

Dietary methionine restriction attenuates cognitive dysfunction associated with sleep deprivation by promoting production of hydrogen sulfide, upregulating expression of BDNF and suppressing neuro-inflammation in rats

Keywords

cognitive dysfunction, methionine, H₂S, sleep deprivation, neuroinflammation

Abstract

Introduction

In this study, we aimed to study the potential role of H₂S and the application of dietary methionine restriction (DMR) in the treatment of cognitive dysfunction in sleep deprived rat models.

Material and methods

The rat groups were established as a Control group, a sleep deprivation (SD) group, a SD + NaSH group and a SD + DMR group. Behavioral data was studied via Morris water maze test. TUNEL assay, real-time PCR, immunohistochemistry (IHC) and enzyme-linked immunosorbent assay (ELISA) were performed to study the differences of cognitive impairment related genes and proteins in different rat groups.

Results

Path length and escape latency were higher in the sleep deprived rats compared with the control rats, which were subsequently recovered by the treatment of NaSH or DMR. Also, DMR most significantly recovered the cognitive impairment and neuron status of sleep deprived rats compared with the administration of NaSH. The elevated level of hippocampal Iba1 and H₂S production was recovered by NaSH and DMR, and the expressions of hippocampal phenotypic-related genes including NOS, CD68, CD32 and CD206 mRNA also showed similar trend as hippocampal Iba1. Meanwhile, the increased relative expression of IL-6 and IL-4 were recovered by DMR in sleep deprived rats, while the reduced level of hippocampal brain-derived neurotrophic factor (BDNF) and cystathionine γ -lyase (CSE) were elevated by the treatment of NaSH and MDR, with MDR exhibiting the most significant effect.

Conclusions

The application of DMR could attenuate cognitive dysfunction in sleep deprivation rats by upregulating the production of H₂S and BDNF expression and alleviating neuro-inflammation responses.

**1Dietary methionine restriction attenuates cognitive
2dysfunction associated with sleep deprivation by promoting
3production of hydrogen sulfide, upregulating expression of
4BDNF and suppressing neuro-inflammation in rats**

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15Abstract

16In this study, we aimed to study the potential role of H₂S and the
17application of dietary methionine restriction (DMR) in the treatment of
18cognitive dysfunction in ~~sleep-deprived~~[sleep-deprived](#) rat models. The
19rat groups were established as a ~~Control~~[Sham](#) group, a sleep
20deprivation (SD) group, a SD + NaSH group and a SD + DMR group.
21Behavioral data was studied via Morris water maze test. TUNEL assay,
22real-time PCR, immunohistochemistry (IHC) and enzyme-linked
23immunosorbent assay (ELISA) were performed to study the differences
24of cognitive impairment related genes and proteins in different rat
25groups. Path length and escape latency were higher in the ~~sleep~~
26~~deprived~~[sleep-deprived](#) rats compared with the ~~sham-operated~~[control](#)
27rats, which were subsequently recovered by the treatment of NaSH or
28DMR. Also, DMR most significantly recovered the cognitive impairment
29and neuron status of ~~sleep-deprived~~[sleep-deprived](#) rats compared with
30the administration of NaSH. The elevated level of hippocampal Iba1
31and H₂S production was recovered by NaSH and DMR, and the

32expressions of hippocampal phenotypic-related genes including NOS,
33CD68, CD32 and CD206 mRNA also showed similar trend as
34hippocampal Iba1. Meanwhile, the increased relative expression of IL-6
35and IL-4 were recovered by DMR in ~~sleep-deprived~~sleep-deprived rats,
36while the reduced level of hippocampal brain-derived neurotrophic
37factor (BDNF) and cystathionine γ -lyase (CSE) were elevated by the
38treatment of NaSH and MDR, with MDR exhibiting the most significant
39effect. The application of DMR could attenuate cognitive dysfunction in
40sleep deprivation rats by upregulating the production of H₂S and BDNF
41expression and alleviating neuro-inflammation responses.

42**Running title:** Dietary methionine restriction attenuates cognitive
43dysfunction

44**Keywords:** methionine, sleep deprivation, cognitive dysfunction, H₂S,
45neuro~~n~~-inflammation

46**Introduction**

47Sleep is a fundamental physiological process crucial to one's health,
48and sleep disorder has been listed as one of the important health-
49related issues worldwide. As previously reported, sleep deprivation ~~not~~
50only impairs vigilant attention (1), increases the formation of false
51memory (2) and weakens the cognitive performance (3), ~~but also~~
52~~damages the immune system and even induce diseases including~~
53~~hypertension and diabetes (4)~~. Moreover, ~~compared with other patient~~
54~~groups~~, sleep deprivation raised more concerns in menopausal women
55(4, 5). Therefore, ~~to seek for other~~ treatment methods which may
56presents few side effects for the management of sleep disturbances
57are necessary, especially for menopausal women.

58Methionine functions as a precursor of homocysteine, a non-protein-
59forming sulfur amino acid (6). Homocysteine was found to be
60associated with cognitive impairment as the increased total plasma
61homocysteine level has been found to induce cognitive decline and
62white matter damage (7). And the deprivation of methionine not only

63 induce rapid reduction of body weight by influencing the metabolism
64 mechanisms of obese adults (8, 9), but also exert therapeutic effects in
65 the management of diseases such as glioma (10), hepatosteatosis
66 (11), and age-associated cognitive decline (12-14). For example, in
67 their study which explore the effect of dietary methionine restriction
68 (DMR) on age-related cognitive decline, Ren et al. found that DMR
69 elevated expression of fibroblast growth factor 21 (FGF21), a key
70 mediator for neuro functions and hippocampal mitochondrial
71 biogenesis (12). And by regulating expression of FGF21, DMR also
72 modulates energy balance and adipose tissue remodeling (13).
73 Therefore, DMR is best known as dietary treatment methods which
74 could reduce the incidence age-related disease.

