Impact of Lipid-Lowering Drug Targets on Genetic Links with Diabetic Retinopathy

Keywords

Diabetic Retinopathy, Type 2 diabetes, drug target, Blood Lipid

Abstract

Introduction

Lipid metabolism is pivotal in diabetic retinopathy (DR) development. Nevertheless, the relationship between lipid-lowering drugs and the risk of DR remains controversial. This study utilized Mendelian randomization (MR) to investigate the potential effects of pharmacological targets for lowering lipid levels on DR and to clarify the causal link between blood lipid characteristics and DR.

Material and methods

The data comprised genetic variations related to lipid traits and genetic variations associated with lipidlowering drug targets obtained from the Global Lipid Consortium. Total DR, non-proliferative DR (NPDR), and proliferative DR (PDR) were sourced from the Finnish R9 database. Lipid-lowering drug targets were tested using inverse variance-weighted MR (IVW-MR) and statistics-based MR (SMR). Colocalization and mediation analysis were conducted to validate the results and explore potential mediating factors.

Results

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Conclusions

This Mendelian randomization study suggests that abnormalities in triglyceride (TG) levels serve as a pathogenic element in DR. Of the nine lipid-lowering drug targets assessed, HMGCR and APOB have emerged as potential promising targets for managing NPDR.

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22	(HbA1c) are critical factors that mediate the impact of HMGCR and APOB on DR risk.
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24	triglyceride (TG) levels serve as a pathogenic element in DR. Of the nine lipid-lowering
25	drug targets assessed, HMGCR and APOB have emerged as potential promising targets
26	for managing NPDR. These findings underscore the importance of controlling both
27	BMI and HbA1c levels to optimize outcomes in diabetic patients at risk for DR. The
28	therapeutic mechanisms of HMGCR and APOB in DR go beyond lipid-lowering alone,
29	and a multimodal lipid-lowering strategy should be selected early and comprehensively
30	to address the patient's medical conditions.

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31 Keywords: Type 2 diabetes, Blood Lipid, Diabetic Retinopathy, drug target

32 Introduction

33 Diabetes is increasingly recognized as a significant global public health challenge. The worldwide diabetic population has escalated to 529 million in 2021 and is anticipated 34 to rise to 1.31 billion by 2050 [1]. With the rising prevalence of diabetes mellitus and a 35 36 trend towards younger patient populations, the global incidence of diabetic retinopathy 37 (DR) has been on the rise. By 2045, it is anticipated that DR cases will increase to 160.5 38 million, impacting 44.82 million individuals with VTDR[2]. DR, a significant 39 microvascular complication, stands as a key contributor to vision impairment among patients. This condition stems from sustained damage to the retinal vasculature, 40 41 resulting in alterations such as hard exudates, cotton-wool spots, and vascular remodeling[3]. 42

43 Laser therapy, vitrectomy, anti-ceramide immunotherapy, and intravitreal injections represent available modalities for managing DR[4]. However, these interventions are 44 45 predominantly aimed at slowing down DR progression rather than providing a definitive cure. Despite the existence of treatment options, the screening rate for DR 46 47 remains below 50% due to various socio-environmental factors, including economic 48 and regional disparities. Consequently, many patients who do not receive timely and regular treatment face irreversible visual impairment[5]. Therefore, the identification 49 and management of DR risk factors are paramount. 50

There are numerous factors that influence the risk of developing DR[6]. Notably, lipid 51 52 profiles within the diabetic cohort have garnered considerable global attention due to 53 their distinct correlation with various medical conditions. Hyperlipidemia is a systemic 54 metabolic disorder[7]. Dyslipidemia escalates the susceptibility to macrovascular complications such as peripheral vascular disease, coronary heart disease, and 55 cerebrovascular disease, as well as microvascular issues like retinopathy and end-stage 56 renal disease in patients with diabetes[8, 9]. Research into the relationship between 57 lipids and DR has evolved significantly over the past decades. Initial investigations 58 have indicated that patients with DR exhibit elevated baseline lipid concentrations 59 60 compared to the broader diabetic population[10]. A growing body of research suggests that this association may be due to the involvement of multiple lipid components. 61 62 Consequently, early initiation of lipid-lowering medications not only reduce the incidence of other complications, but also significantly reduce mortality. 63

64 A diverse array of lipid-lowering medications are presently accessible for managing

65	dyslipidemia. The evolution of therapeutic approaches has seen significant
66	advancement, from the introduction of first-generation statins to the development of
67	more targeted therapies. Guidelines, including those from the European Atherosclerosis
68	Society (EAS), advocate for statins as the primary therapeutic option for addressing
69	dyslipidemia in diabetic patients[11]. In addition, the combination of ezetimibe, PCSK9
70	inhibitors, and fibrates is frequently employed to augment lipid-lowering
71	interventions[11, 12]. However, the evidence concerning the influence of commonly
72	used lipid-lowering medications on the initiation and advancement of DR remains
73	contentious. Among the studies of fenofibrate drugs, the well-known Fenofibrate
74	Intervention and Event Lowering in Diabetes (FIELD) study and the Action to Control
75	Cardiovascular Risk in Diabetes (ACCORD) study have shown that fenofibrate appears
76	to slow the progression of diabetic retinopathy. However, these studies also revealed
77	differences in the drug's efficacy in patients with different subtypes and severity of
78	DR[13][14]. The role of statins in DR management has been similarly debated over
79	time. Early small-scale studies and extensive observational studies spanning the past
80	two decades have indicated potential advantages of statins in mitigating late-stage
81	complications of DR and averting vision impairment[15-17]. These findings seemed
82	promising until a recent study suggests that statin usage might elevate the prevalence
83	of DR in both proliferative and non-proliferative diabetic retinopathy (NPDR and PDR)
84	subgroups, as well as in the broader DR population[18]. These conflicting outcomes
85	underscore the critical need for additional comprehensive research to delineate the
86	precise impact of lipid-lowering drugs on DR.

