measured by qRT-PCR. Western blot was carried out to determine the protein levels that used in this research. Cell viability was identified via Cell Counting Kit-8 (CCK-8). Transwell assay was employed to determine migration and invasion. The relationship between miR-320b and NEAT1 or MSI2 was clarified by dual-luciferase reporter assay, RNA immunoprecipitation (RIP) and RNA pull down assay. Also, a murine xenograft assay was used to explore the effect of NEAT1 on cisplatin resistance in OC in vivo.

Results
The level of NEAT1 was significantly increased in cisplatin resistant OC cell lines. Downregulation of NEAT1 enhanced cisplatin sensibility in OVCAR-3/DDP and HEY/DDP cells. Furthermore, miR-320b was a target of NEAT1, and the effects of knockdown of NEAT1 on the cell viability, IC50 of cisplatin, migration and invasion in OVCAR-3/DDP and HEY/DDP were restored by the inhibitor of miR-320. In addition, miR-320b directly targeted MSI2 to regulate cisplatin sensibility in cisplatin resistant OC cells. In addition, downregulation of NEAT1 decreased cisplatin resistance in OC in vivo.

Conclusions
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Long non-coding RNA NEAT1/miR-320b/MSI2 axis regulates cisplatin resistance in ovarian cancer

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**Introduction**

Ovarian cancer (OC) is a common cause of cancer-related death in women. It frequently occurs in postmenopausal women, along with several months of abdominal pain and distension. Most patients come under observation have been in International Federation of Gynecology and Obstetrics [FIGO] stage III [1], usually therapy by using surgery and systemic treatment including chemotherapy, antiangiogenic drugs and PARP inhibitors and in selected cases radiotherapy. However, because of lack of effective chemotherapy strategy, the mortality of OC patients is still high, accompanied by a cure rate of only 30% [2]. Exploring effective anti-cancer drugs or enhance chemosensitivity is of great significance. Oridonin was reported to repress OC cell metastasis [3], but whether it could be used for clinical treatment remains uncertain.

Cisplatin (DDP) is a common and effective chemotherapy drug for the therapy of OC [4]. Thus this study focused on the regulatory mechanism of cisplatin (DDP) resistance.

Over the past decades, studies indicated that chemoresistance was influenced by multiple factors, including long non-coding RNAs (lncRNAs) [5]. LncRNA is a kind of transcript, which has no function of translating into protein, but involved in the pathogenesis of various diseases, including but not limited to tumors. Accumulating researches proved that lncRNA modified cancer-associated processes, like metastasis and loss of imprinting [6]. Specially, upregulation of lncRNA anti-differentiation non-
coding RNA (DANCR) involved in the advanced progression of tumor and poor prognosis in colorectal cancer [7], downregulation of it enhanced osteoblast differentiation by regulating enhancer of zeste homolog 2 (EZH2) [8]. What’s more, IncRNAs was associated with the function and regulatory mechanism of OC [9]. Such as metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) induced epithelial ovarian cancer cell proliferation and metastasis by phosphatidylinositol 3-kinase/ protein kinase B (PI3K/AKT) signaling pathway [10]. Prostate cancer-associated transcript-1 (PCAT-1) was upregulated in OC and facilitated OC cell progression [11]. Myocardial infarction-associated transcript MIAT negative regulated miR-330-5p and acted as a tumor oncogenic IncRNA in epithelial ovarian cancer tumorigenesis [12]. Moreover, nuclear paraspeckle assembly transcript 1 (NEAT1) facilitated paclitaxel resistance in OC cells through microRNA-194/zincfinger ebox binding homeobox 1 (miR-194/ZEB1) axis [13]. All evidence suggested that NEAT1 participated in the progression and chemoresistance of OC, further researches for the molecular mechanism of the process needed to be conducted.