75 Hydrogen sulfide (H₂S) has been widely acknowledged as a key
76 signaling factor which exhibit positive effects in the management of
77 sleep deprivation. For example, H₂S was found to antagonize
78 hippocampal damage in sleep-deprived rat models (15), and further
79 alleviate cognitive dysfunction in sleep-deprived rat models by
80 suppressing excessive hippocampal autophagy (16). Moreover, H₂S
81 could also prevent the depression and anxiety symptoms caused by
82 sleep deprivation by modulating the signaling pathway of silence
83 information regulating factor-1 (Sirt-1), which plays a key role in
84 neuroprotection— (17, 18) ~~(17, 18)~~. Also, multiple investigations
85 demonstrated the anti-oxidative effect, ~~anti-ER stress, and~~
86 antiapoptotic properties of H₂S in central nervous system (19, 20) ~~(19-~~
87 ~~21)~~, the performance of which is strongly associated with the adverse
88 effects caused by sleep deprivation (21).

89 Among previous publications which reported the effect of H₂S in brain
90 functions, a report by Wei et al. suggested that in SD rats treated with
91 H₂S donor NaHS, the endogenous production of H₂S as well as the
92 expression of brain-derived neurotrophic factor (BDNF) was evidently
93 increased, leading to the attenuated endoplasmic reticulum stress and

94 neuronal apoptosis in hippocampus (22). Meanwhile, it is also reported
95 that paradoxical sleep deprivation could down-regulate the level of
96 hippocampal BDNF (23, 24). And BDNF has been proved to play key
97 roles in cognition-related functions including the efficacy of exercise on
98 synaptic plasticity in spatial learning (25) and synaptic plasticity (26). In
99 this study, we aimed to study the role of H₂S and the application of
100 ~~dietary methionine restriction~~ [DMR](#) in the treatment of cognitive
101 dysfunction in ~~sleep-deprived~~ [sleep-deprived](#) rat models.

102 **Methods**

103 **Animals**

104 ~~Thirty-two~~ 32 male [Sprague Dawley](#) ~~SD~~ rats (weight between 220 g to
105 255 g; 6 weeks old) were enrolled in this study. The rats were divided
106 into four groups with 8 random rats in each group. All animals were
107 housed under a 12-h light-dark cycle with free access to food and
108 water. The housing facility temperature was maintained between 22 °C
109 to 25 °C, with the humidity ranging between 45 % to 60%. All
110 procedures were carried out according to the instructions in the
111 National Research Council's Guide for the Care and Use of Laboratory
112 Animals and the institutional Ethics Committee approved the study (ID:
113 H-HNNHY-190813X01).

114 **Experimental groups**

115 The animal groups were established as a ~~Control~~ [SHAM](#) group, a sleep
116 deprivation (SD) group, a SD+[Sodium hydrosulfide \(NaSH\)](#) group, a
117 SD+DMR group. The rats assigned to the ~~SHAM~~ [Control](#) group were kept
118 in the home cages, and the rats subjected to establish SD models were
119 exercised in the water pool designed for the multiple platforms method
120 (MPM) (27). There were 10 columns located 2 cm above the water
121 surface in the pool, and each column was 7 cm in diameter. These
122 columns enabled the rats to move between different platforms.
123 Moreover, once the rapid eye movement sleep (REM) occurs in a rat,
124 the lost muscle tone would result in contact with water, thus achieving

125the establishment of SD rats. For rats in the SD+NaSH group, the SD
126rats were injected with 100 $\mu\text{mol/mL}$ NaHS resolved in PBS. For rats in
127the SD+DMR group, the rats were fed with diets containing a restricted
128dosage of 0.15% D,L-methionine for 72 hours (28).

129**Morris water maze (MWM) test**

130MWM test was performed with the rat groups to evaluate the effect of
131DMR on rat spatial learning and memory according to previously
132published methods (29, 30). The MWM test equipment is a ring-shaped
133pool divided into four quadrants, with 155 cm in diameter and 70cm in
134height. The pool was filled with ~~hygienic~~ water with 40 cm in depth.
135The temperature of water pool is maintained at 23–25°C. Before the
136grouping of experiment animals, all rats were trained to find an escape
137platform with 10 cm in diameter placed above the water surface under
138the guidance of visual cues for at least 3 hours. These visual cues
139during the training contributed to the spatial learning of rats.
140Subsequently, after 30 mins of animal group establishment, the
141behavioral test was carried out by releasing the rats into the pool for
142the search of unmarked hidden platform with submerged 1 cm below
143the water surface in the target quadrant. Each experiment phase lasts
144for 60 seconds for 4 phases for each quadrant. The interval between
145each phase is 60 seconds. And the MWM test was repeated in 3
146quadrants. The path length and escape latency to reach the hidden
147platform were recorded to indicate the changes of spatial learning in
148different animal groups.

149Additionally, a probe test was performed with the rat groups to
150evaluate the effect of DMR on rat spatial memory. The hidden platform
151was removed and the rats were individually released into the pool to
152swim for 60 seconds. The percentage time and distance in the target
153quadrant where the platform was submerged before were recorded to
154indicate the changes of spatial memory in different animal groups.

155**TUNEL**

156The neuron apoptosis was evaluated by TUNEL assays. Prior to TUNEL
157assays, the brain tissues were processed with PBS, and 3% hydrogen
158peroxide to block the endogenous peroxidase activities. After being
159dried, the tissues were subjected to TUNEL assays with a TUNEL assay
160kit (Beyotime Biotechnology, Shanghai, China) according to the kit's
161instruction. Subsequently, the sections were stained with a DAB
162substrate (Thermo Fisher Scientific, MA, US) before being observed and
163assessed under an Olympus light microscope (Olympus Corporation,
164JP).

165RNA isolation and real-time PCR

166Total RNA was isolated by from peripheral blood samples utilizing a
167Trizol reagent (Invitrogen, Carlsbad, CA). And the extracted total RNA
168was reverse-transcribed into cDNA using a Taqman Reverse
169Transcription assay kit (Invitrogen, Carlsbad, CA) according to the kit's
170instructions. Finally, real-time PCR analysis was performed to evaluate
171the expression level of Iba1 mRNA, iNOS mRNA, CD68 mRNA, CD32
172mRNA, CD206 mRNA, BDNF mRNA, cystathionine γ -lyase (CSE) mRNA
173on a PRISM 7500 real time PCR machine (Applied Biosystems, Foster
174City, CA). The quantification of relative expression of these genes
175utilized GAPDH as an internal reference gene following the $2^{-\Delta\Delta Ct}$
176calculation method.

