Exploring the mediating role of the plasma lipidome in the pathway from the gut microbiota to dementia: a Mendelian randomization study

Binghan Li¹, Xu Sun¹, Xiao Luo², Yuting Kan¹, Weisen Wang¹, Tianren Wang¹, Cheng Wu^{2*}, Yongbo Hu^{1*}, Xiaoying Bi^{1*}

¹Department of Neurology, Shanghai Changhai Hospital, Naval Medical University, Shanghai, China

²Department of Military Health Statistics, Naval Medical University, Shanghai, China

Submitted: 15 August 2024; Accepted: 15 February 2025 Online publication: 27 April 2025

Arch Med Sci DOI: https://doi.org/10.5114/aoms/201447 Copyright © 2025 Termedia & Banach

Abstract

Introduction: Previous studies have indicated a potential association between the gut microbiota (GM) and dementia; however, the exact causeand-effect relationships between GM, various types of dementia, and the potential influence of the plasma lipidome as intermediaries are still unclear. **Material and methods:** We used genome-wide association study (GWAS) data to identify GM taxa, plasma lipid species (lipidome), and five types of dementia: Alzheimer's disease (AD), dementia with Lewy bodies (DLB), Parkinson's disease dementia (PDD), frontotemporal dementia (FTD) and vascular dementia (VD). We used Mendelian randomization (MR) to investigate the possible causal connections among the GM, plasma lipidome, and dementias. The inverse variance weighting (IVW) method served as the primary statistical approach. We investigated the role of plasma lipidome as a potential mediating factor in this relationship.

Results: A total of 41 positive and 39 negative causal relationships between genetic susceptibility in the GM taxa or bacterial pathways and dementia, as well as 14 negative causal relationships between the plasma lipidome and dementias, were identified. Additionally, only 1 potential mediation pathway was identified as having a significant mediating effect.

Conclusions: Our results suggest a link between the GM and the plasma lipidome with five distinct types of dementia, indicating that the phosphatidylcholine (O-16:1_18:2) level could play a role in the pathway from the species *Bacteroides coprocola* to vascular dementia.

Key words: dementia, plasma lipidome, gut microbiota, Mendelian randomization.

Introduction

Dementia is a prevalent neurodegenerative disorder distinguished by cognitive dysfunction and gradual deterioration in daily functioning [1]. The World Health Organization has recognized dementia as the seventh most common cause of death globally. As the global population ages, the current estimate of over 50 million individuals affected by dementia is projected to increase to 152 million by the year 2050 [2]. Dementia is a multifaceted neurological syndrome characterized by deficits in

*Corresponding authors:

Yongbo Hu Xiaoying Bi Department of Neurology Shanghai Changhai Hospital Naval Medical University 168 Changhai Road Yangpu District 200433, Shanghai China E-mail: huyongbo91@126.com, bixiaoying2013@163.com

Cheng Wu

Department of Military Health Statistics Naval Medical University 800 Xiangyin Road Yangpu District 200433, Shanghai, China E-mail: wucheng_wu@126.com



Attribution-NonCommercial-ShareAlike 4.0 International (CC BY -NC -SA 4.0). License (http://creativecommons.org/licenses/by-nc-sa/4.0/)

Creative Commons licenses: This is an Open Access article distributed under the terms of the Creative Commons

cognition and memory, with specific subtypes including Alzheimer's disease (AD), dementia with Lewy bodies (DLB), Parkinson's disease dementia (PDD), frontotemporal dementia (FTD), and vascular dementia (VD) [3, 4]. The high occurrence of dementia presents considerable obstacles in healthcare, financial resources, and caregiver burden [5]. Thus, the identification of risk factors and biomarkers is essential for the prevention and management of dementia.

The human gut microbiota (GM) consists of 100 trillion microorganisms with over 3 million genes that impact human physiology, health, and behavior [6]. Recent studies have revealed that the GM is important for the nervous system and is connected to neurodegenerative diseases through the microbiota-gut-brain axis (MGBA) [7]. The dysregulation of the MGBA may induce neuroinflammation, lipid metabolic disorder, and synaptic impairment, and subsequently cause cognitive decline [8]. Moreover, the GM offers great potential as a reservoir for new therapeutic opportunities, enabling treatment of numerous neurological disorders by targeting the MGBA [9].

Plasma lipids, commonly assessed through high/low-density lipoprotein cholesterol, triglycerides, and total cholesterol, have been identified as significant factors associated with dementia in numerous studies [10, 11]. Nevertheless, advancements in lipidomics technologies have greatly expanded our comprehension of the diversity and breadth of circulating lipids. Lipid species such as phosphatidylcholine, sterol ester, ceramide and phosphatidylethanolamine have the potential to enhance dementia risk evaluation beyond traditional lipid measures [12, 13].

Lipids play a vital role in cellular function by contributing to membrane structure, intercellular communication, energy storage, and homeostasis regulation [14]. Neurodegenerative diseases and other neurological disorders have been linked to dysregulation of brain lipids [15]. Recent studies have suggested that the GM may influence lipid profiles and lipidomics in dementia, implicating both the GM and lipidome in dementia pathogenesis [16]. It is hypothesized that the lipidome could serve as a mediator in the pathway linking GM to the development of dementia. MR was used in this study to investigate causal relationships between exposure variables and outcomes using single nucleotide polymorphisms (SNPs) as instrumental variables (IVs). The two-sample MR technique enhances statistical power by leveraging published summary estimates from diverse genome-wide association study (GWAS) to identify causal effects between exposure variables and outcomes [17].