87 To overcome the limitations of observational studies, the use of MR methods leveraging comprehensive summary data from genome-wide association studies (GWAS) is 88 89 gaining traction. MR harnesses genetic variations as inherent experiments to provide insights into potential causal relationships between risk factors and diseases, with 90 91 reduced susceptibility to environmental confounders or reverse causation[19]. In drug 92 target MR analysis, the simulation involves the pharmacological blockade of genetic 93 drug targets utilizing pertinent genetic variants as instrumental variables, encompassing quantitative trait loci for expression (eQTLs) and protein (pQTLs). This approach is 94 employed to assess the consequences of drug exposure[20]. Using a drug-target MR 95 approach, this study simulates exposure to lipid-lowering drugs in diabetic patients to 96 elucidate the causal relationship between these drugs and DR and to provide a basis for 97 98 clinical strategies for the prevention and treatment of DR.

99 Methods

100 Study Design

The recommendations for MR (STROBE-MR), which strengthens the reporting of observational studies in epidemiology, were followed in this investigation (see Table S1)[21]. Mendelian Randomization (SMR) utilizing summary data and two-sample MR methods was employed to explore the relationship between diabetic retinopathy (DR) risk and targets of lipid-lowering medications. All data utilized in this investigation were sourced from published, publicly available summary statistics, as detailed in Table 107 S2. Approval from the respective ethics committees was obtained for all original studies.

108 The study design workflow is depicted in Figure 1.

109 Data Sources and Selection of Genetic Instrumental Variables

110 Lipid Biomarkers

Genetic association data for lipid biomarkers were obtained from the Global Lipids 111 112 Genetics Consortium, which represents the largest genome-wide association study (GWAS) meta-analysis to date, encompassing approximately 1.5 million people of 113 European heritage[22]. The primary biomarkers considered were triglycerides (TG), 114 total cholesterol (TC), and low-density lipoprotein cholesterol (LDL-C), demonstrating 115 significance ($p < 5 \times 10^{8}$). These biomarkers met the criteria of a physical distance 116 requirement of 10,000 kb and a chain unbalance [LD] aggregation threshold of $r_2 < r_2$ 117 0.001. To address potential sample overlap bias, particularly given that the outcome 118 119 variables were derived from a Finnish database, participants from the Finnish Biobank (n = 177,987) were excluded from the dataset. 120

121 Lipid-Lowering Drug Targets

Utilizing information on both established and emerging lipid-lowering drugs [16, 23], we identified pertinent drug target genes using the DrugBank database. Subsequently, we conducted a comprehensive analysis integrating insights from existing literature and research findings [24, 25]. The study ultimately encompassed a total of 7 lipid-lowering drugs corresponding to 9 targets. Statins function by inhibiting 3-hydroxy-3-

127	methylglutaryl-CoA reductase (HMG-CoA reductase), resulting in the upregulation of
128	hepatic low-density lipoprotein receptors (LDL receptors). This mechanism enhances
129	the efficiency of LDL clearance. On the other hand, Ezetimibe operates by inhibiting
130	the Niemann-Pick C1-Like 1 (NPC1L1) gene, which is accountable for cholesterol
131	absorption in the intestine and liver, significantly reducing plasma total cholesterol and
132	LDL-C levels. Proprotein convertase subtilisin/kexin type 9 (PCSK9) inhibitors,
133	function by impeding the interaction of PCSK9 with LDL receptors (LDLR). This
134	action prevents the degradation of LDL receptors by PCSK9, leading to an increase in
135	the quantity of LDL receptors on the liver surface and a pronounced reduction in plasma
136	LDL-C levels. Bile acid sequestrants operate by binding bile acids within the intestine,
137	impeding their reabsorption and consequently lowering LDL-C levels. Mipomersen
138	acts to diminish the synthesis of apolipoprotein B-100 (APOB-100), resulting in
139	decreased levels of very low-density lipoprotein (VLDL), LDL, and lipoprotein(a)[9].
140	Fibrates specifically target peroxisome proliferator-activated receptor α (PPAR α),
141	enhancing the activity of lipoprotein lipase, thereby significantly reducing plasma
142	triglyceride (TG) levels[26] . Angiopoietin-like 3 (ANGPTL3) function by suppressing
143	ANGPTL3 protein, thereby enhancing the activity of lipoprotein lipase. Antisense
144	oligonucleotides directed at APOC3 mRNA serve to inhibit APOC3 synthesis, resulting
145	in a notable decrease in plasma TG levels[27]. Additionally, lipoprotein lipase (LPL)
146	mainly functions on the surface of capillary endothelial cells, catalyzing the hydrolysis
147	of triglycerides within circulating triglyceride-rich lipoproteins like chylomicrons and
148	VLDL. Predicated on the fundamental pharmacological functions of these target genes,

we subsequently categorized them into genes targeting the reduction of LDL-C and TG
levels. Detailed information is summarized in Table 1.