177IHC

178The levels of Iba1, BDNF, CSE in the hippocampal tissues were
179investigated by IHC assays. Prior to the IHC assay, the collected
180hippocampus tissues were fixed in 10% formalin solution (pH value =
1817.4), paraffin embedded, sliced into 2 mm thick sections and de-
182paraffined using xylene. Subsequently, the hippocampal sections were
183blocked with 0.3% hydrogen peroxide for 20 min and blocked with 10%
184FBS. Finally, the hippocampal sections were incubated with primary
185antibodies against Iba1 (Catalogue number: ab178846, Abcam,
186Cambridge, MA), BDNF (Catalogue number: ab178846, Abcam,

187Cambridge, MA), CSE (Catalogue number: #30068, Cell Signaling
188Technology, MA, US) overnight, followed by the incubation with
189horseradish peroxidase-conjugated secondary antibodies (Abcam,
190Cambridge, MA) for 60 min. Finally, the sections were counter-stained
191with a DAB substrate (Sigma-Aldrich, St. Louis, MO) before being
192observed and assessed under an Olympus light microscope (Olympus,
193Tokyo, Japan).

194**ELISA**

195To evaluate the hippocampal production of IL-6, IL-4, H₂S in the animal
196models, ELISA assays were performed with ELISA kits of IL-6, IL-4, H₂S
197(Abcam, Cambridge, UK) according to the kit's instructions. Prior to the
198ELISA assays, the collected hippocampus tissues were processed with
199RIPA lysis buffer and centrifugated for the removal cell debris.

200**Statistical analysis**

201Data obtained from the above experiments were collected and
202analyzed with statistical analysis software SPSS v 19.0 (IBM, NY, US).
203The comparisons between multiple groups were analyzed using one-
204way ANOVA, and Tukey's test was used as the post-hoc test. All data
205were expressed as mean \pm standard deviation, and the statistical
206significance level was set as less than $P < 0.05$.

207**Results**

208**Recovery effect of DMR on the behavioral data of sleep-** 209**deprived rats**

210Compared with the [controlsham-operated](#) rats, the [sleep](#)
211[deprivedsleep-deprived](#) rats travelled longer distance (Fig.1A) and
212spent more time (Fig.1B) to reach the hidden platform. However, NaSH
213treatment and DMR both reduced the path length and escape latency
214during platforms, and DMR exhibited a more significant recovery effect
215upon these behavioral data. Moreover, when observing the percentage
216of their swimming distance and escape latencies among the rat groups
217(Fig.1C), compared with the rats in the [SHAMControl](#) group, a

218 significant reduction in the ~~sleep-deprived~~ sleep-deprived rats was
219 found. Also, the rats in the SD + NaSH and the SD + DMR groups spent
220 less time and travelled less distance in finding the target quadrant
221 than the rats in the SD group. Besides, TUNEL assay upon the
222 apoptosis of neurons (Fig.1D) showed significant recovery of neurons in
223 ~~sleep-deprived~~ sleep-deprived rats treated with NaSH or DMR, with the
224 recovery effect of DMR being most significant.

225 **Recovery effect of DMR on the expression of hippocampal Iba1**

226 As shown in Figure 2, we measured the level of hippocampal Iba1 in
227 the rat models to study the effect of DMR upon the expression of Iba1
228 in vivo. IHC assay (Fig.2A and 2B) showed that the number of Iba1
229 positive cells was evidently higher in the ~~sleep-deprived~~ sleep-deprived
230 rats compared with the ShamControl group, and the administration of
231 NaSH and DMR both reduced the number of Iba1 positive cells.
232 Specially, the number of Iba1 positive cells in the SD + DMR group is
233 lowest among all ~~sleep-deprived~~ sleep-deprived rats. Similarly, the
234 relative expression of Iba1 mRNA (Fig.2C) also presented the same
235 trend among the rat groups, indicating the recovery effect of DMR on
236 the expression of hippocampal Iba1.

237 **Recovery effect of DMR on the expressions of phenotypic-** 238 **related genes in hippocampus**

239 The upregulated expression of hippocampal phenotypic-related genes,
240 including NOS mRNA, CD68 mRNA, CD32 mRNA and CD206 mRNA,
241 commonly characterize the incidence of cognitive impairment. As
242 shown in Figure 3, we observed the expression of NOS mRNA (Fig.3A),
243 CD68 mRNA (Fig.3B), CD32 mRNA (Fig.3C) and CD206 mRNA (Fig.3D)
244 in the hippocampal tissues collected from the rat models. Accordingly,
245 we found that the relative hippocampal expressions of phenotypic-
246 related genes were all evidently increased in ~~sleep-deprived~~ sleep-
247 deprived rats compared with sham-operated control rats, and these
248 highly expressed genes in ~~sleep-deprived~~ sleep-deprived rats were

249significantly recovered by the administration of NaSH or the application
250of DMR. Specially, the recovery effect of DMR was more significant
251than that of the NaSH treatment. Also, ELISA assays indicated that the
252increased relative expression of IL-6 (Fig.3E) and IL-4 (Fig.3F) were also
253recovered by DMR in ~~sleep-deprived~~ rats.

254**Promotive effect of DMR on the expression of hippocampal** 255**BDNF and CSE**

256In this study, we also evaluated the mRNA level of hippocampal BDNF
257(Fig.4) and CSE (Fig.5). Accordingly, the numbers of BDNF positive cells
258(Fig.4A and 4B) or CSE positive cells (Fig.5A and 5B) were both
259significantly decreased due to the deprivation of sleep in rat models.
260And compared with the SD group, DMR most significantly promoted the
261reduced numbers of BDNF and CSE positive cells. Meanwhile, the
262evidently downregulated hippocampal BDNF mRNA (Fig.4C)
263expressions were also elevated by the treatment of NaSH and MDR,
264with MDR exhibiting the most significant effect. Similar results were
265obtained in respect to the relative expressions of CSE mRNA (Fig.5C)
266and H₂S production (Fig.5D).