This study hypothesized that certain types of lipids may mediate the associations between the gut microbiota and the development of different types of dementia. Therefore, MR analysis was employed to explore the links between the GM, plasma lipidome, and different types of dementia. It also investigated the potential role of the plasma lipidome in the pathway from the GM to dementia, and analyzed the impact of genetic predisposition to dementia risk on the GM and plasma lipidome.

Material and methods

Study design

This study comprises three primary components: an examination of the influence of GM on dementia, an investigation into the impact of plasma lipids on dementia, and an analysis of the role of plasma lipids in the pathway from GM to dementia. The study used SNP as IVs and followed the fundamental assumptions of MR [18]. The MR study was reported according to the MR-STROBE guidelines (Figure 1).

Data source

The data on the GM are derived from Esteban *et al.*'s study, which reported 412 microbes [19]. The genetic data for the plasma lipidome came from a GWAS with 7174 individuals and 179 lipid species [20].

Data on VD, AD, PDD, FTD, and DLB were collected from FinnGen's tenth version (https://r10. risteys.finngen.fi/) and Chia's study [21]. Patients were screened using ICD diagnosis codes for dementia subtypes and genetic data were downloaded from the FinnGen database. DLB data were included in the IEU Open GWAS (https://gwas.mrcieu.ac.uk/) database based on Chia's study. Participants were diagnosed using consensus criteria.

GWAS summary statistics were used as secondary data in the study, following ethical guidelines. Ethical approval was obtained for the original studies, and the results can be accessed on the website provided. The data are publicly available and do not require further ethical review.

Selection of instrumental variables

The genetic instruments met the following criteria: (1) we selected the SNPs with a *p*-value of 1×10^{-5} for GM as the threshold; (2) we selected the SNPs with significant associations for the plasma lipidome ($p < 5 \times 10^{-8}$); (3) we demonstrated independent association linkage disequilibrium (LD) clumping $r^2 < 0.001$ and distance > 10,000 kb; (4) we removed palindromic SNPs (SNP with the A/T or G/C alleles) after matching the outcome.

Mendelian randomization analysis

In this study, we used 412 GM taxa and 179 plasma lipid species as exposure factors for de-



Figure 1. Overview of the study (by Figdraw 2.0). Step 1 describes the bi-directional causal effects between GM and dementia. Step 2 describes the bi-directional causal effects between the plasma lipidome and dementia. In Step 3, the mediation analysis of the plasma lipidome from the GM to dementia is outlined

mentia, analyzed using MR with the TwoSample-MR package. We used the IVW method to assess causal relationships, presenting results with odds ratios and 95% confidence intervals. Statistical significance was determined with a *p*-value less than 0.05 for the IVW method, with consistency in direction between IVW and MR-Egger. Suggestive associations were those with a *p*-value below 0.05 but above the Bonferroni-corrected threshold.

In the mediation analysis, this study identified the GM and plasma lipidome as having significant causal effects on dementias through two-sample analysis. The investigation aimed to determine whether GM had a causal impact on plasma lipidome, and subsequently utilized multiple MR analyses to assess whether the plasma lipidome acted as a mediator in the pathway from the GM to dementia. Bi-directional causal effects were tested between the GM, plasma lipidome and dementias. We employed dementias as the "exposure" variable and identified GM or plasma lipidome linked to dementias as the outcome variables. We utilized SNPs that were found to be significantly correlated with dementia ($p < 5 \times 10^{-8}$) as IVs.

We tested heterogeneity in IVW estimates with Cochran's Q test and visualized MR results with scatter plots. We conducted sensitivity analyses and used MR-PRESSO and MR-Egger regression to check for horizontal pleiotropy [22]. MR-PRESSO was used to identify and correct outliers, addressing horizontal pleiotropy effects [23]. The Steiger directionality test was used to establish causality, identifying genetic variants with stronger correlations with the outcome than the exposure. Variants identified by the Steiger test were excluded from subsequent analysis. Analysis of MR was carried out using the R package TwoSampleMR (version 4.3.2).

Results

A total of 2774 SNPs were chosen as IVs for the 412 GM taxa and bacterial pathways (Supplementary Table SI). Subsequently, 601 SNPs were identified as being associated with 179 plasma lipid species (Supplementary Table SII).

Causal effects of GM and plasma lipidome on multiple dementia types

AD

The findings of our study demonstrated that 8 GM taxa and 11 bacterial pathways were associated with AD (Supplementary Table SIII, Figure 2). MR analysis suggested that genetic prediction of 6 GM taxa and 4 bacterial pathways were positively correlated with AD. The species *Desulfovibrio piger* (OR = 1.206, p = 0.003), species *Paraprevotella* unclassified (OR = 1.162,