MicroRNAs (miRNAs) have been discovered to involve in the initiation and progression of tumors [14]. MicroRNA research in the context of tumor is a new strategy for pharmacologic approaches [15]. For example, microRNA-200, which was a family of tumor repressor miRNAs was strongly associated with inhibition of epithelial mesenchymal transformation, caused the inhibiting effect of cancer stem cells [16]. MicroRNA-179 targeted to PTEN-mediated PI3K/AKT signaling pathway to regulate proliferation and chemosensitivity of human ovarian cancer cells [17].
MicroRNA-331-3p reduced regulator of chromosome condensation 2 (RCC2) expression to impede OC cell proliferation and metastasis [18]. Previous research also shown that microRNA-320 acted as a hypoxia-regulated microRNA, was downregulated in hypoxic cells and regulated the expression of VEGF [15]. Meanwhile, miRNA also modulated the chemoresistance in human cancer [19]. MicroRNA-214 promoted cell survival and cisplatin resistance in OC by targeting phosphatase and tensin homolog deleted on chromosome ten (PTEN) [20]. MicroRNA-320b (miR-320b) as a target of X-inactive specific transcript (XIST), regulated the progression of osteosarcoma [21]. However, whether miR-320b modulates cisplatin resistance of human OC is unknown.

Musashi-2 (MSI2) has been proved to regulate the progression in amounts of human diseases [22]. For example, MSI2 contributed to cell metastasis by supporting transforming growth factor-β (TGF-β) [23]. MSI2 was a predictive biomarker for liver metastasis and was related to poor prognosis in colorectal cancer [24]. Moreover, MSI2 induced the chemoresistance of liver cancer stem cells by regulating the activation of lin-28 homolog A (LIN28A) [25]. All evidence revealed that MSI2 may participate in chemoresistance in the therapy of human OC.

Herein, the research paid attention to the regulatory mechanism of NEAT1, miR-320b and MSI2 in OC chemoresistance, and uncovered that NEAT1/miR-320b/MSI2 axis regulated the chemoresistance of OC in vivo and in vitro.

**Materials and methods**
Cell culture

Firstly, human OC cell lines OVCAR-3 and HEY were obtained from Chuan Qiu Biotechnology (Shanghai, China) and cultured in Roswell Park Memorial Institute (RPMI) medium. The cisplatin-resistant cell lines OVCAR-3/DDP and HEY/DDP were generated from their parental cell lines OVCAR-3 and HEY through treating with increasing concentration of cisplatin. All mediums were mixed with 10% fetal bovine serum (FBS, Gibco, Carlsbad, CA, USA) and 100 U/mL penicillin&streptomycin (Gibco), in addition, OVCAR-3/DDP and HEY/DDP cells were added cisplatin (APEX BIO Technology, Austin, TX, USA) with a final concentration of 2 ug/mL. OC cells were incubated in 37°C with 5% CO₂.

Cell Counting Kit-8 (CCK-8) assay

CCK-8 assay was carried out for detection of cell viability and IC₅₀ of cisplatin. Firstly, transfected cells were seeded in 96-well plates (Thermo Fisher Scientific, Waltham, MA, USA), and incubated for 48 h, three repetitions were prepared for per sample. Subsequently, WST-8 (Sigma-Aldrich, Louis, MO, USA) was added into cells which was added different concentrations of cisplatin and cultured for another 2 h. Finally, microplate reader (Bio-Rad Laboratories, Philadelphia, PA, USA) was performed to measure OD value at 450 nm. The cisplatin concentration repressed cell viability of 50% was considered cisplatin IC₅₀ in vitro.

Transwell assay

Cell migration and invasion was determined by transwell assay. The upper chamber was coated without or with Matrigel (BD Biosciences, San Jose, CA, USA) for
migration or invasion detection, respectively. Then, serum-free medium containing
cells was added (1 × 10^4 cells for migration and 2 × 10^4 cells for invasion). Otherwise,
the basolateral chamber was added cell medium with 10% serum. After incubation for
48 h, the cells throughout the membrane was fixed by 4% paraformaldehyde and dyed
with 0.5% crystal violet solution. The cells were counted under a microscope (Thermo Fisher Scientific).