267**Discussion**

268~~Sleep deprivation is characterized as a state of inadequate quantity or~~
269~~quality of sleep (31). And sleep deprivation has been reported to~~
270~~induce brain function impairment (1), aggravate oxidative stress and~~
271~~inflammation (32), activate DNA damage response (33), suppresses~~
272~~neuronal proliferation (34), and reduce hippocampal gliogenesis (35).~~
273~~Moreover, epidemiological reports have stated that sleep deprivation,~~
274~~especially chronic sleep deprivation, increased the risk of diseases~~
275~~including coronary artery disease, hypertension, arrhythmias, diabetes~~
276~~and obesity (36). Among all the personal characteristics, aging is a~~
277~~factor which influences the individual's ability of deal with sleep~~
278~~deprivation, supported by the fact that the brain function of an aged~~
279~~individual is generally weaker (37). Besides, sex is also an influencing~~

280factor. It was reported that women, especially menopausal women,
281may suffer more from sleep deprivation than man with slower recovery
282duration (38). In this study, by using indicators including path length,
283escape latency, cognitive impairment and neuron status, we studied
284the effect of dietary methionine restriction-DMR on sleep deprivation,
285and cognitive dysfunction associated with sleep deprivation.
286Behavioral data such as path length and escape latency were higher in
287the sleep-deprivedsleep-deprived rats compared with the sham-
288operatedcontrol rats, which were subsequently recovered by the
289treatment of NaSH or DMR. Also, DMR most significantly recovered the
290cognitive impairment and neuron status of sleep-deprivedsleep-
291deprived rats compared with the administration of NaSH.

292As a health intervention which is deemed to exert positive effect on the
293cognitive dysfunctions in postmenopausal women, the method of
294exercise could also alter the expression of hippocampal BDNF (31).
295BDNF not only functions as a mediator in CNS, but also provides
296neurotrophic and neuroprotective support to different subpopulations
297of neurons (32). The decreased hippocampal BDNF levels were
298reported to be associated with the impairment in the cognitive function
299in aged individuals (33). Moreover, sleep deprivation after contextual
300conditioning has been proved to inhibit the level of BDNF and the
301according signaling (34). In this study, we found that the mRNA and
302protein level of hippocampal BDNF and CSE were reduced in sleep
303deprivedsleep-deprived rats, which is in-consistent with the results of
304previous studies. And the reduced BDNF levels were elevated by the
305treatment of NaSH and MDR, with MDR exhibiting the most significant
306effect.

307The hippocampal protective effect of H₂S against cognitive dysfunction
308induced by sleep deprivation has been validated in sleep-deprived rat
309models (15, 35). Meanwhile, CSE, which is produced in neurons, is the
310biosynthetic enzyme for the H₂S (36). In CSE mutant mice, the

311 suppressive effect of NF- κ B upon apoptosis is diminished as well (36).
312 Moreover, NF- κ B regulates the immune and inflammatory responses in
313 brain ischemia-reperfusion rats (37, 38). Therefore, H₂S production is
314 also associated with neuro-inflammatory responses. In this study, we
315 found that the elevated level of hippocampal H₂S production was
316 recovered by NaSH and DMR, with DMR exhibiting the most significant
317 recovery effect in ~~sleep-deprived~~ rats.

318 Several molecules have been reported to be associated with cognitive
319 dysfunctions. For example, the up-regulation of CD68 was reported to
320 be correlated with longer latency times of normally aged rats in the
321 MWM test (39). And the combination of CD68 and Iba1, another
322 microglial marker, could function as biomarkers for the characterization
323 of microglial phenotype in age-associated deep subcortical white
324 matter lesions (40). Moreover, CD32 also functions as a surface protein
325 on brain microglia for the characterization of M2b macrophage in
326 Alzheimer's disease patients (41). IL-6 was reported to be associated
327 with cognitive function (42) while IL-4 could attenuate reference
328 memory impairment by modulating the hippocampal-BDNF signaling
329 pathway (43). In this study, the expressions of hippocampal
330 phenotypic-related genes including NOS mRNA, CD68 mRNA, CD32
331 mRNA and CD206 mRNA also showed similar trend as hippocampal
332 Iba1. Meanwhile, the increased relative expression of IL-6 and IL-4
333 were also recovered by DMR in ~~sleep-deprived~~ rats.

334 However, this study is limited as further validation due to ~~the lack of~~
335 ~~further validation of the relatively small sample size in the animal~~
336 ~~experiment. Also, the effect of DMR should be further verified~~ in clinical
337 practice in sleep-deprived patients, especially the mechanism of
338 modulating the production of hydrogen sulfide and the level of BDNF.
339 Therefore, in our future studies, larger sample size for the more
340 comprehensive animal study will be exploited. And further
341 observations in clinical trials are also necessary.

342 **Conclusions**

343 In conclusion, we identified that the application of DMR could attenuate
344 cognitive dysfunction in sleep deprivation rat models.

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354 **Competing interests**

355 None.

356 **Data availability statement**

357 The datasets generated in this study is available from the
358 corresponding authors upon reasonable request.

359 **Figure legends**

360 **Figure 1**

361 Recovery effect of DMR on the behavioral data of sleep-deprived rats
362 ([scale bar = 50 μm](#))

363 A: The increased path length of sleep-deprived rats to reach the
364 hidden platform was inhibited by NaSH or DMR;

365 B: The prolonged escape latency of ~~sleep-deprived~~ sleep-deprived rats
366 to find the hidden platform was reduced by NaSH or DMR;

367 C: The percentage of distance, time and crossing in the target
368 quadrant during the probe phase of ~~sleep-deprived~~ sleep-deprived rats
369 was increased by NaSH or DMR;

370 D: TUNEL assay indicated that NaSH or DMR could alleviated the
371 aggravated neuron apoptosis in ~~sleep-deprived~~ sleep-deprived rats.

372 **Figure 2**

373Recovery effect of DMR on the expression of hippocampal Iba1 mRNA
374and protein (* P value < 0.05 vs. [SHAMControl](#) group; ** P value < 0.05
375vs. SD group; # P value < 0.05 vs. SD+NaSH group; [scale bar = 50](#)
376[μm](#)).

377A: IHC assay upon Iba1 expression in hippocampal tissues showed that
378NaSH or DMR could reduce the up-regulated Iba1 expression;

379B: The increased number of Iba1 positive microglia in [sleep](#)
380[deprivedsleep-deprived](#) rats was reduced by NaSH or DMR;

381C: The relative expression level of Iba1 mRNA was elevated in [sleep](#)
382[deprivedsleep-deprived](#) rats, and NaSH or DMR could restore the
383elevated Iba1 mRNA expression.