Exposure	No.of SNP	Outcome	OR(95% CI)		Р
s Desulfovibrio piger	6	AD	1.206 (1.066 to 1.364)	jee .	0.003
s Paraprevotella unclassified	9	AD	1.162 (1.011 to 1.335)		0.035
s Alistipes indistinctus	12	AD	1.223 (1.076 to 1.389)	H	0.002
s Phaseolarctobacterium succinatutens	9	AD	0.897 (0.819 to 0.981)	104	0.018
g Phascolarctobacterium	9	AD	0.897(0.819 to 0.981)	H	0.018
f Clostridiales noname	8	AD	1.183(1.017 to 1.380) 1.227 (1.059 to 1.421)	-	0.029
g Paraprevotella	10	AD	1.227 (1.039 to 1.421) 1 154 (1 027 to 1 297)		0.000
PWY.7392	10	AD	0.864 (0.790 to 0.945)	101	0.001
PWY.6285	12	AD	0.917 (0.849 to 0.991)	E.	0.029
PWY.66.422	10	AD	1.180 (1.014 to 1.374)		0.033
PRPP.PWY	14	AD	0.860 (0.747 to 0.990)		0.036
PWY.3001	8	AD	0.728 (0.576 to 0.920)	Here I	0.008
PWY.5104	9	AD	0.817 (0.695 to 0.961)		0.014
PW Y.5188	12	AD	1.325 (1.143 to 1.536)	H++1	0.000
PW 1.5080 NACI IDASVNI DWV	0	AD	0.705(0.57100.871) 1.170(1.001 to 1.368)		0.001
NONOXIPENT PWV	6	AD	1.170(1.001 to 1.308) 1 230 (1 014 to 1 493)		0.048
PENTOSE P.PWY	13	AD	0.868 (0.760 to 0.990)	144	0.035
s Bilophila unclassified	11	FTD	0.277 (0.110 to 0.701)	H	0.007
s Bacteroides thetaiotaomicron	8	FTD	0.189 (0.072 to 0.495)		0.001
s Lachnospiraceae bacterium 5_1_63FAA	46	FTD	0.572 (0.338 to 0.967)	→→→	0.037
s Adlercreutzia equolifaciens	7	FTD	3.024 (1.142 to 8.009)		→0.026
s Lactobacillus delbrueckii	13	FTD	1.450 (1.029 to 2.044)		0.034
g Escherichia	9	FTD	3.613 (1.357 to 9.619)		→0.010
o Enterobacteriales	5	FID	9.328 (2.442 to 35.624)		$\rightarrow 0.001$
c Gammaproteobacteria	4	FID	4.844 (1.080 to 21.714)		0.039
f Enterobacteriaceae	5	FTD	9 324 (2 442 to 35 604)		0.030
g Adlercreutzia	7	FTD	3.016(1.141 to 7.974)		$\rightarrow 0.001$
g Eubacterium	6	FTD	4.815 (1.161 to 19.969))	
PWY.6151	12	FTD	3.234 (1.334 to 7.839)		→0.009
HOMOSER.METSYN.PWY	9	FTD	2.998 (1.077 to 8.348)		-0.036
P23.PWY	10	FTD	1.640 (1.026 to 2.621)		• 0.039
ANAEROFRUCAT.PWY	5	FTD	0.174 (0.038 to 0.807)	H=====1	0.025
s Escherichia coli	11	DLB	0.775 (0.633 to 0.949)	HH.	0.014
g Roseburia	13	DLB	1.340 (1.024 to 1.752)		0.033
PWV 5101	12	DLB	1.382(1.037 to 1.807) 0.758 (0.575 to 0.999)		0.018
PWY 5705	9	DLB	0.756 (0.644 to 0.889)	144	0.001
s Bacteroides coprocola	4	PDD	0.719 (0.518 to 0.999)		0.049
s Bacteroides intestinalis	3	PDD	0.492 (0.283 to 0.854)	H	0.012
p Proteobacteria	10	PDD	1.643 (1.062 to 2.541)		• 0.026
s Bifidobacterium bifidum	15	PDD	1.275 (1.031 to 1.578)		0.025
f Clostridiaceae	10	PDD	1.461 (1.060 to 2.013)		0.021
g Lactobacillus	11	PDD	1.356 (1.080 to 1.704)		0.009
PW Y./196	7	PDD	1.897 (1.071 to 3.358)		$\rightarrow 0.028$
PWY 7400	8		1.041 (1.095 to 2.459) 0.568 (0.341 to 0.948)		0.010
SULFATE CYS PWY	12	PDD	0.628 (0.395 to 0.997)		0.030
PWY.5791	15	PDD	0.757 (0.575 to 0.998)	+++	0.048
PWY.6121	10	PDD	1.867 (1.001 to 3.481)		→0.050
PWY.6147	14	PDD	0.644 (0.429 to 0.967)		0.034
PWY.1269	6	PDD	2.182 (1.092 to 4.360)		→0.027
PWY.5384	8	PDD	1.733 (1.076 to 2.789)		- 0.024
PWY.5695	11	PDD	1.767 (1.060 to 2.947)	· · ·	-0.029
GLYCOCALPWY	11	PDD VD	0.564 (0.384 to 0.828)		0.004
s Bacteroides correcola	8	VD	0.939 (0.895 to 0.986) 0.939 (0.882 to 1.000)	- 3	0.012
s Bacteroides fragilis	18	VD	1.058 (1.005 to 1.114)		0.033
s Ruminococcus torques	7	VD	0.917 (0.844 to 0.997)	H	0.041
s Alistipes senegalensis	12	VD	1.108 (1.012 to 1.213)		0.027
s Eubacterium hallii	11	VD	1.111 (1.042 to 1.184)	-	0.001
s Ruminococcus bromii	7	VD	0.870 (0.784 to 0.966)	and the	0.009
s Phascolarctobacterium succinatutens	9	VD	0.928 (0.872 to 0.988)	H	0.020
g Phascolarctobacterium	9	VD	0.928 (0.871 to 0.988)	H	0.020
PWY.GLYOXYLATE.BYPASS	11	VD	1.078 (1.002 to 1.160)	eel .	0.043
PWY0 1470	10	VD	0.957 (0.917 to 0.999)	N	0.045
PWV 5188	12	VD	1.117 (1.018 to 1.225) 1 124 (1 029 to 1 227)	let	0.020
1 11 1.5100	12		1.124 (1.027 (0 1.227)		
				0 0.5 1 1.5 2 2	.5 3