151 A systematic methodology was employed to ascertain pertinent genetic variants, drawing upon established methodologies from previous studies[28]. Our selection 152153process involves two main steps. Firstly, we initially identified variants that achieved 154 genome-wide significance ($p < 5 \times 10^{-8}$) within a 100 kb vicinity of the gene under investigation. This threshold is widely accepted in GWAS as indicative of strong 155evidence for association. To ensure independence among the selected variants, we 156 157 further refined our selection using LD-based clumping. We applied an LD threshold of $r^2 < 0.1$. This step helps to mitigate the risk of including multiple correlated variants 158 that could introduce biased estimates in subsequent analyses. 159

160 Genetic Associations with Diabetic Retinopathy

Genetic association data for three outcomes were selected from the FinnGen R9 release 161 GWAS summary statistics. The primary outcome was total DR, with secondary 162 163 outcomes including NPDR and PDR. These outcomes were diagnosed according to the International Classification of Diseases, 10th Revision (ICD-10) classification system. 164 Specifically, DR diagnosis primarily relied on the H36.0 code, with supplementary 165 codes within the H35 category utilized for specific classifications. Specifically, H35.0 166 was used for NPDR diagnosis, while H35.2 was commonly used for PDR diagnosis. 167 The sample sizes were as follows: DR included 10,413 cases and 308,633 controls; 168 NPDR included 3,494 cases and 366,864 controls; PDR included 9,511 cases and 169

170 **362,581** controls.

171 Statistical Analysis

172To investigate the causal relationship between cholesterol-lowering medications and genetically instrumented circulating lipid traits in relation to DR, NPDR, and PDR, we 173used two-sample MR analysis. To represent the impact of a 1 mmol/L shift in lipid 174 175 levels, all estimates (given as odds ratios [ORs]) were normalized to be 38.7 mg/dL for LDL-C, 88.5 mg/dL for TG, and 38.7 mg/dL for TC. 176 177 For drug target genes showing positive associations with outcome variables in the twosample analysis, we utilized the GTEx database to examine their expression in high-178expression tissues. Subsequently, we conducted SMR analysis to evaluate the 179 180 association between a 1-standard deviation (1-SD) change in drug target gene expression levels and outcome variables. 181 We utilized the Bonferroni adjustment for multiple testing, setting significance 182 183 thresholds at p < 0.006 (0.05/9) for the nine pharmacological targets and p < 0.016(0.05/3) for the three lipid characteristics. For all remaining analyses, statistical 184 185 significance was defined as a two-sided p-value < 0.05. The statistical analyses were

conducted using the R software (version 4.3.1) and involved the "TwoSampleMR",
"MendelianRandomization" and "coloc" packages.

The MR method is underpinned by three core assumptions[29]: exclusion limitation, independence, and relevance (Figure 2). Avoiding bias resulting from inadequate instrumental variables, we verified the strength of drug target instrumental variables by

calculating F-statistics (β^2 / SE²), with F > 10 indicating sufficient instrument 191 strength[30, 31]. Considering the well-documented advantages of lipid-lowering 192 medication therapy in lowering the risk of coronary artery disease (CAD), we 193 designated CAD as a positive control to confirm the efficacy of pertinent instrumental 194 variables. Genetic association data for CAD were derived from a genome-wide 195 196 association study involving 361,194 controls and 42,096 clinically diagnosed patients. To exclude bias from confounding factors beyond the study exposure, we conducted 197 Bayesian colocalization analysis for drug targets significantly associated with outcome 198 199 variables. Bayesian colocalization, founded on Bayes' theorem, serves as a tool to evaluate whether distinct molecules (e.g., proteins, RNAs) are situated in close 200 proximity in a cell, with the central idea being to combine prior knowledge with 201 202 observed data to derive posterior probabilities of colocalization[32]. This analysis assessed whether drug targets and DR-related SNPs were driven by the same causal 203 variant (posterior probability PP.H4) or influenced by different but linkage 204 disequilibrium-related causal variants (PP.H3)[32]. A posterior probability exceeding 205 0.80 was considered indicative of support for the colocalization hypothesis. 206 To explore the specific pathways through which positive drug targets affect DR, We 207

10 explore the specific pathways through which positive drug targets affect DR, we
assessed the connection between recognized risk factors (e.g., age at diabetes diagnosis,
fasting blood glucose, glycated hemoglobin (HbA1c), diabetic nephropathy, hypertension,
body mass index (BMI)) for DR and genetically proxied lipid-lowering treatments[33].
Subsequently, after considering mediating effects, we evaluated these effects using a
two-step MR approach. This approach enabled the quantification of the direct impact

of genetically linked lipid-lowering medications on DR, the evaluation of the indirect influence of the mediator via the product of coefficients method, and the determination of the standard error of the indirect effect using the Delta method. To confirm the strength and reliability of the findings, we conducted a test of heterogeneity (Cochran's Q test) and a test of multiple validity (MR-Egger regression intercept test).

218 **Results**

219 Traits of Lipids and DR Risk

As instrumental factors for lipid characteristics, we found separate SNPs linked to TG, TC, and LDL-C (Tables S3-S5). Genetically proxied increases in TG levels were linked to a higher risk of DR in the general population, according to a two-sample MR analysis (OR = 1.34; 95% CI: 1.20-1.50; p = 1.25×10^{-7}). However, no significant associations were observed for LDL-C and TC with DR or its subtypes in the overall population (Table 2 and S7).

We performed genetic simulations for nine lipid-lowering drug targets (Table S8). A positive control analysis was carried out to validate the effectiveness of the genetic instruments, revealing that eight genetically proxied pharmacological targets (except ANGPTL3) decreased CAD risk (Figure 2). The genetic instruments' F-values varied from 10 to 5810, indicating that the potential influence of instrumental variable bias on the study outcomes was unlikely (Table S6).