384**Figure 3**

385Recovery effect of DMR on the expressions of phenotypic-related genes
386in hippocampus (* P value < 0.05 vs. [ControlSHAM](#) group; ** P value <
3870.05 vs. SD group; # P value < 0.05 vs. SD+NaSH group).

388A: Relative expression of iNOS mRNA was increased in [sleep](#)
389[deprivedsleep-deprived](#) rats, while both NaSH and DMR restored the
390dysregulation of iNOS mRNA level;

391B: Relative expression of CD68 mRNA was increased in [sleep](#)
392[deprivedsleep-deprived](#) rats, while both NaSH and DMR restored the
393dysregulation of CD68 mRNA level;

394C: Relative expression of CD32 mRNA was increased in [sleep](#)
395[deprivedsleep-deprived](#) rats, while both NaSH and DMR restored the
396dysregulation of CD32 mRNA level;

397D: Relative expression of CD206 mRNA was increased in [sleep](#)
398[deprivedsleep-deprived](#) rats, while both NaSH and DMR restored the
399dysregulation of CD206 mRNA level;

400E: Relative expression of IL-6 was increased in [sleep-deprivedsleep-](#)
401[deprived](#) rats, which was suppressed by treatment of NaSH and DMR;

402F: Relative expression of IL-4 was increased in [sleep-deprivedsleep-](#)
403[deprived](#) rats, which was suppressed by treatment of NaSH and DMR.

404**Figure 4**

405Promotive effect of DMR on the expression of hippocampal BDNF mRNA
406and protein (* P value < 0.05 vs. [ControlSHAM](#) group; ** P value < 0.05
407vs. SD group; # P value < 0.05 vs. SD+NaSH group; [scale bar = 50](#)
408[μm](#)).

409A: IHC assay showed that hippocampal BDNF expression was reduced
410in [sleep-deprivedsleep-deprived](#) rats, and NaSH or DMR antagonized
411the changes of BDNF expression;

412B: The reduced number of BDNF positive microglia in [sleep](#)
413[deprivedsleep-deprived](#) rats was recovered by NaSH or DMR;

414C: The relative expression level of BDNF mRNA was reduced in [sleep](#)
415[deprivedsleep-deprived](#) rats, and NaSH or DMR antagonized the
416changes of BDNF expression.

417**Figure 5**

418Promotive effect of DMR on the expression of hippocampal CSE mRNA
419and production of H₂S (* P value < 0.05 vs. [ControlSHAM](#) group; ** P
420value < 0.05 vs. SD group; # P value < 0.05 vs. SD+NaSH group; [scale](#)
421[bar = 50 μm](#))

422A: IHC assay showed that hippocampal CSE expression was reduced in
423[sleep-deprivedsleep-deprived](#) rats, and NaSH or DMR antagonized the
424changes of BDNF expression;

425B: The reduced number of CSE positive microglia in [sleep](#)
426[deprivedsleep-deprived](#) rats was recovered by NaSH or DMR;

427C: The relative expression level of CSE mRNA was reduced in [sleep](#)
428[deprivedsleep-deprived](#) rats, and NaSH or DMR antagonized the
429changes of BDNF expression;

430D: H₂S production was lowest in [sleep-deprivedsleep-deprived](#) rats, and
431NaSH or DMR treatment antagonized the reduction of H₂S production.

432

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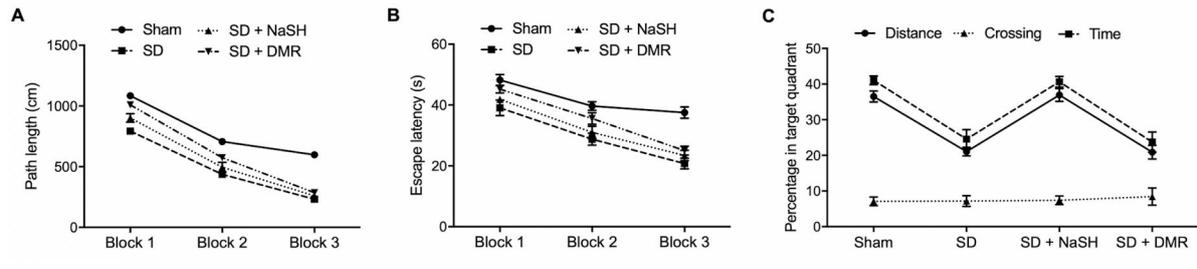
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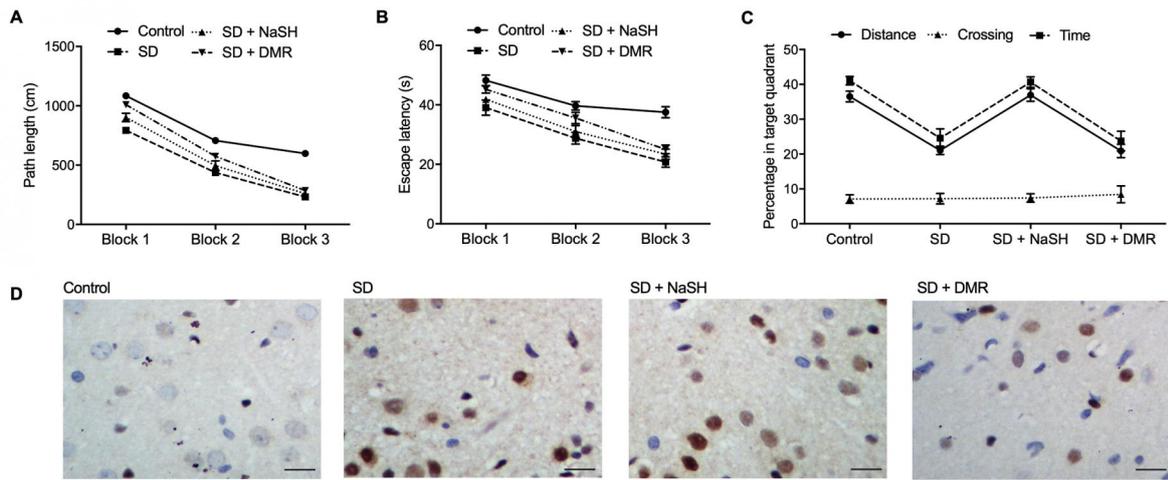


Figure 1

Recovery effect of DMR on the behavioral data of sleep-deprived rats (scale bar = 50 μ m)

A: The increased path length of sleep-deprived rats to reach the hidden platform was inhibited by NaSH or DMR;

B: The prolonged escape latency of sleep-deprived rats to find the hidden platform was reduced by NaSH or DMR;

C: The percentage of distance, time and crossing in the target quadrant during the probe phase of sleep-deprived rats was increased by NaSH or DMR;

D: TUNEL assay indicated that NaSH or DMR could alleviate the aggravated neuron apoptosis in sleep-deprived rats.