Figure 2. Mendelian randomization results of causal effects between GM taxa and dementias

p = 0.035), species Alistipes indistinctus (OR = 1.223, p = 0.002), genus Escherichia (OR = 1.185, p = 0.029), family Clostridiales noname (OR = 1.227, p = 0.006), genus Paraprevotella (OR = 1.154, p = 0.016), PWY.66.422 (OR = 1.180, p = 0.033), PWY.5188 (OR = 1.325, p < 0.001), NAGLI-PASYN.PWY (OR = 1.170, p = 0.048) and NONOX-IPENT.PWY (OR = 1.230, p = 0.036) may increase the risk of developing AD.

Genetic prediction of 2 GM taxa and 7 bacterial pathways were negatively correlated with AD. The species *Phascolarctobacterium succinatutens* (OR = 0.897, p = 0.018), genus *Phascolarctobacterium* (OR = 0.897, p = 0.018), PWY.7392 (OR = 0.864, p = 0.001), PWY.6285 (OR = 0.917, p = 0.029), PRPP.PWY (OR = 0.860, p = 0.036), PWY.3001 (OR = 0.728, p = 0.008), PWY.5104 (OR = 0.817, p = 0.014), PWY.5686 (OR = 0.705, p = 0.001) and PENTOSE.P.PWY (OR = 0.868, p = 0.035) may reduce the risk of developing AD.

The findings of our study demonstrated that 6 lipid species exhibit associations with AD (Supplementary Table SIV, Figure 3). MR analysis suggested that genetic prediction of all 6 lipid species were negatively correlated with AD. Ceramide (d40:1) levels (OR = 0.875, p = 0.024), ceramide (d42:1) levels (OR = 0.825, p = 0.009), ceramide (d42:2) levels (OR = 0.896, p = 0.034), phosphatidylcholine (18:0_20:3) levels (OR = 0.835, p = 0.015), phosphatidylethanolamine (16:0_18:2) levels (OR = 0.910, p = 0.010) and phosphatidylethanolamine (18:0_18:2) levels (OR = 0.936, p = 0.046) may reduce the risk of developing AD.

FTD

The findings of our study demonstrated that 12 GM taxa and 4 bacterial pathways were associated with FTD (SupplementaryTable SIII, Figure 2). MR analysis suggested that genetic prediction of 9 GM taxa and 3 bacterial pathways were positively correlated with FTD. The species Adlercreutzia equolifaciens (OR = 3.024, p = 0.026), species

Lactobacillus delbrueckii (OR = 1.450, p = 0.034), genus Escherichia (OR = 3.613, p = 0.010), order Enterobacteriales (OR = 9.328, p = 0.001), class Gammaproteobacteria (OR = 4.844, p = 0.039), family Eubacteriaceae (OR = 4.813, p = 0.030), family Enterobacteriaceae (OR = 9.324, p = 0.001), genus Adlercreutzia (OR = 3.016, p = 0.026), genus Eubacterium (OR = 4.815, p = 0.030), PWY.6151 (OR = 3.234, p = 0.009), HOMOSER.METSYN.PWY (OR = 2.998, p = 0.036) and P23.PWY (OR = 1.640, p = 0.039) may increase the risk of developing FTD.

Genetic prediction of 3 GM taxa and 1 bacterial pathway were negatively correlated with FTD. The species *Bilophila* unclassified (OR = 0.277, p = 0.001), species *Bacteroides thetaiotaomicron* (OR = 0.189, p = 0.001), species *Lachnospiraceae bacterium* 5_1_63FAA (OR = 0.572, p = 0.037), and ANAEROFRUCAT.PWY (OR = 0.174, p = 0.025) may reduce the risk of developing FTD.

No lipid species were associated with FTD.

DLB

The findings of our study demonstrated that 3 GM taxa and 2 bacterial pathways were associated with DLB (Supplementary Table SIII, Figure 2). MR analysis suggested that genetic prediction of 2 GM taxa were positively correlated with DLB. The genus *Roseburia* (OR = 1.340, p = 0.033) and genus *Parabacteroides* (OR = 1.382, p = 0.018) may increase the risk of developing DLB.

Genetic prediction of 1 GM taxon and 2 bacterial pathways were negatively correlated with DLB. The species *Escherichia coli* (OR = 0.775, p = 0.014), PWY.5101 (OR = 0.758, p = 0.049) and PWY.5705 (OR = 0.756, p = 0.001) may reduce the risk of developing DLB.