Figure 3 illustrates the effects of genetically proxied lipid-lowering drugs on DR. Our

analysis revealed nuanced and differential effects across various drug targets. HMGCR

234	targets showed the most consistent and significant protective effects with DR subtypes.
235	In the DR (OR = 0.62; 95% CI: 0.46-0.83; $p = 0.01$] and NPDR OR = 0.49; 95% CI:
236	0.34-0.70; p = 9.70 \times 10^-3] populations, genetically-modelled HMGCR
237	augmentation and a 1 mmol/L (88.9 mg/dL) rise in TG were associated with a lower
238	risk of DR and NPDR. APOB targets showed consistent risk reduction across DR
239	subtypes. Genetic simulation of APOB enhancement also showed associations with low
240	DR risk (DR: OR = 0.75; 95% CI: 0.60–0.94; p = 0.01; NPDR: OR = 0.64; 95% CI:
241	0.48–0.87; p = 4.30 × 10 ⁻³ ; PDR: OR = 0.82; 95% CI: 0.69–0.99; p = 0.03). Notably,
242	the protective effect was most pronounced in the NPDR subgroup. Conversely,
243	genetic simulation of ANGPTL3 enhancement was observed to increase the risk of DR
244	and PDR. However, the significance of the associations of ANGPTL3 and APOB with
245	outcomes diminished post-Bonferroni correction, warranting cautious interpretation.
246	Notably, no significant relationships were identified between the other genetically
247	simulated drug targets and DR outcomes.

The outcomes obtained from alternative analysis approaches were largely consistent with those from the primary analysis method (inverse variance-weighted method) (Table S8). No indications of pleiotropy were detected for the variables, except in the analyses involving LDL-C and TG, where the MR-Egger intercepts exceeded 0, thereby enhancing the validity of causal inference.

253 Gene Expression and DR Risk

Blood, liver, and subcutaneous adipose tissues exhibiting the highest expression levels 254of HMGCR, APOB, and ANGPTL3 genes were selected for SMR analysis using the 255 GTEx database. The results revealed a significant correlation between a 1-standard 256 deviation (1-SD) rise in HMGCR expression in blood tissue with a lower incidence of 257 DR (DR: OR = 0.66; 95% CI:0.52–0.85; p = 7.31× 10^-4; NPDR: OR = 0.64; 95% CI: 258 0.44–0.93; $p = 2.03 \times 10^{-2}$) (Table S10). No significant associations were identified 259between genes related to APOB or ANGPTL3 and DR or NPDR. Colocalization 260 analysis was conducted to determine the likelihood of shared causal SNPs between 261 genetic variants linked to HMGCR expression in whole blood tissue and DR/NPDR. 262 263 The results revealed a common causal variant for HMGCR expression in whole blood tissue and DR (PP.H4 = 0.85) (Table S11), but no strong evidence for a shared causal 264 265 variant with NPDR (PP.H4 = 0.27).

266 Mediation Analysis

To explore mediating factors in HMGCR's influence on DR risk, we performed a twostep MR analysis of six potential mediating variables to assess their role in the effects of HMGCR and APOB on DR. The results showed significant causal associations between HMGCR and HbA1c, BMI, and hypertension, whereas APOB was causally associated with HbA1c and BMI (Table S12). For HMGCR, we found that HbA1c mediated 9.92% (95% CI: 3.72%, 17.09%) of the total effect of HMGCR on NPDR and 20.33% (95% CI: 12.68%, 29.38%) of the total effect of HMGCR on DR. The 274 mediating effect of BMI was more significant, accounting for 17.26% (95% CI: 9.37%, 26.60%) of the total effect of HMGCR on NPDR and 36.83% (95% CI: 24.95%, 275 276 49.91%) of the total effect on DR (Figure 4). The mediating effect of hypertension, although statistically significant, was relatively small, accounting for only 2.34% (95% 277 278 CI: 0.14%, 5.65%) of the total effect of HMGCR on NPDR and 3.82% (95% CI: 0.40%, 279 8.25%) of the total effect on DR. For APOB, the mediating effect of HbA1c in its total effect on NPDR was 11.73% (95% CI: 4.06%, 21.21%), while the mediating effect of 280 BMI was 5.64% (95% CI: 2.48%, 9.70%). These results suggest that the genetically 281 282 modelled effects of HMGCR and APOB on reducing the risk of DR are partly mediated by these mediators. In particular, BMI and HbA1c played a large mediating role in the 283 effect of HMGCR on DR, whereas the mediating role of HbA1c was more significant 284 285 in the effect of APOB on NPDR. Detailed statistical results are presented in Tables S12, S13 and S14 of the Supplementary Material. 286

287 **Discussion**

This study examined the possible effects of lipid-lowering treatment targets while utilizing MR and drug target SMR methodologies to explore the causal link between blood lipid levels and DR[34]. The key findings of our study are as follows: elevated TG levels significantly increase the risk of DR and NPDR; HMGCR is negatively correlated with DR risk in the total population and NPDR; APOB is also negatively correlated with NPDR risk; Nevertheless, there is no proof that lipid characteristics or the nine pharmacological targets that lipid-lowering have an effect on PDR. Mediation analyses highlight the importance of glycaemic control and weight management in the
prevention and management of DR. Although hypertension is a known risk factor for
DR, the role of HMGCR-related lipid-lowering drugs in DR appears to be less
prominent than that of BMI and HbA1c.