Western blot assay

OC cells were lysed by using Radio Immunoprecipitation Assay (RIPA) lysis buffer
(Millipore, Bedford, MA, USA), and Extraction Buffer (Millipore) contained with
protease inhibitors (Thermo Fisher Scientific) were applied to extract protein. Then
protein concentration was examined by BCA Protein Assay Kit (Sangon Biotech, Shanghai, China). Subsequently, quantified protein which was denatured by boiling
water was separated on SDS-polyacrylamide gel via electrophoresis and blotted onto
polyvinylidene fluoride (PVDF) membranes (Millipore). Then membranes were
blocked with 5% (w/v) bovine serum albumin (Solarbio, Beijing, China) for 2 h at room
temperature. Whereafter, PVDF membranes were incubated with primary antibodies
which purchased from Abcam (Cambridge, MA, USA), including anti-Matrix
metalloproteinases (MMP) 2(ab97779; 1:1000), anti-MMP9 (ab38898; 1:1000), anti-
MIS2 (ab73164; 1:1000) and anti-glyceraldehyde 3-phosphate dehydrogenase
(GAPDH) (ab181602; 1:5000) overnight at 4°C. Next, the membranes were incubated
for 1 h at room temperature with secondary antibody (ab205718; 1:5000). Finally,
protein signals were visualized utilizing Pierce™ ECL Western Blotting Substrate
Quantitative real-time polymerase chain reaction (qRT-PCR)

The total RNA were extracted by Trizol (Invitrogen), and RNA was reverse transcribed into cDNA by using Thermo Scientific RevertAid First Strand cDNA Synthesis kit (Thermo Fisher Scientific). Then qRT-PCR was carried out to examine the expression of special RNA via SYBR Green real-time PCR kit (Takara, Dalian, China). In the end point, solution was reacted and signals were collected via ABI 7500 fast system (Applied Biosystems, Rockford, IL, USA). The levels of NEAT1, miR-320b and MSI2 were quantified via $2^{-\Delta\Delta Ct}$ method and normalized by GAPDH (for NEAT1 and MSI2) and U6 (for miR-320b). Special primers were listed as below: NEAT1: (forward 5’-CTT CCT CCC TTT AAC TTA TCC ATT CAC-3’, reverse 5’-CTC TTC CTC CAC CAT TAC CAA CAA TAC-3’); MSI2: forward 5’-( and GAPDH: (forward 5’- ATT CCA TGG CAC CGT CAA GGC TGA -3’, reverse 5’- TTC TCC ATG GTG GTG AAG ACG CCA -3’), which were synthesized in Ribobio, and The forward and reverse primers for miR-320b and U6 were purchased from Ribobio. The gene expression was presented using $2^{-\Delta\Delta Ct}$ method.

Transient transfection

Special sequences, including Small interfering RNA (siRNA) against NEAT1 and siRNA negative control (si-control) miR-320 mimics (miR-320b), mimics blank control (miR-control), miR-320 inhibitor (miR-320 inhibitor), inhibitor negative control (inhibitor-control), overexpression vector of MSI2 (MSI2) and overexpression empty vector (pcDNA), bio-labeled probe of miR-320b (Bio-miR-320b), blank control
(Bio-NC) and input (NC-Input, miR-320b-Input) as well as vectors of binding sites of wildtype (NEAT1 WT) and mutant (NEAT1 MUT) of NEAT1 and miR-320b, and 3′-UTR sequences of wildtype (MSI2 3′UTR WT) and mutant (MSI2 3′UTR MUT) of MSI2 contained common fragments with miR-320b, short hairpin RNA (shRNA) of NEAT1 (sh-NEAT1) and shRNA negative control (sh-control) were synthesized in Ribobio (Guangzhou, China). Subsequently, vectors and oligonucleotide were transfected into OC cell lines by using Lipofectamine™ 2000 (Invitrogen, Carlsbad, CA, USA) according to producer’s manual, respectively.

**Dual-luciferase reporter assay**

LncBase Predicted v.2 was performed to predict the relationship between NEAT1 and miR-320b, and the interrelation between miR-320b and MSI2 was predicted utilizing Targetscan. Then vectors of binding sites of wildtype (NEAT1 WT) and mutant (NEAT1 MUT) of NEAT1 and miR-320b, and 3′-UTR sequences of wildtype (MSI2 3′UTR WT) and mutant (MSI2 3′UTR MUT) of MSI2 containing common fragments with miR-320b were synthesized in Ribobio. OVCAR-3/DDP and HEY/DDP cells were transfected with one of them and renilla plasmid, severally. Finally, luciferase activity was examined by Dual-Luciferase® Reporter Assay System (Promega, Madison, WI, USA) according to manual and analyzed via Varioskan Flash (Thermo Fisher Scientific).