The findings of our study demonstrated that 4 lipid species exhibit associations with DLB (Supplementary Table SIV, Figure 3). MR analysis suggested that genetic prediction of all 4 lipid species was negatively correlated with DLB. Sterol ester

Exposure	No.of SNP	Outcome	OR(95% CI)			Р
Ceramide (d40:1) levels	6	AD	0.875 (0.780 to 0.983)	H+++		0.024
Ceramide (d42:1) levels	3	AD	0.825 (0.714 to 0.953)			0.009
Ceramide (d42:2) levels	8	AD	0.896 (0.810 to 0.992)	1-0-1		0.034
Phosphatidylcholine (18:0_20:3) levels	3	AD	0.835 (0.721 to 0.966)	I		0.015
Phosphatidylethanolamine (16:0_18:2) leve	ls 6	AD	0.910 (0.847 to 0.978)	Het		0.010
Phosphatidylethanolamine (18:0_18:2) leve	ls10	AD	0.936 (0.877 to 0.999)	104		0.046
Sterol ester (27:1/16:0) levels	8	DLB	0.785 (0.626 to 0.986)	→ →→		0.037
Phosphatidylcholine (O-16:0_18:1) levels	3	DLB	0.647 (0.433 to 0.969)			0.035
Phosphatidylcholine (O-16:1_18:2) levels	2	DLB	0.581 (0.371 to 0.909)	—		0.018
Phosphatidylcholine (O-18:1_20:4) levels	4	DLB	0.791 (0.628 to 0.995)	H		0.046
Ceramide (d42:1) levels	3	VD	0.889 (0.816 to 0.968)	144		0.007
Phosphatidylcholine (18:0_18:2) levels	5	VD	0.929 (0.866 to 0.996)	101		0.039
Phosphatidylcholine $(O-16:1_18:2)$ levels	2	VD	0.784 (0.656 to 0.937)	H+++		0.007
Phosphatidylethanolamine (18:0_18:2) leve	ls10	VD	0.946 (0.901 to 0.993)	141		0.024
				0 05 1	1.5	
				0 0 5 1	1.5	2

Figure 3. Mendelian randomization results of causal effects between plasma lipidome and dementias

(27:1/16:0) levels (OR = 0.785, p = 0.037), phosphatidylcholine (O-16:0_18:1) levels (OR = 0.647, p = 0.035), phosphatidylcholine (O-16:1_18:2) levels (OR = 0.581, p = 0.018) and phosphatidylcholine (O-18:1_20:4) levels (OR = 0.791, p = 0.046) may reduce the risk of developing DLB.

PDD

The findings of our study demonstrated that 6 GM taxa and 11 bacterial pathways were associated with PDD (Supplementary Table SIII, Figure 2). MR analysis suggested that genetic prediction of 4 GM taxa and 6 bacterial pathways were positively correlated with PDD. The phylum Proteobacteria (OR = 1.643, p = 0.026), species *Bifidobacterium bifidum* (OR = 1.275, p = 0.025), family Clostridiaceae (OR = 1.461, p = 0.021), genus Lactobacillus (OR = 1.356, p = 0.009), PWY.7196 (OR = 1.897, p = 0.028), PWY.7315 (OR = 1.641, p = 0.016), PWY.6121 (OR = 1.867, p = 0.050), PWY.1269 (OR = 2.182, p = 0.027), PWY.5384 (OR = 1.733, p = 0.024) and PWY.5695 (OR = 1.767, p = 0.029) may increase the risk of developing PDD.

Genetic prediction of 2 GM taxa and 5 bacterial pathways were negatively correlated with PDD. The species *Bacteroides coprocola* (OR = 0.719, p = 0.049), species *Bacteroides intestinalis* (OR = 0.492, p = 0.012), PWY.7400 (OR = 0.568, p = 0.030), SULFATE.CYS.PWY (OR = 0.628, p = 0.049), PWY.5791 (OR = 0.757, p = 0.048), PWY.6147 (OR = 0.644, p = 0.034) and GLYCOCAT.PWY (OR = 0.564, p = 0.004) may reduce the risk of developing PDD.

No lipid species were associated with PDD.

VD

The findings of our study demonstrated that 9 GM taxa and 4 bacterial pathways were associated with VD (Supplementary Table SIII, Figure 2). MR analysis suggested that genetic prediction of 3 GM taxa and 3 bacterial pathways were positively correlated with VD. The species *Bacteroides fragilis* (OR = 1.058, p = 0.033), species *Alistipes senegalensis* (OR = 1.108, p = 0.027), species *Eubacterium hallii* (OR = 1.111, p = 0.001), PWY.GLYOXYL-ATE.BYPASS (OR = 1.078, p = 0.043), PWY0.1479 (OR = 1.117, p = 0.020) and PWY.5188 (OR = 1.124, p = 0.009) may increase the risk of developing VD.

Genetic prediction of 6 GM taxa and 1 bacterial pathway were negatively correlated with VD. The species *Bacteroides clarus* (OR = 0.939, p = 0.012), species *Bacteroides coprocola* (OR = 0.939, p = 0.050), species *Ruminococcus torques* (OR = 0.917, p = 0.041), species *Ruminococcus bromii* (OR = 0.870, p = 0.009), species *Phascolarctobacterium succinatutens* (OR = 0.928, p = 0.020), genus *Phascolarctobacterium* (OR = 0.928, p = 0.020)

and PWY.5850 (OR = 0.957, p = 0.045) may reduce the risk of developing VD.

The findings of our study demonstrated that 4 lipid species exhibit associations with VD (Supplementary Table SIV, Figure 3). MR analysis suggested that genetic prediction of all 4 lipid species were negatively correlated with VD. Ceramide (d42:1) levels (OR = 0.889, p = 0.007), phosphatidylcholine (18:0_18:2) levels (OR = 0.929, p = 0.039), phosphatidylcholine (O-16:1_18:2) levels (OR = 0.784, p = 0.007) and phosphatidylethanolamine (18:0_18:2) levels (OR = 0.946, p = 0.024) may reduce the risk of developing VD.