The correlation between DR risk and blood lipid levels remains controversial. Previous 299 300 studies have reported conflicting results, with some investigations showing no clear 301 relationship between any blood lipid component and any form of DR[35, 36]. For instance, a MR study conducted by Sobrin et al. did not identify any causal effect of 302 four lipid components (HDL, LDL, TG, TC) on DR [33]. However, a study by Dornan 303 304 et al. suggested that LDL levels were higher in PDR populations compared to NPDR and normal populations, implying a correlation between LDL and DR severity [37]. 305 Furthermore, a large Spanish follow-up study reported that LDL levels and the TC to 306 LDL ratio were associated with an increased risk of DR[34, 38]. These findings align 307 308 with a meta-analysis of 13 cohort studies, which indicated that baseline TG levels were linked to the development of diabetic retinopathy in individuals with diabetes, while no 309 310 significant associations were found between the prevalence of diabetic retinopathy and 311 LDL and TC levels[39]. This inconsistency reflects the complex relationship between lipids and DR, possibly related to methodological differences, population 312 characteristics, or different stages of DR. 313

In the current study, genetic simulation of HMGCR and APOB was linked to a decreased risk of any DR and NPDR, which is consistent with the findings of Chen et 316 al. However, this protective effect does not seem to extend to PDR. Additionally, through mediation analyses, we demonstrated that the protective impact of HMGCR on 317 318 DR was partially mediated by HbA1c and BMI, with the causal effect of HMGCR on 319 these two mediating variables resembling previous findings [40, 41]. An animal model-320 based investigation revealed that reduced Hmgcr expression led to elevated dietary 321 intake and fat storage, potentially mediated by the target of brain insulin (TOBI) regulation through modulation of a-glucosidase gene expression to regulate blood 322 glucose levels[42]. This discovery implies that genetic mimicry of HMGCR to promote 323 324 blood glucose and BMI reduction may be linked to this phenomenon. The study further confirmed the association between HMGCR and BMI by observing weight gain 325 resulting from statin drug administration[43].Chronic hyperglycaemia induces 326 327 oxidative stress by increasing reactive oxygen species (ROS) production, triggering an inflammatory cascade resulting in vascular wall damage and increased vascular 328 permeability, activation of protein kinase C and advanced glycation end-products 329 330 (AGEs), and endothelial dysfunction. In addition, sustained high blood glucose levels lead to pericyte detachment and basement membrane thickening, impaired 331 332 neurovascular coupling, and disruption of retinal microcirculation. These series of metabolic and physiological disturbances ultimately induce the development of DR. 333

Additionally, genetic mimicry of APOB linked to lowering LDL-C was identified as contributing to the reduction in NPDR risk. It is noteworthy that the impact of apolipoprotein B (APOB) on DR in previous investigations was multifaceted, with some studies indicating that APOB increased retinal small artery tortuosity, a factor correlated with the severity of DR[44, 45]. However, Li et al. did not find an association
between genetically determined APOB and DR risk. This inconsistency may prompt us
to revisit the relationship between APOB and DR. It is possible that genetic variations
could influence the function of APOB beyond its levels, and specific instrumental
variables related to the APOB gene might exert protective effects. Furthermore, the
complex interplay of genetic background, environmental factors, and population
variances may all contribute to the diverse outcomes observed in this association.

Previous large clinical studies such as FIELD and ACCORD-EYE have demonstrated 345 the protective effect of betablockers in patients with diabetic retinopathy (DR)[46]. 346 347 However, in the present study, only inhibition of the ANGPTL3 gene showed a trend towards reducing the risk of DR. This discrepancy may stem from several aspects: first, 348 349 fibrates such as fenofibrate exert their triglyceride (TG) lowering effects by activating multiple signaling pathways, rather than relying on a single pathway. This multi-350 351 targeted action may be an important reason for the superior therapeutic effect over single gene inhibition observed in clinical trials. Second, although studies have 352 353 confirmed the correlation between certain circulating lipid levels and DR progression, 354 the final clinical phenotype is modulated by multiple factors. In addition to changes at the transcriptional and translational levels of genes, environmental factors, epigenetic 355 modifications, and other unknown regulatory mechanisms may influence disease 356 357 progression and therapeutic efficacy. This complex interaction may lead to differences 358 in the effects of single gene interventions versus the results of drug therapy.

359 These seemingly contradictory findings suggest that lipid-lowering drugs may have 360 different mechanisms and effects in the prevention and treatment of DR. It was found 361 that microglia aggregation and systemic inflammation were more severe in patients 362 with PDR, whereas fenofibrate ameliorated oxidative stress and systemic inflammation, 363 while also inhibiting infiltration and activation of retinal cells[47]. Previous studies 364 have shown that the degree of systemic inflammation in patients with DR is related to the grade of the disease, and the differences in the efficacy of different types of lipid-365 lowering drugs in patients with different DR grades may be related to the above factors. 366 367 Therefore, it is important to explore the mechanism of action of these drugs in different types of DR, especially the relationship with inflammatory reactions and cell activation. 368

The pathological process of DR is complex, with evidence indicating that plasma LDL-369 370 C and cholesterol levels are linked to retinal hard exudates [48, 49]. Statin medications 371 have been shown by Gupta et al. to lessen the intensity of hard exudates and central 372 foveal lipid migration [18, 50], further supporting the association between blood lipids and retinal exudation. The intricate relationship between circulating lipids and DR has 373 374 been confounded by numerous factors, prompting an increasing number of studies to 375 explore and emphasize non-lipid mechanisms. Recent research has indicated that disruption of retinal cholesterol metabolism and impaired retinal capillary repair may 376 377 serve as the underlying mechanisms of DR. Initially, retinal cholesterol accumulation leads to the formation of highly reflective crystalline deposits (CCS), which activate 378 the immune response, triggering the NLRP3 inflammasome and the release of various 379 380 inflammatory factors, such as IL-1, thereby inciting local tissue inflammation [47].

Various microbial infections activate Toll-like receptors (TLRs), inhibit liver X receptors (LXRs) through the viral response to the transcription factor interferon regulatory factor 3 (IRF3), reduce expression of ABCA1 transporter proteins, and inhibit cholesterol efflux by macrophages [51]. In addition, chronic inflammatory activation in diabetic patients can disrupt bone marrow microenvironmental homeostasis and slow retinal vascular endothelial cell repair, thus exacerbating the progression of DR [52].