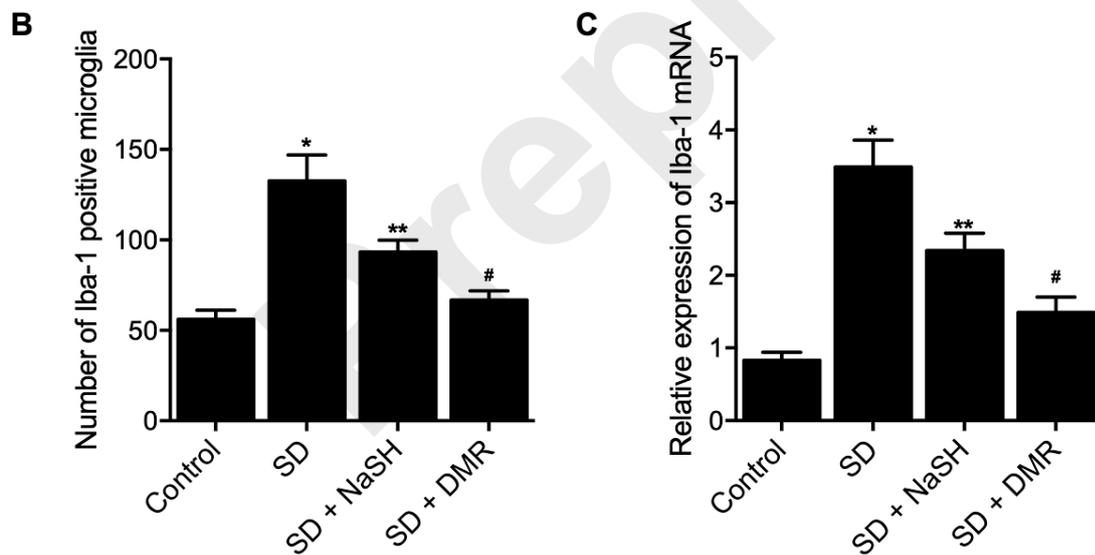
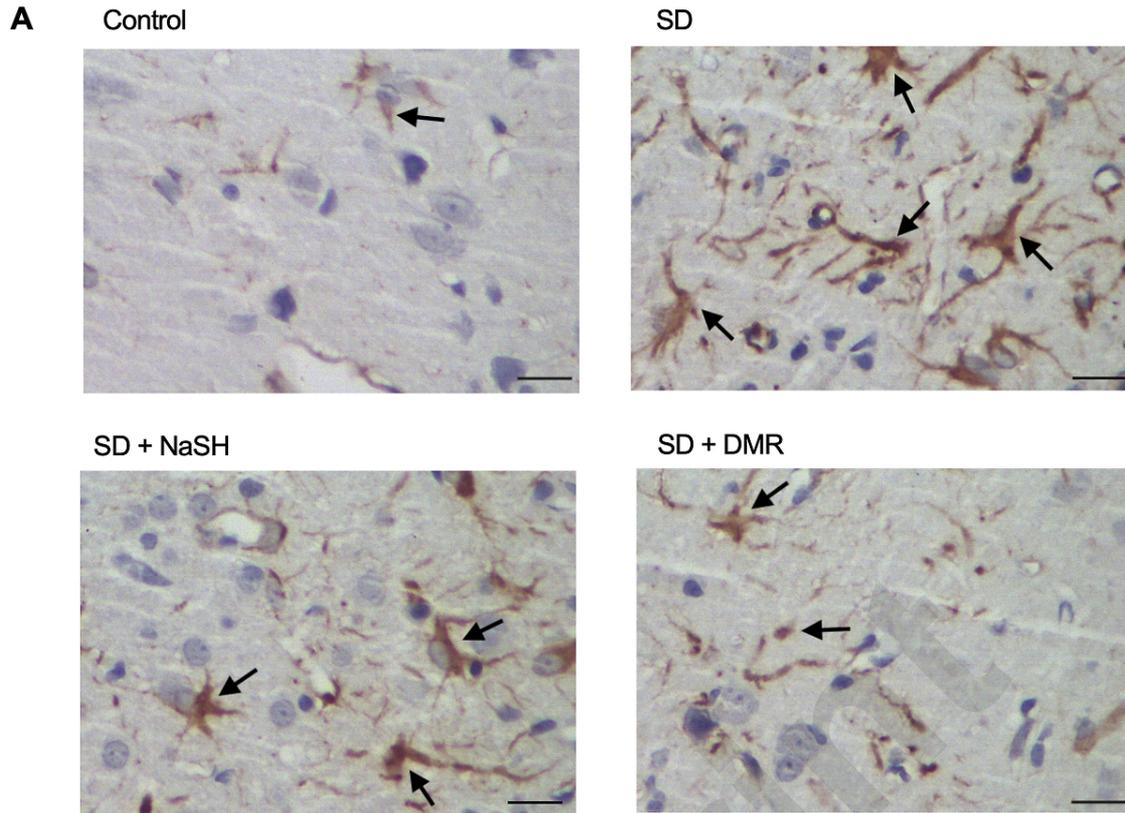


Figure 2

Recovery effect of DMR on the expression of hippocampal Iba1 mRNA and protein (* P value < 0.05 vs. Control group; ** P value < 0.05 vs. SD group; # P value < 0.05 vs. SD+NaSH group; scale bar = 50 μ m).