Sensitivity analyses

Our research found no genetic pleiotropy or horizontal pleiotropy influencing the results, and there was no statistically significant heterogeneity in the dataset (Supplementary Table SV). The "leave-one-out" analysis demonstrated the reliability of the MR analysis. Scatter plots illustrated the collective impact of the GM on dementia. Furthermore, the forest plots showed causal associations between the GM and dementia (Supplementary Figures S1–S8).

Bi-directional causal effects of dementias on GM and plasma lipidome

Based on Supplementary Table SVI, there was no reverse effect between GM or plasma lipidome on DLB, PDD and VD. No SNP can be used as an IV after matching FTD with the GM or plasma lipidome. AD had causal effects on phosphatidylethanolamine (16:0_18:2) levels (OR = 1.063, p = 0.021).

Mediation effect of plasma lipidome

This study discovered that both the gut microbiota (GM) and plasma lipidome play a significant role in the development of dementias. The plasma lipidome may act as a mediator between the GM and dementia, with a key condition being the association between the GM and plasma lipidome. Additional analysis identified 11 potential mediation pathways (Supplementary Table SVII, Figure 4). Ultimately, only 1 potential mediation pathway was found to have a real mediating effect (Table I).

Discussion

In our study, we found that certain GM taxa, bacterial pathways, and plasma lipids are linked to dementia risk. Specifically, phosphatidylcholine (O-16:1_18:2) levels may explain a significant portion of the effect of *Bacteroides coprocola* on reducing vascular dementia. Our analysis supports a causal relationship between GM taxa, plasma lipids, and dementia.

Exploring the mediating role of the plasma lipidome in the pathway from the gut microbiota to dementia: a Mendelian randomization study

Type of dementia	Exposure	No.of SNP	Outcome	OR(95% CI)		Р
AD	PWY.3001	8	Ceramide (d40:1) levels	1.198 (1.030 to 1.393)) (0.019
AD	PWY.3001	8	Ceramide (d42:1) levels	1.182 (1.017 to 1.373)) 🛏	0.030
AD	PWY.6285	12	Ceramide (d42:2) levels	0.933 (0.873 to 0.996	H (0.038
AD	s Paraprevotella unclassified	9	Phosphatidylcholine (18:0 20:3) levels	1.151 (1.007 to 1.316) 🛏	0.039
AD	s Phascolarctobacterium succinatutens	8	Ceramide (d42:2) levels	0.917 (0.845 to 0.996) 🛏	0.039
AD	g Phascolarctobacterium	8	Ceramide (d42:2) levels	0.917 (0.845 to 0.996) 🛏	0.039
VD	s Bacteroides coprocola	4	Phosphatidylcholine (O-16:1 18:2) levels	1.178 (1.077 to 1.289)	0.000
VD	s Bacteroides coprocola	4	Phosphatidylcholine (18:0 18:2) levels	1.145 (1.047 to 1.251) 🛏	0.003
VD	s Phascolarctobacterium succinatutens	7	Phosphatidylethanolamine (18:1_18:1) levels	s1.127 (1.029 to 1.234) 🛏	0.010
VD	g Phascolarctobacterium	7	Phosphatidylethanolamine (18:1 18:1) levels	s1.127 (1.029 to 1.234) 🛏	0.010
VD	s Bacteroides coprocola	4	Phosphatidylcholine (O-16:0_18:1) levels	1.119 (1.022 to 1.225) 🛏	0.015
	•					
					0 05 1 1	5 2

Figure 4. Mendelian randomization results of causal effects between plasma lipidome and dementias

Table I. Two-step	MR estimates	for the gut r	nicrobiota on	VD risk b	y plasma	lipidome i	mediato
-------------------	--------------	---------------	---------------	-----------	----------	------------	---------

Gut microbiota	Plasma lipidome	Outcome	Direct effect	Total effect	Mediated proportion	<i>P</i> -value
Species Bacteroides coprocola	Phosphatidylcholine (O-16:1_18:2) levels	VD	-0.023	-0.063	63.6%	0.032

The GM and gut-derived metabolites play a crucial role in maintaining an individual's physiological functions, particularly brain functions. The MGBA helps communication between the nervous system and gastrointestinal tract, involving the central nervous system, enteric nervous system, and hypothalamic-pituitary-adrenal axi [24]. Previous research has linked the GM, the plasma lipidome, and dementia, but our study goes further to identify a causal relationship between specific GM taxa, plasma lipid species, and dementia [25].

Our study found a positive correlation between 6 GM taxa and 4 bacterial pathways with AD, and a negative correlation between the genetic prediction of 2 GM taxa and 7 bacterial pathways with AD. Li et al. discovered a decrease in oxidative stress and inflammatory-related GM taxa, like Alistipes and Desulfovibrio, after treating an AD mouse model. They also observed a decrease in Aβ accumulation in the hippocampus and an increase in antioxidation enzyme activity with PC12 cells [26]. Sun et al. conducted a comparative analysis of the GM and metabolome in APP^{swe}/ PS1^{AE9}(PAP) exhibiting cognitive decline and agematched controls. Their findings revealed a significant increase in the abundance of Paraprevotella in the GM of the cognitive decline group [27]. For Phascolarctobacterium, there are heterogeneous findings in different studies. In a meta-analysis that included 11 studies, researchers found that the intestinal Phascolarctobacterium of AD patients increased significantly [28]. However, Jemimah's study found that the intestinal *Phascolarc*tobacterium of AD patients decreased significantly [29]. Galactose degradation (Leloir pathway) has been closely linked to brain senescence and has frequently been used in the construction of AD mouse models [30, 31]. Tynkkynen's study found a link between isoleucine and reduction of AD risk, supporting our findings on the potential role of MGBA [32]. Our research also showed conflicting effects of the pentose phosphate pathway and its non-oxidative branch on AD, suggesting a possible risk factor in disrupting the complete pathway in the relationship between MGBA and AD.