388 This study leverages the advantages of large-scale samples from the Finnish Biobank, providing important insights into the genetic risk of DR. This study demonstrates the 389 390 potential for more targeted and individualized interventions beyond the current "onesize-fits-all" approach. The research methods employed help to avoid the reverse 391 causality issues and confounding variables that are prevalent in traditional 392 observational studies. From a clinical application perspective, our findings provide new 393 opportunities for precision medicine. It is worth noting that although the study found 394 that genetic mimicry of HMGCR and APOB enhancement were beneficial in reducing 395 396 the risk of DR, we acknowledge the therapeutic benefits of lipid-lowering medications 397 like statins and mipomersen in individuals with diabetes. Our findings offer a nuanced approach to clinical implementation for patients with DR and hyperlipidemia. A 398 comprehensive assessment of the patient's genetic and metabolic profile is 399 recommended to develop a personalized lipid-lowering strategy for optimal 400 management based on the patient's individual profile. Future research endeavors may 401

focus on identifying the most suitable lipid-lowering approach based on the unique
characteristics of patients (e.g., DR stage, genotype).

404 However, we also acknowledge the limitations of the methodology of this study: (1) 405 Despite conducting sensitivity analyses, studies based on GWAS data still cannot completely rule out pleiotropy, there may be other confounding factors affecting 406 outcomes, and the results have the potential to be false-positive. (2) Given that the 407 408 GWAS data originated from European cohorts, caution is warranted in generalizing the findings to other ethnic populations, as population-specific effects may not be 409 accurately represented. (3) Although the study detailed specific retinal effects of lipid-410 411 lowering drugs in patients with DR, the GWAS-based data were limited, and it was not possible to obtain the eQTL data of HMGCR and APOB in retinal tissues, and the 412 413 results for the intraretinal effects. (4) The potential disparities between the direct effects of lipid-lowering therapies and the effects of genetic variants on DR risk were 414 415 not directly evaluated in this study. (5) the original GWAS data can only be used to make the main classification based on DR, but the progression of DR includes many 416 417 other important pathological processes, and refinement of the effects of lipid-lowering 418 therapies on these pathological processes will help to strengthen the results, thereby facilitating a more comprehensive interpretation of the results. 419

We acknowledge several critical limitations in our current study that necessitate future research. Firstly, the multiplicity of studies can be reduced by developing advanced statistical methods to more accurately distinguish between direct and indirect genetic 423 effects and by using a multi-omics approach to reveal complex genetic interactions. 424 Secondly, in order to generalise the results of the study, multi-ethnic studies can be 425 conducted at a later stage to incorporate different genetic backgrounds to ensure the 426 robustness of the risk assessment. Stratified analysis protocols could also be developed 427 to take into account genetic variation in specific populations.

428 Based on our findings, future research directions may include: 1) Further elucidating 429 the mechanisms of action of lipid-lowering drugs at different DR stages, especially nonlipid effects; 2) Develop comprehensive genetic screening protocols, explore, 430 individualized treatment strategies based on genotype risk prediction and treatment 431 432 selection and develop predictive models that integrate genetic, metabolic, and clinical variables. 3) Creating algorithm-based treatment selection models, considering the 433 interaction between DR and other diabetic complications and developing an integrated 434 435 management approach that considers multiple metabolic pathways; 4) Conduct longitudinal studies investigating long-term outcomes of lipid management in DR 436 populations, ongoing monitoring of DR progression, and validation of HMGCR and 437 438 APOB-targeted therapies. This will lead to a comprehensive assessment of potential 439 side effects and long-term efficacy.; 5) Combining genetic insights with clinical practice for personalised diabetic retinopathy management and developing precision medicine 440 441 approaches.

442 Conclusion

443 In conclusion, our research uses MR and SMR techniques to illuminate the intricate

connection between lipid metabolism and DR, particularly the potential protective role 444 of HMGCR and APOB in NPDR risk. The mediation analysis highlighted the 445 446 importance of glycemic control and weight management in the prevention and management of DR. These results contribute to our understanding of the pathogenic 447 448 underpinnings of DR and offer fresh perspectives for preventative and therapeutic 449 approaches in the future. While further study is required to confirm and elaborate on these findings, our work translates genetic insights into practical clinical applications, 450 ultimately improving patient outcomes and developing more precise, personalized 451 452 approaches to diabetic retinopathy management.

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The GWAS authors and participants are much appreciated by the authors for their contributions of summary statistics data.

456 **Potential Conflicts of Interest**

According to the study's authors, there were no business or financial relationships that
 may have created a conflict of interest throughout the study's implementation.

459 **Patient and public involvement**

460 Patients or the general public were not involved in the conception, methodology,

461 reporting, or distribution methods of the study.

462 **Patient consent for publication**

463 It's not applicable.

464 Data availability statement

- 465 All information is accessible to the general audience. Table S2 summarizes the specific
- 466 details for these datasets.