A: IHC assay upon Iba1 expression in hippocampal tissues showed that NaSH or DMR could reduce the up-regulated Iba1 expression;

B: The increased number of Iba1 positive microglia in sleep-deprived rats was reduced by NaSH or DMR;

C: The relative expression level of Iba1 mRNA was elevated in sleep-deprived rats, and NaSH or DMR could restored the elevated Iba1 mRNA expression.

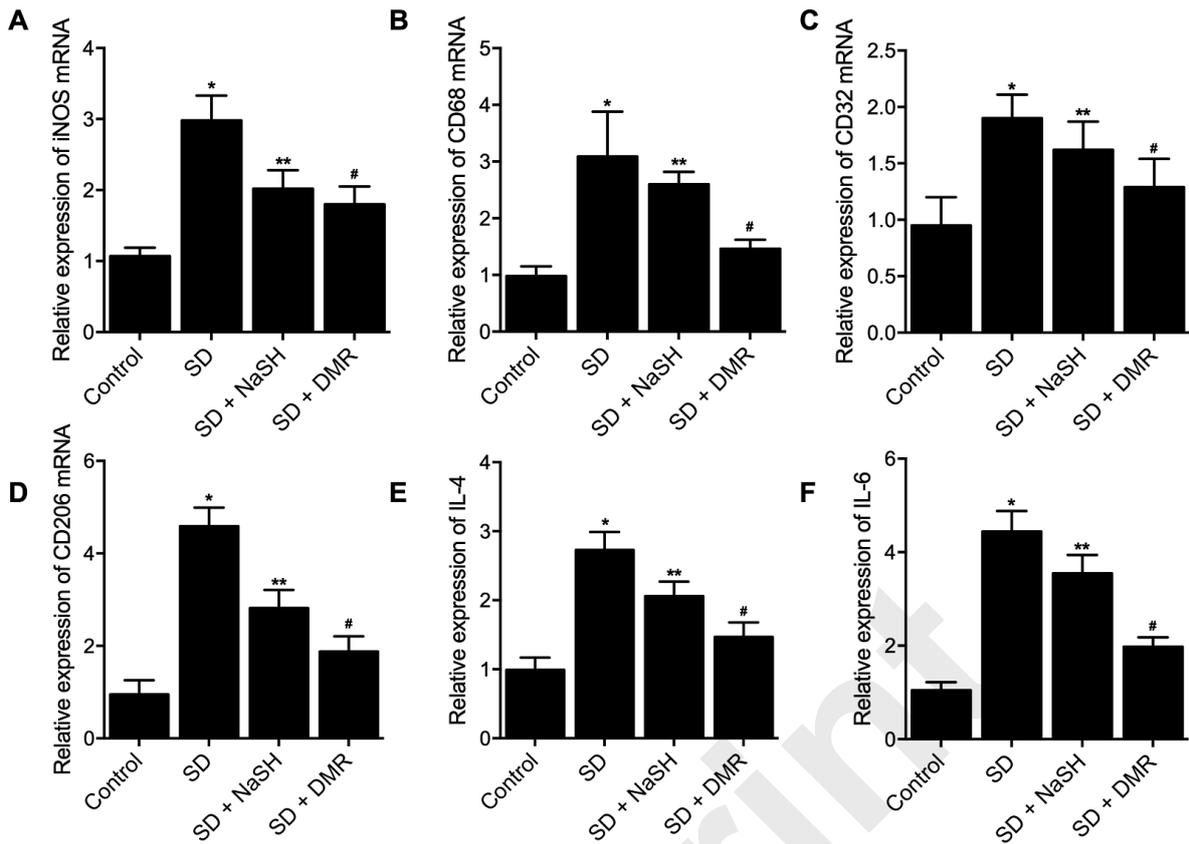


Figure 3

Recovery effect of DMR on the expressions of phenotypic-related genes in hippocampus (* P value < 0.05 vs. Control group; ** P value < 0.05 vs. SD group; # P value < 0.05 vs. SD+NaSH group).

A: Relative expression of iNOS mRNA was increased in sleep-deprived rats, while both NaSH and DMR restored the dysregulation of iNOS mRNA level;

B: Relative expression of CD68 mRNA was increased in sleep-deprived rats, while both NaSH and DMR restored the dysregulation of CD68 mRNA level;

C: Relative expression of CD32 mRNA was increased in sleep-deprived rats, while both NaSH and DMR restored the dysregulation of CD32 mRNA level;

D: Relative expression of CD206 mRNA was increased in sleep-deprived rats, while both NaSH and DMR restored the dysregulation of CD206 mRNA level;

E: Relative expression of IL-6 was increased in sleep-deprived rats, which was suppressed by treatment of NaSH and DMR;

F: Relative expression of IL-4 was increased in sleep-deprived rats, which was suppressed by treatment of NaSH and DMR.