**RNA immunoprecipitation (RIP) assay**

The assay was used EZ-Magna RIP™ RNA-Binding Protein Immunoprecipitation Kit (Millipore) to verify the relationship between NEAT1 and miR-320b. Firstly, OVCAR-3/DDP and HEY/DDP cells were incubated for 48 h and lysed by using RIP-buffer.
Then lysates were incubated with magnetic beads coated AGO2 (Argonaute-2) or IgG antibody which obtained from Abcam. Finally, residuum were washed off and RNA were harvested, qRT-PCR was carried to detect the enrichment of NEAT1 in OVCAR-3/DDP and HEY/DDP cells.

**RNA pull down assay**

In the assay, bio-labeled probe of miR-320b (Bio-miR-320b), blank control (Bio-NC) and input (NC-Input, miR-320b-Input) were synthesized in Ribobio. Subsequently, OVCAR-3/DDP and HEY/DDP cells were transfected with probes, respectively, and incubated for 48 h. Then, cells were lysed and lysates were incubated with Streptavidin-Dyna beads overnight at 4°C accompanied by RNA separation. Then the enrichment of NEAT1 was measured by qRT-PCR *in vitro.*

**Murine xenograft assay**

BALB/c nude mice (male, four-week-old) were purchased from Shanghai SLAC Laboratory Animal Co, Ltd. (Shanghai, China) and divided in three group (n = 6 per group) randomly. The assay was performed according to guidelines of the National Animal Care and Ethics Institution and approved by Animal Research Committee of Eye, Ear, Nose and Larynx Hospital, Liaocheng People's Hospital. OVCAR-3/DDP cells were transfected with the lentivirus harboring sh-NEAT1 or negative control (sh-control) constructed by Ribobio. Subsequently, stably transfected cells and OVCAR-3/DDP cells were infected into mice subcutaneously, and then the mice were treated with DDP. The tumor volumes were examined after injected one week and then measured every three days until the mice were sacrificed at the 22 d post injection. The
volumes were calculated following the formula: volume (mm$^3$) = width$^2 \times$ length/2. Tumor samples were also taken out and weighted after mice were sacrificed.

**Statistical analysis**

All the data were put forward as means ± standard deviation (means ± SD) for three independent assays, Student’s $t$ test was carried out to examine the difference between two groups. Additionally, one-way analysis of variance (ANOVA) was applied for multiple groups. Differences were considered significant statistically at $P$ less than 0.05.

**Results**

**The level of NEAT1 was notably increased in cisplatin-resistant OC cell lines**

Firstly, cisplatin-resistant cell lines were established. The cell viability, $IC_{50}$ of cisplatin, migration and invasion of the parental OC cells and cisplatin-resistant OC cells were detected to assess the cisplatin resistance. CCK-8 assay revealed that the cell viability and $IC_{50}$ of cisplatin of OVCAR-3/DDP and HEY/DDP cells were higher than that of OVCAR-3 and HEY cells (Fig. 1A and 1B). Moreover, the capacities of migration and invasion of OVCAR-3/DDP ($P = 0.0002$, $P < 0.0001$) and HEY/DDP cells ($P = 0.0004$, $P < 0.0001$) were both enhanced compared with their parental cells (Fig. 1C and 1D). Besides, the protein levels of migration-related proteins (MMP2 and MMP9) were also measured, and the results showed that MMP2 and MMP9 levels were significantly elevated in OVCAR-3/DDP and HEY/DDP cells ($P < 0.0001$) (Fig. 1E and 1F), which was consistent with the transwell result. These results indicated that cisplatin-resistant
OC cell lines were successfully constructed. Then the level of NEAT1 were detected. The qRT-PCR result manifested that NEAT1 level was strikingly increased in cisplatin resistant OC cells (OVCAR-3/DDP and HEY/DDP cells) \( (P < 0.0001) \) (Fig. 1G and 1H). The aberrant expression of NEAT1 suggested that it might be involved in the regulation of cisplatin resistance in human OC.

Knockdown of NEAT1 enhanced cisplatin sensitivity of cisplatin resistance OC cell lines.