The results of our study revealed a positive correlation between genetic prediction of 9 GM taxa and 3 bacterial pathways with FTD, as well as a negative correlation between genetic prediction of 3 GM taxa and 1 bacterial pathway with FTD. Yang's research revealed that Bacteroides thetaiotaomicron significantly contributed to cognitive impairment in a mouse model of dementia [33]. Furthermore, as an enzyme that converts cholesterol to cholesterol-3-sulfonate, Bacteroides thetaiotaomicron played a role in regulating blood cholesterol levels by sulfonating steroidal metabolites, suggesting a potential avenue for mitigating frontotemporal dementia [34]. The findings regarding the impact of Lachnospiraceae on dementia research have been inconsistent. Li's study suggested that Lachnospiraceae may contribute to excitotoxic effects, metabolic damage, inflammatory responses, and neural and astrocytic apoptosis through quinolinic acid synthesis [35]. Additionally, the study highlights a potential link between the biosynthesis of the methionine-related pathway and the development of FTD. Stopa's research revealed a notable decrease in the decomposition of methionine in individuals diagnosed with frontotemporal dementia [36].

The results of our study revealed a positive correlation between genetic prediction of 2 GM taxa with DLB, as well as a negative correlation between genetic prediction of 1 GM taxon and 2 bacterial pathways with DLB. Klein found that the functional amyloid fibers produced by *Escherichia*

had a similar structure to alpha synuclein, which was closely related to the pathogenesis of DLB [37]. This study is the first to report that *Roseburia* and *Parabacteroides* are risk factors for DLB, which may have potential biological application value.

The findings of our study revealed a positive correlation of the genetic prediction of 4 GM taxa and 6 bacterial pathways with PDD, alongside a negative correlation of the genetic prediction of 2 GM taxa and 5 bacterial pathways with PDD. Chang's research suggested that *Bacteroides* may have a positive impact on cognitive function in individuals with Parkinson's disease by metabolizing D-glutamate [38]. Additionally, Heravi's study indicated an increase in the expression of Bifidobacterium in patients with PDD, while no significant difference was observed in Proteobacteria levels between PDD patients and the general population [39]. Despite previous beliefs associating Lactobacillus with beneficial effects on health, this study revealed that Lactobacillus may actually be a risk factor for PDD [40]. This finding underscored the importance of further examining the use of certain probiotics in the context of neurological diseases. In terms of protective factors for PDD, our findings were similar to those of recent studies in PD patients [41].

The results of our study revealed a positive correlation between genetic prediction of 3 GM taxa and 3 bacterial pathways with VD, as well as a negative correlation between genetic prediction of 6 GM taxa and 1 bacterial pathway with VD. There is an ongoing debate surrounding the role of *Bacteroides* in cognitive function. While certain studies had reported a notable increase in *Bacteroides* within the GM of individuals with vascular dementia and post-stroke cognitive impairment, other research suggested a decrease in *Bacteroides* among those with cognitive impairment [42, 43].

Our research involved a detailed classification and analysis of Bacteroides, revealing Bacteroides clarus and Bacteroides coprocola as protective factors against VD, while Bacteroides fragilis emerged as a risk factor for VD. Additionally, Wu's study identified an enrichment of Bacteroides clarus and Bacteroides coprocola in individuals with low levels of indole-3-acetic acid, whereas Bacteroides fragilis was found to be enriched in those with high levels of indole-3-acetic acid, indicating a significant risk factor for vascular cognitive impairment [44, 45]. Xia's investigation revealed the involvement of Bacteroides fragilis in the activation of microglia and the induction of Alzheimer's disease pathologies in Thy1-C/EBPB transgenic mice [46]. In a separate study, Zhao demonstrated that Bacteroides fragilis may contribute to the development of neuroinflammation via lipopolysaccharides, resulting in cognitive impairment [47]. These findings underscore the significance of identifying precise bacterial strains in future research on the MGBA.

This study aimed to investigate the impact of GM taxa on dementia by analyzing their relative abundance expression. However, the precise mechanism underlying the relationship between GM and dementia remains unclear. It was hypothesized that plasma lipid species may serve as mediators in the interaction between the GM and the development of dementia.

MR analysis suggested that genetic predictions of all 6 lipid species were negatively correlated with AD, no lipid species were associated with FTD, all 4 lipid species were negatively correlated with DLB, no lipid species were associated with PDD, and all 4 lipid species were negatively correlated with VD. Interestingly, our research revealed a significant correlation between lipid levels and the reduction of dementia incidence. Numerous studies have demonstrated that maintaining normal lipid metabolism in the central nervous system could greatly decrease the risk of dementia. Ceramide, phosphatidylcholine, phosphatidylethanolamine, and sterol ester all exhibited protective properties against various forms of dementia, a finding supported by multiple studies. Additionally, research suggested that proper metabolism of sphingomyelin and ceramide may facilitate synaptic plasticity and cognitive enhancement [48]. Ylilauri's research revealed a significant association between increased phosphatidylcholine intake and reduced risk of dementia and enhanced cognitive function [43].