467

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Impact	Impact of genetic variants associated with lipid profiles and drug targets on DR assessed by Mendelian randomization (MR) analysis								
Lipid deposition damages the retina		s lipid deposition atherosclerosis	retinal damage						
Lipid traits	LDL TG TC	Genetic simulation HMGCR↓ Representative drugs(Rosuvastatin)	PDR ↑						
Positive drug targets	HMGCR and APOB	Genetic simulation APOB↓ Representative drugs(mipomersen sodium)	R↑						
interm (Medi	ediary variable iated proportion (%))	HbAlc \downarrow 20.33% 9.92% 17.13% HMGCR \uparrow 36.83% 17.26% 17.26% 17.26% 17.26% 17.13% 17.26% 17.26% 17.26% 17.13% 17.26% 17.26% 17.13% 17.26% 17.26% 17.13% 17.26% 17.26% 17.13% 17.26% 17.	APOB ↑						

Primary pharmacological Action	Drug targets	Target genes	Gene region (GRCh37)	Genetic instruments(nSNPs)
Reduced LDL-C	LDL Receptor	LDLR	chr19:11200139-11244496	50
	HMG-CoA reductase	HMGCR	Chr5:74632993-74657941	23
	Niemann-Pick C1-like protein 1	NPC1L1	Chr7:44552134-44580929	14
	Proprotein convertase subtilisin/kexin type 9	PCSK9	Chr1:55505221-55530525	43
	Apolipoprotein B-100	APOB	Chr2:21224301-21266945	24
Reduced TG	Lipoprotein Lipase	LPL	Chr8:19759228-19824769	34
	APOC3 mRNA	APOC3	Chr11:116700422-116703788	31
	Peroxisome proliferator-activated receptor α	PPARA	Chr22:46546424-46639653	9
	ANGPTL3 protein	ANGPTL3	Chr1:63063158-63071830	20

Table 1 Information of genetic instruments.

Abbreviation: SNPs, single-nucleotide polymorphisms; chr, chromosome; LDL-C, low-density lipoprotein cholesterol; TG, triglyceride.

Table 2 Risk association between blood lipids and DR.

F	Madha J	DR		NPDR		PDR	
Exposure	Method	OR(95% CI)	P Value	OR(95% CI)	P Value	OR(95% CI)	P Value
LDL	Inverse variance weighted	0.94(0.84,1.05)	0.28	0.94(0.79,1.10)	0.42	0.96(0.88,1.06)	0.46
	MR Egger	0.76(0.65,0.90)	0.001	0.78(0.61,1.00)	0.05	0.87(0.75,1.00)	0.05
	Weighted median	0.96(0.84,1.11)	0.60	0.96(0.78,1.19)	0.73	1.01(0.88,1.16)	0.91
	Weighted mode	0.91(0.80,1.04)	0.15	0.96(0.78,1.18)	0.69	1.03(0.89,1.18)	0.70
TG	Inverse variance weighted	1.34(1.20,1.50)	1.25E-07	1.13(1.00,1.28)	0.05	1.05(0.96,1.14)	0.29
	MR Egger	0.89(0.77,1.04)	0.15	0.88(0.74,1.06)	0.19	0.83(0.73,0.94)	0.003
	Weighted median	1.12(0.96,1.30)	0.16	1.01(0.80,1.26)	0.96	0.94(0.81,1.09)	0.40
	Weighted mode	1.05(0.91,1.21)	0.53	1.00(0.84,1.20)	0.96	0.90(0.80,1.02)	0.10
ТС	Inverse variance weighted	0.91(0.82,1.01)	0.09	0.90(0.78,1.04)	0.15	0.95(0.88,1.04)	0.27
	MR Egger	0.81(0.70,0.95)	0.009	0.80(0.65,0.99)	0.04	0.87(0.77,0.99)	0.03
	Weighted median	0.99(0.87,1.13)	0.87	0.92(0.76,1.12)	0.40	0.95(0.85,1.07)	0.43
	Weighted mode	0.95(0.83,1.08)	0.431	0.87(0.73,1.05)	0.14	0.94(0.83,1.07)	0.37

Abbreviation: LDL-C, low-density lipoprotein cholesterol; TG, triglyceride, TG, total cholesterol, NPDR, non-proliferative diabetic retinopathy; PDR, proliferative diabetic retinopathy.



Figure1: Study Design Flowchart



Figure2: Mendelian randomization principles and presumptions.