In order to investigate the potential role of NEAT1 in the regulatory mechanism of OC cisplatin resistance, si-NEAT1 or si-control was transfected into OVCAR-3/DDP and HEY/DDP cells, and the level of NEAT1 was significantly decreased in cisplatin resistant OC cells transfected with si-NEAT1 \( (P = 0.0013, P = 0.0002) \) (Fig. 2A and 2B). Cell viability and cisplatin IC50 were notably reduced by si-NEAT1 in OVCAR-3/DDP and HEY/DDP cells (Fig. 2C and 2D). Moreover, migration and invasion of OVCAR-3/DDP \( (P = 0.0001, P < 0.0001) \) and HEY/DDP cells \( (P < 0.0002, P < 0.0001) \) were also inhibited by NEAT1 knockdown (Fig. 2E and 2F). Furthermore, knockdown of NEAT1 downregulated the expression of MMP2 and MMP9 \textit{in vitro} \( (P < 0.0001) \) (Fig. 2G and 2H). From the above, knockdown of NEAT1 increased cisplatin sensitivity of OVCAR-3/DDP and HEY/DDP cells.

MiR-320b was a target gene of NEAT1.

The binding sites between NEAT1 and miR-320b were predicted by LncBase Predicted
v.2 (Fig. 3A). Dual-luciferase reporter assay was performed to clarify the interrelation between them, luciferase activity was evidently decreased in NEAT1 WT group ($P < 0.0001$), while had no significant difference in NEAT1 MUT group ($P = 0.6884$, $P = 0.9922$), the evidence suggested that miR-320b was a target of NEAT1 (Fig. 3B and 3C). Moreover, the results of RIP ($P < 0.0001$) and RNA pull down assay ($P = 0.0005$, $P = 0.0001$) were both verified the above conclusion (Fig. 3D-3G). In addition, the level of miR-320b was enhanced by sh-NEAT1 whereas repressed via overexpression of NEAT1 in OVCAR-3/DDP and HEY/DDP cells. These evidences revealed that NEAT1 directly targeted miR-320b, it might regulate OC cisplatin resistance by sponging miR-320b.

The effect of knockdown of NEAT1 on cisplatin sensitivity was reversed by miR-320b inhibitor in cisplatin-resistant OC cell lines

In order to research regulatory mechanism between NEAT1 and miR-320b, si-control, si-NEAT1, si-NEAT1+miR-control or si-NEAT1+miR-320b was transfected into OVCAR-3/DDP and HEY/DDP cells, severally. The effect of si-NEAT1 on the expression of miR-320b was evidently reversed by miR-320b inhibitor ($P < 0.0001$) (Fig. 4A and 4B). The cell viability and $IC_{50}$ of cisplatin which decreased by si-NEAT1 was regained by knockdown of miR-320b in SKOV3/DDP and A2780/DDP cells (Fig. 4C and 4D). What’s more, si-NEAT1 repressed migration and invasion which was rescued by miR-320b inhibitor ($P < 0.0001$) (Fig. 4E and F). In addition, the downregulated MMP2 and MMP9 expression resulted by NEAT1 knockdown were
restored by miR-320b inhibitor ($P < 0.0001$) (Fig. 4G and 4H). All evidences proved that the effect of NEAT1 knockdown on cisplatin sensitivity was reversed by miR-320b inhibitor in OC in vitro.

MiR-320b directly targeted MSI2

The interrelation between miR-320b and MSI2 was predicted via Targetscan and the binding sites were exhibited (Fig. 5A). Luciferase activity was significantly repressed by MIS2 3’UTR WT in OVCAR-3/DDP and HEY-3/DDP cell lines ($P < 0.0001$), while no notably change was observed in MSI2 3’UTR MUT group ($P = 0.9706$, $P = 0.9846$) (Fig. 5B and 5C). Besides, the expression of MSI2 was distinctly decreased by miR-320b mimics ($P < 0.0001$) (Fig. 5D). Knockdown of NEAT1 downregulated MSI2 expression, and this effect was reversed by miR-320b inhibitor (Fig. 5E and 5F).

Additionally, the expression of MSI2 was markedly increased in OVCAR-3/DDP and HEY/DDP cells relative to VCAR-3 and HEY cells ($P < 0.0001$) (Fig. 5G and 5H). In short, MSI2 was a target of miR-320b, and NEAT1 regulated MSI2 expression through sponging miR-320b.