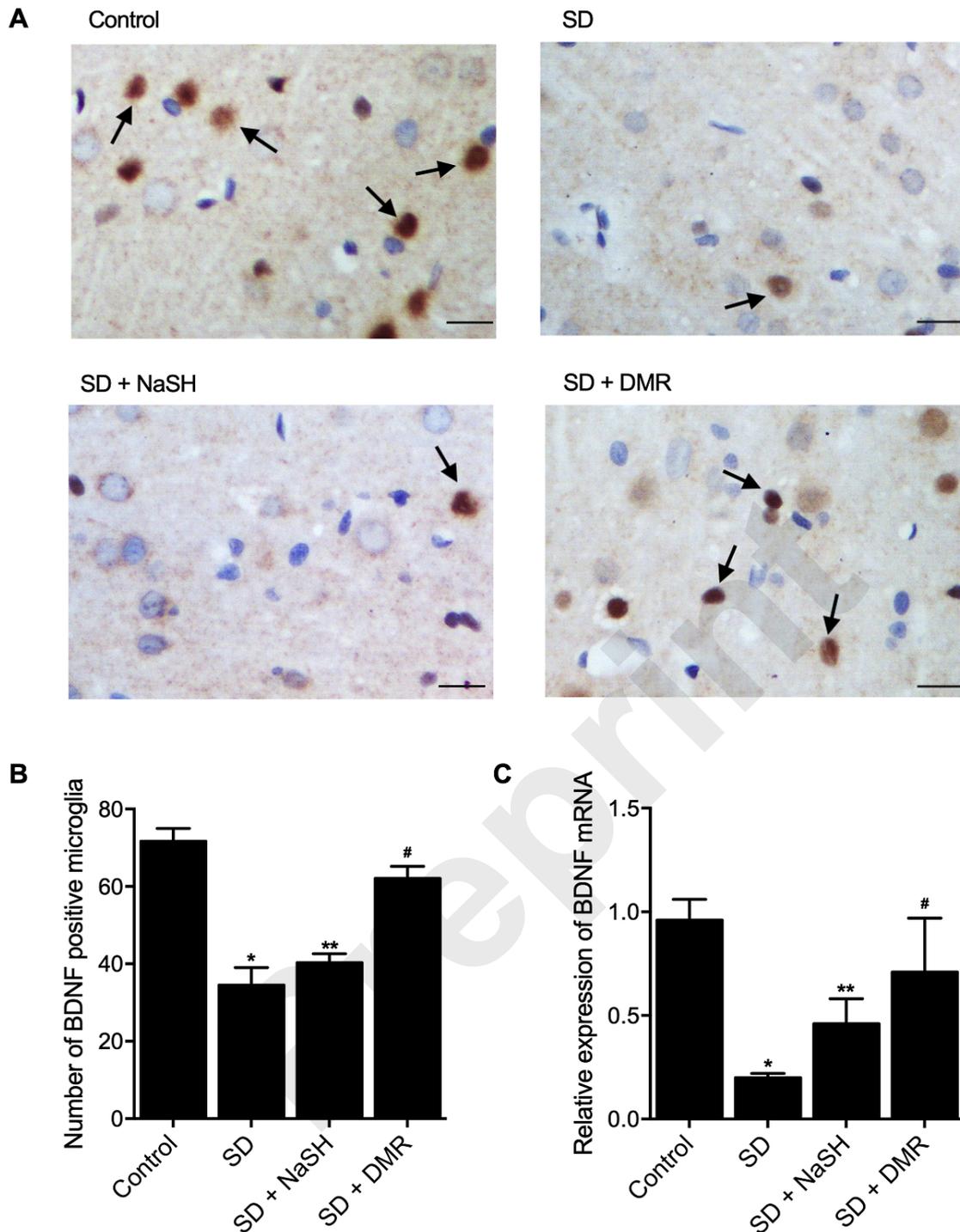


Figure 4

Promotive effect of DMR on the expression of hippocampal BDNF mRNA and protein (* P value < 0.05 vs. Control group; ** P value < 0.05 vs. SD group; # P value < 0.05 vs. SD+NaSH group; scale bar = 50 μ m).

A: IHC assay showed that hippocampal BDNF expression was reduced in sleep-deprived rats, and NaSH or DMR antagonized the changes of BDNF expression;

B: The reduced number of BDNF positive microglia in sleep-deprived rats was recovered by NaSH or DMR;

C: The relative expression level of BDNF mRNA was reduced in sleep-deprived rats, and NaSH or DMR antagonized the changes of BDNF expression.

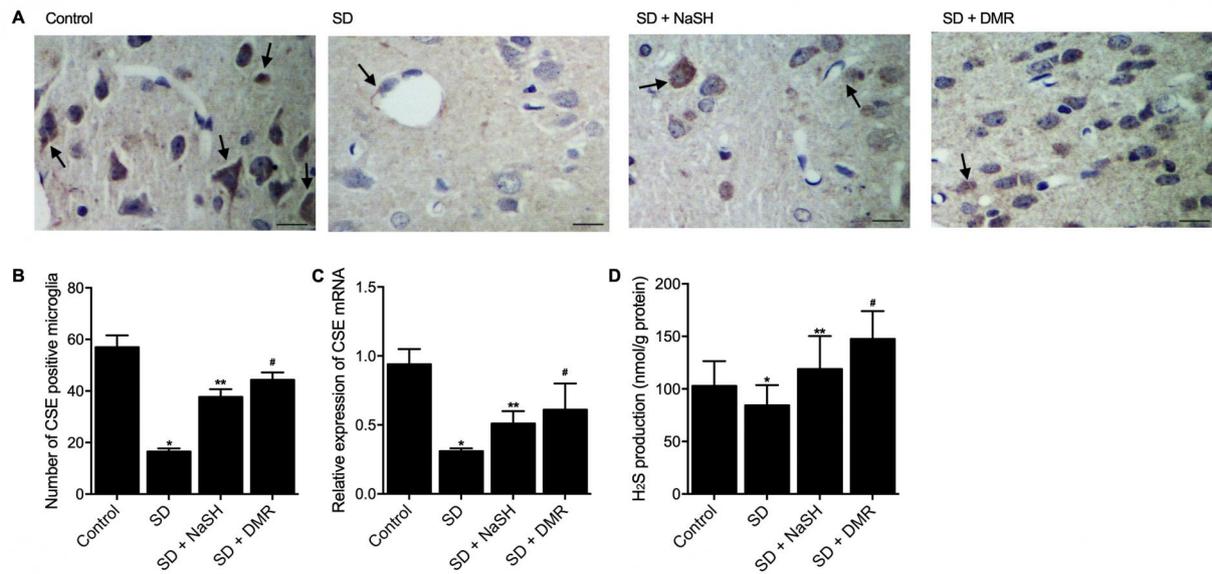


Figure 5

Promotive effect of DMR on the expression of hippocampal CSE mRNA and production of H₂S (* P value < 0.05 vs. Control group; ** P value < 0.05 vs. SD group; # P value < 0.05 vs. SD+NaSH group; scale bar = 50 μ m)

A: IHC assay showed that hippocampal CSE expression was reduced in sleep-deprived rats, and NaSH or DMR antagonized the changes of BDNF expression;

B: The reduced number of CSE positive microglia in sleep-deprived rats was recovered by NaSH or DMR;

C: The relative expression level of CSE mRNA was reduced in sleep-deprived rats, and NaSH or DMR antagonized the changes of BDNF expression;

D: H₂S production was lowest in sleep-deprived rats, and NaSH or DMR treatment antagonized the reduction of H₂S production.