Drug Target	nSNP	P.value	DR(Total population)	OR(95%CI)	Drug Target	nSNP	P.value	PDR(Subgroup)	OR(95%CI)
PCSK9	43	6.10e-01		0.97(0.84, 1.11)	PCSK9	43	3.30e-01		0.94(0.83, 1.06)
LPL	34	1.70e-01		1.15(0.94, 1.41)	LPL	34	6.40e-01		1.04(0.88, 1.23)
PPARA	9	4.00e-02	⊢→	4.20(1.06, 16.60)	PPARA	9	6.20e-01		0.71(0.18, 2.77)
LDLR	50	5.60e-01		1.04(0.90, 1.20)	LDLR	50	8.70e-01		0.99(0.85, 1.15)
ANGPTL3	20	2.00e-02		1.39(1.05, 1.83)	ANGPTL3	20	2.00e-02		1.42(1.06, 1.90)
APOC3	31	9.10e-01		1.01(0.87, 1.18)	APOC3	31	6.70e-01		0.97(0.85, 1.11)
APOB	24	1.00e-02		0.75(0.60, 0.94)	APOB	24	3.00e-02	H-0	0.82(0.69, 0.99)
HMGCR	23	1.00e-03		0.62(0.46, 0.83)	HMGCR	23	3.30e-01		0.87(0.66, 1.15)
NPC1L1	14	4.00e-02	⊢−−−→	1.76(1.02, 3.05)	NPC1L1	14	1.00e-01		1.59(0.91, 2.77)
		0.	5 0.75 1 1.25 1.	5			0	.5 0.75 1 1.25 1	.5
Drug Target	nSNP	P.value	CAD(Positive Control)	OR(95%CI)	Drug Target	nSNP	P.value	NPDR(Subgroup)	OR(95%CI)
Drug Target PCSK9	nSNP 43	P.value 4.00e-33	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21)	Drug Target PCSK9	nSNP 43	P.value 1.30e-01	NPDR(Subgroup)	OR(95%CI) 1.17(0.96, 1.43)
Drug Target PCSK9 LPL	nSNP 43 34	P.value 4.00e-33 1.65e-19	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77)	Drug Target PCSK9 LPL	nSNP 43 34	P.value 1.30e-01 9.00e-02	NPDR(Subgroup)	OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04)
Drug Target PCSK9 LPL PPARA	nSNP 43 34 9	P.value 4.00e-33 1.65e-19 1.86e-02	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19)	Drug Target PCSK9 LPL PPARA	nSNP 43 34 9	P.value 1.30e-01 9.00e-02 2.10e-01	NPDR(Subgroup)	OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04) ► 8.06(0.30, 213.18)
Drug Target PCSK9 LPL PPARA LDLR	nSNP 43 34 9 50	P.value 4.00e-33 1.65e-19 1.86e-02 1.21e-20	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19) 1.91(1.67, 2.19)	Drug Target PCSK9 LPL PPARA LDLR	nSNP 43 34 9 50	P.value 1.30e-01 9.00e-02 2.10e-01 9.70e-01	NPDR(Subgroup)	OR(95%CI) 1.17(0.96,1.43) 0.78(0.58,1.04) → 8.06(0.30,213.18) 1.00(0.80,1.24)
Drug Target PCSK9 LPL PPARA LDLR ANGPTL3	nSNP 43 34 9 50 20	P.value 4.00e-33 1.65e-19 1.86e-02 1.21e-20 1.63e-01	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19) 1.91(1.67, 2.19) 1.17(0.94, 1.45)	Drug Target PCSK9 LPL PPARA LDLR ANGPTL3	nSNP 43 34 9 50 20	P.value 1.30e-01 9.00e-02 2.10e-01 9.70e-01 7.00e-02	NPDR(Subgroup)	 OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04) 8.06(0.30, 213.18) 1.00(0.80, 1.24) 1.42(0.98, 2.07)
Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3	nSNP 43 34 9 50 20 31	P.value 4.00e-33 1.65e-19 1.86e-02 1.21e-20 1.63e-01 7.25e-08	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19) 1.91(1.67, 2.19) 1.17(0.94, 1.45) 1.29(1.17, 1.41)	Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3	nSNP 43 34 9 50 20 31	P.value 1.30e-01 9.00e-02 2.10e-01 9.70e-01 7.00e-02 9.00e-02	NPDR(Subgroup)	 OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04) 8.06(0.30, 213.18) 1.00(0.80, 1.24) 1.42(0.98, 2.07) 1.19(0.97, 1.46)
Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3 APOB	nSNP 43 34 9 50 20 31 24	P.value 4.00e-33 1.65e-19 1.86e-02 1.21e-20 1.63e-01 7.25e-08 5.25e-06	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19) 1.91(1.67, 2.19) 1.17(0.94, 1.45) 1.29(1.17, 1.41) 1.54(1.28, 1.85)	Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3 APOB	nSNP 43 34 9 50 20 31 24	P.value 1.30e-01 9.00e-02 2.10e-01 9.70e-01 7.00e-02 9.00e-02 4.30e-03	NPDR(Subgroup)	 OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04) 8.06(0.30, 213.18) 1.00(0.80, 1.24) 1.42(0.98, 2.07) 1.19(0.97, 1.46) 0.64(0.48, 0.87)
Drug Targed PCSK9 LPL PPARA LDLR ANGPTL3 APOC3 APOB HMGCR	nSNP 43 34 9 50 20 31 24 23	P.value 4.00e-33 1.65e-19 1.86e-02 1.21e-20 1.63e-01 7.25e-08 5.25e-06 8.02e-03	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19) 1.91(1.67, 2.19) 1.17(0.94, 1.45) 1.29(1.17, 1.41) 1.54(1.28, 1.85) 1.39(1.09, 1.77)	Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3 APOB HMGCR	nSNP 43 34 9 50 20 31 24 23	P.value 1.30e-01 9.00e-02 2.10e-01 9.70e-01 7.00e-02 9.00e-02 4.30e-03 9.68e-05	NPDR(Subgroup)	 OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04) 8.06(0.30, 213.18) 1.00(0.80, 1.24) 1.42(0.98, 2.07) 1.19(0.97, 1.46) 0.64(0.48, 0.87) 0.49(0.34, 0.70)
Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3 APOB HMGCR NPC1L1	nSNP 43 34 9 50 20 31 24 23 14	P.value 4.00e-33 1.65e-19 1.86e-02 1.21e-20 1.63e-01 7.25e-08 5.25e-06 8.02e-03 2.94e-06	CAD(Positive Control)	OR(95%CI) 1.98(1.77, 2.21) 1.60(1.45, 1.77) 3.74(1.25, 11.19) 1.91(1.67, 2.19) 1.17(0.94, 1.45) 1.29(1.17, 1.41) 1.54(1.28, 1.85) 1.39(1.09, 1.77) 2.04(1.51, 2.75)	Drug Target PCSK9 LPL PPARA LDLR ANGPTL3 APOC3 APOB HMGCR NPC1L1	nSNP 43 34 9 50 20 31 24 23 14	P.value 1.30e-01 9.00e-02 2.10e-01 9.70e-01 7.00e-02 9.00e-02 4.30e-03 9.68e-05 4.00e-02	NPDR(Subgroup)	 OR(95%CI) 1.17(0.96, 1.43) 0.78(0.58, 1.04) 8.06(0.30, 213.18) 1.00(0.80, 1.24) 1.42(0.98, 2.07) 1.19(0.97, 1.46) 0.64(0.48, 0.87) 0.49(0.34, 0.70) 2.02(1.04, 3.91)

Figure3: Associations Between Genetically Proxied Lipid-Lowering Drugs and Diabetic Retinopathy



Figure4: Intermediary Analysis Chart