Overexpression of MSI2 restored the effect of miR-320b on cisplatin sensitivity in cisplatin-resistant OC cell lines

To further investigate the molecular mechanism between miR-320b and MSI2, OVCAR-3/DDP and HEY/DDP cells were transfected with miR-control, miR-320b, miR-320b+pcDNA or miR-320b+pcDNA-MSI2, respectively. The effect of miR-320b
on MSI2 expression was rescued by pcDNA-MSI2 (Fig. 6A and 6B). In addition, the cell viability and cisplatin IC$_{50}$ was decreased by miR-320b while regained by overexpression of MSI2 in OVCAR-3/DDP and HEY/DDP cells (Fig. 6C and 6D). The effects of miR-320b mimics on migration and invasion were restored via co-transfecting with pcDNA-MSI2 in OVCAR-3/DDP and HEY/DDP cells (Fig. 6E and 6F). Moreover, overexpression of MSI2 recovered the effect of miR-320b mimics on MMP2 and MMP9 expression in cisplatin resistant OC cells (Fig. 6G and 6H). In brief, overexpression of MSI2 reversed the effect of miR-320b mimics on cell viability, cisplatin IC$_{50}$, migration and invasion in cisplatin resistant OC cell lines, which meant that the effect of miR-320b mimics on increasing cisplatin sensibility was recovered by pcDNA-MSI2 in vitro.

**Knockdown of NEAT1 retarded the progression of OC tumors after cured with DDP or not in vivo**

Nude mice were injected with OVCAR-3/DDP cells transfected with sh-control or sh-NEAT1 followed by treating with DDP. The tumor volumes and weight were both dramatically decreased in sh-NEAT1+DDP and sh-control+DDP groups, especially in sh-NEAT1+DDP group ($P < 0.0001$, $P = 0.0134$) (Fig. 7A-7B). Meanwhile, the expression level of NEAT1 was significantly curbed while miR-320b expression was markedly enhanced via treatment with sh-NEAT1+DDP ($P = 0.0003$, $P < 0.0001$) (Fig. 7C-7D). Finally, MSI2 expression was obviously decreased in sh-NEAT1+DDP group ($P = 0.0001$) (Fig. 7E). In conclusion, tumor progression was blocked via knockdown
Discussion

OC is a gynecologic malignancy which causing highly fatality ratio, and according to statistics, about 70% of patients will eventually succumb to their disease [26]. A large number of OC patients, with the exception of some well-differentiated early-stage cancer patients, undergo the treatment of chemotherapy with platinum (cisplatin or carboplatin) and paclitaxel [27]. The therapy shows promising effects at first, however, the occurrence and development of chemotherapy chemoresistance impose restrictions on successful treatment outcomes [28]. Therefore, chemoresistance is the primary problem in the treatment of ovarian cancer. The study used OVCAR-3/DDP and HEY/DDP cells as cisplatin resistant OC cell lines, and investigated the molecular mechanism of cisplatin resistance in OC.

Accumulating evidences have revealed that IncRNA, whose length over than 200 nucleotides, participated in the tumorigenesis of multiple cancers [29-31]. Specially, urothelial cancer associated 1 (UCA1) promoted breast cancer epithelial-mesenchymal transition by inducing Wnt/beta-catenin pathway [32]. Furthermore, IncRNA was also referred to human OC [33]. Human ovarian cancer-specific transcript 2 (HOST2) modulated cell biological behaviors by sponging microRNA let-7b in epithelial ovarian cancer [34]. NEAT1 accelerated metastasis of ovarian cancer via upregulating ROCK1 through miR-382-3p [35]. Also, NEAT1 could be regulated by p53 and was closely related with chemoresistance [36]. Previous studies revealed that NEAT1 enhanced
cisplatin resistance in bladder cancer, nasopharyngeal carcinoma and anaplastic thyroid carcinoma [37-39]. On the contrary, NEAT1 reduced cisplatin resistance in lung cancer [40]. Besides, NEAT1 expression was consistent with the reactive oxygen species (ROS) level in cancer cells [41,42]. The levels of ROS in drug resistant cancer cells are typically increased compared to non-multidrug resistance (MDR) cancer and normal cells [43]. Mounting evidence suggest that modulating cellular ROS levels can enhance MDR cancer cell death and sensitize MDR cancer cells to certain chemotherapeutic drugs [44,45]. These researches revealed that NEAT1 played different roles on cisplatin resistance in different cancers. Thus we explored the effect of NEAT1 on cisplatin resistance in OC. Our data uncovered that knockdown of NEAT1 augmented cisplatin sensibility in cisplatin resistant OC cells.

NEAT1 may exert its role through regulating downstream gene in the progress. For example, NEAT1 has been reported to regulate chemoresistance via miR-194/ZEB1, let-7a-5p/Rsf-1 and miR-9-5p/SPAG9 axis [10, 38, 39]. We also found that miR-194, let-7a-5p and miR-9-5p were upregulated, while ZEB1, Rsf-1 and SPAG9 were downregulated in cisplatin resistant OC cells (Fig. S1), suggesting that NEAT1 may response to cisplatin resistant through various pathways in OC cells. However, it still needs to be verified by further studies. In this study, we identified miR-320b as a target of NEAT1. A previous report proved that miR-320b was significantly downregulated in OC and might serve as biomarker for the prognosis of OC [45]. Yet there was no researches reported the effect of miR-320b on cisplatin resistance. Then this study showed that miR-320b was downregulated in cisplatin resistant OC cells, and miR-
320b inhibitor reversed the effect of knockdown of NEAT1 on enhancing cisplatin sensibility in OC cells, implying the inhibitory role of miR-320b in cisplatin resistance. Additionally, further researches were also needed to investigate the regulatory mechanism of miR-320b in chemoresistance in cancers.

MSI2 has been suggested to participate in human haematopoietic stem cells via repressing aryl hydrocarbon receptor (AHR) signaling during the past decades [46]. It also contributed to cell migration and invasion of bladder cancer via activating Janus kinase 2/ signal transducer and activation of transcription 3 (JAK2/STAT3) pathway [47]. What’s more, MSI2 was a novel regulator to modify paclitaxel sensitivity in OC cell lines [48]. In the research, MSI2 was a target by miR-320b, overexpression of it fortified cisplatin resistance in OVCAR-3/DDP and HEY/DDP cells. MSI2 may function as an oncogene, involve in chemoresistance in OC patients.

In conclusion, the level of NEAT1 was upregulated in cisplatin-resistant OC cells, subsequent assay showed that it regulated cisplatin resistance by NEAT1/miR-320b/MSI2 axis. However, the chemotherapy resistance of various tumors still needs to be further studied in the future.

**Conclusion**

The level of NEAT1 was notably increased in cisplatin resistant OC cell lines. Knockdown of NEAT1 significantly augmented the cisplatin sensibility in OVCAR-3/DDP and HEY/DDP cells. What’s more, miR-320b as a target gene of NEAT1, miR-320b inhibitor reversed the effect of knockdown of NEAT1 on the cell viability, IC$_{50}$ of cisplatin, migration and invasion in OVCAR-3/DDP and HEY/DDP cells. In addition,
MSI2 was directly targeted by miR-320b. MiR-320b mimics significantly decreased cell viability, cisplatin IC$_{50}$ migration and invasion in OVCAR-3/DDP and HEY/DDP cells, while these effects were rescued by MSI2 overexpression vector. Furthermore, downregulation of NEAT1 curbed the progression of tumor in vivo. In brief, NEAT1 regulated cisplatin resistant through NEAT1/miR-320b/MSI2 axis in human cisplatin resistant OC.

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Availability of data and materials

The analyzed data sets generated during the present study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The present study was approved by the ethical review committee of Eye, Ear, Nose and Larynx Hospital, Liaocheng People's Hospital.
Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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Yan P, Su Z, Zhang Z, Gao T. LncRNA NEAT1 enhances the resistance of anaplastic


Figure legends

Fig. 1 The level of NEAT1 was notably increased in cisplatin resistant OC cell lines. (A and B) The cell viability and IC₅₀ of cisplatin in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells were assessed by CCK-8 assay. (C and D) Migration and invasion of OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells were determined by transwell assay. (E and F) The protein levels of MMP2 and MMP9 were detected by
western blot. (G and H) The expression of NEAT1 was detected in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells by qRT-PCR. Data are represented as means ± SD; *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001.

Fig. 2 Knockdown of NEAT1 enhanced cisplatin sensitivity of cisplatin resistance OC cell lines.

OVCAR-3/DDP and HEY/DDP cells were transfected with si-NEAT1 or si-control, respectively. (A and B) The level of NEAT1 was determined by qRT-PCR. (C and D) CCK-8 assay was carried out to detect cell viability and IC_{50} of cisplatin. (E and F) Migration and invasion were determined by transwell assay (G and H) The protein levels of MMP2 and MMP9 were detected by western blot. Data are represented as means ± SD; **P < 0.01, ***P < 0.001, ****P < 0.0001.

Fig. 3 MiR-320b was a target gene of NEAT1.

(A) The relationship between NEAT1 and miR-320b was predicted by LncBase Predicted v.2. (B and C) NEAT1 WT and NEAT1 MUT was transfected into OVCAR-3/DDP and HEY/DDP cells, severally, and luciferase activity was determined by dual-luciferase reporter assay in vitro. (D-G) RIP assay and RNA pull down assay were performed to verify the interrelation between NEAT1 and miR-320b in SKOV3/DDP and A2780/DDP cells. Data are represented as means ± SD; ***P < 0.001, ****P < 0.0001.

Fig. 4 The effect of knockdown of NEAT1 on cisplatin sensitivity was reversed by
miR-320b in cisplatin resistance OC cell lines.

SKOV3/DDP and A2780/DDP cells were transfected with si-control, si-NEAT1, si-NEAT1+miR-control or si-NEAT1+miR-320b, respectively. (A and B) The level of miR-320b was measured by qRT-PCR. (C and D) CCK-8 assay was carried out to detect cell viability and IC50 of cisplatin. (E and F) Migration and invasion were determined by transwell assay. (G and H) The protein levels of MMP2 and MMP9 were detected by western blot. Date are represented as means ± SD; **P < 0.01, ***P < 0.001, ****P < 0.0001.

Fig. 5 MiR-320b directly targeted MSI2.

(A) Targetscan was used to predict the relationship between miR-320b and MSI2. (B and C) Luciferase activity was measured utilizing dual-luciferase reporter assay. (D) The expression of MSI2 in OVCAR-3/DDP and HEY/DDP cells transfected with miR-control or miR-320b was determined by western blot, severally. (E and F) The expression of MSI2 in OVCAR-3/DDP and HEY/DDP cells transfected with si-control, si-NEAT1, si-NEAT1+miR-control or si-NEAT1+miR-320b was determined by western blot, severally. (G and H) expression of MSI2 in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells was determined by western blot. Date are represented as means ± SD; ***P < 0.001, ****P < 0.0001.

Fig. 6 Overexpression of MSI2 restored the effect of miR-320b on cisplatin sensitivity in cisplatin resistance OC cell lines.
MiR-control, miR-320b, miR-320b+pcDNA or miR-320b+MSI2 were transfected into OVCAR-3/DDP and HEY/DDP cells, respectively. (A and B) The expression of MSI2 were measured by and western blot. (C and D) CCK-8 assay was carried out to detect cell viability and IC50 of cisplatin. (E and F) Migration and invasion were determined by transwell assay (G and H) The protein levels of MMP2 and MMP9 were detected by western blot. Date are represented as means ± SD; **P < 0.01, ***P < 0.001, ****P < 0.0001.

Fig. 7 Knockdown of NEAT1 retarded the progression of OC tumors after cured with cisplatin or not in vivo.

OVCAR-3/DDP cells transfected with sh-control or sh-NEAT1 were injected nude mice subcutaneously. (A) Tumor volumes were measured every three days after injection for one week. (B) Tumor weights were also measured after mice were sacrificed. (C and D) QRT-PCT was carried out to determine the levels of NEAT1 and miR-320b. (E) The expression of MSI2 was analyzed western blot. Date are represented as means ± SD; *P<0.05, ***P < 0.001, ****P < 0.0001.

Fig. S1 MiR-194, let-7a-5p and miR-9-5p were upregulated, while ZEB1, Rsf-1 and SPAG9 were downregulated in cisplatin resistant OC cells. (A and B) The expression of miR-194, let-7a-5p and miR-9-5p in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells was detected by qRT-PCR. (C and D) The mRNA levels of ZEB1, Rsf-1 and SPAG9 were in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells was
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Fig. 1 The level of NEAT1 was notably increased in cisplatin resistant OC cell lines. 
(A and B) The cell viability and IC50 of cisplatin in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells were assessed by CCK-8 assay. (C and D) Migration and invasion of OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells were determined by transwell assay. 
(E and F) The protein levels of MMP2 and MMP9 were detected by western blot. (G and H) The expression of NEAT1 was detected in OVCAR-3, OVCAR-3/DDP, HEY, and HEY/DDP cells by qRT-PCR. Date are represented as means ± SD; *P<0.05 **P < 0.01, ***P < 0.001, ****P < 0.0001.
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