Our research found that phosphatidylcholine $(O-16:1_18:2)$ level mediated the causal effects of the species *Bacteroides coprocola* on reduction of VD (proportion mediated = 63.6%). While previous research has linked the GM to dementia, the exact ways it affects vascular dementia are still unclear. More research in this area could improve our understanding. Our findings suggest that targeting *Bacteroides coprocola* could help treat vascular dementia. This approach involves using various treatments such as antibiotics, modified bacteria, prebiotics, and metabolites to control its levels. Further research is needed to understand the role of *Bacteroides coprocola* in clinical practice.

In conclusion, this study identified important associations between GM taxa, bacterial pathways, plasma lipids, and different types of dementia. These findings offer insights into potential biomarkers and treatment options for these complex diseases. Additionally, the study revealed the diverse mechanisms involved in dementia development, showing that GM taxa can be both protective and risk factors for dementia, emphasizing the complex relationship between microbial communities and disease progression. More research is needed to understand how the GM is connected to dementia. However, there are certain limitations to this study. Firstly, the study population was limited to European individuals, excluding other ethnic groups. Secondly, the specific mechanisms through which the gut microbiota affects the occurrence and progression of dementia via lipids were not elucidated in this study. Our study suggests that altering the GM could help reduce dementia risk and improve patient outcomes. However, more research is necessary to apply these findings in clinical practice.

Acknowledgments

The authors are grateful to the investigators of the original studies for sharing the GWAS summary statistics.

Binghan Li, Xu Sun and Xiao Luo contributed equally.

Funding

The study was supported by the Natural Science Foundation of China (82101484,82371495), Shanghai Sailing Program (21YF1437600), Major clinical research projects of Shanghai Shen-Kang Hospital Development Center (SHDC-2020CR1038B) and Scientific research project of Shanghai Health Commission (202340066).

Ethical approval

Not applicable.

Conflict of interest

The authors declare no conflict of interest.

References

- GBD 2016 Neurology Collaborators. Global, regional, and national burden of neurological disorders, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016. Lancet Neurol 2019; 18: 459-80.
- 2. GBD 2019 Dementia Forecasting Collaborators. Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the Global Burden of Disease Study 2019. Lancet Public Health 2022; 7: e105-25.
- Elahi FM, Miller BL. A clinicopathological approach to the diagnosis of dementia. Nat Rev Neurol 2017; 13: 457-76.
- 4. Valletta M, Vetrano DL, Xia X, et al. Multimorbidity patterns and 18-year transitions from normal cognition to dementia and death: a population-based study. J Internal Med 2023; 294: 326-35.
- 5. Wimo A, Seeher K, Cataldi R, et al. The worldwide costs of dementia in 2019. Alzheimers Dement 2023; 19: 2865-73.
- 6. Sorboni SG, Moghaddam HS, Jafarzadeh-Esfehani R, Soleimanpour S. A comprehensive review on the role of the gut microbiome in human neurological disorders. Clin Microbiol Rev 2022; 35: e0033820.

- 7. Fan S, Guo W, Xiao D, et al. Microbiota-gut-brain axis drives overeating disorders. Cell Metabol 2023; 35: 2011-27.
- 8. Agirman G, Yu KB, Hsiao EY. Signaling inflammation across the gut-brain axis. Science 2021; 374: 1087-92.
- 9. Loh JS, Mak WQ, Tan LKS, et al. Microbiota-gut-brain axis and its therapeutic applications in neurodegenerative diseases. Signal Transd Targeted Ther 2024; 9: 37.
- 10. Wang X, Ma H, Li X, et al. Association of cardiovascular health with life expectancy free of cardiovascular disease, diabetes, cancer, and dementia in UK adults. JAMA Inter Med 2023; 183: 340-9.
- 11. Ferguson EL, Zimmerman SC, Jiang C, et al. Low- and high-density lipoprotein cholesterol and dementia risk over 17 years of follow-up among members of a large health care plan. Neurology 2023; 101: e2172-84.
- 12. Baloni P, Arnold M, Buitrago L, et al. Multi-Omic analyses characterize the ceramide/sphingomyelin pathway as a therapeutic target in Alzheimer's disease. Commun Biol 2022; 5: 1074.
- 13. Green RE, Lord J, Scelsi MA, et al.; Insight 46 study team. Investigating associations between blood metabolites, later life brain imaging measures, and genetic risk for Alzheimer's disease. Alzheimer's Res Ther 2023; 15: 38.
- Yoon JH, Seo Y, Jo YS, et al. Brain lipidomics: From functional landscape to clinical significance. Sci Adv 2022; 8: eadc9317.
- 15. Luo J, Yang H, Song BL. Mechanisms and regulation of cholesterol homeostasis. Nat Rev Mol Cell Biol 2020; 21: 225-45.
- 16. Chen C, Liao J, Xia Y, et al. Gut microbiota regulate Alzheimer's disease pathologies and cognitive disorders via PUFA-associated neuroinflammation. Gut 2022; 71: 2233-52.
- 17. Tam V, Patel N, Turcotte M, Bossé Y, Paré G, Meyre D. Benefits and limitations of genome-wide association studies. Nat Rev Genet 2019; 20: 467-84.
- 18. Bowden J, Holmes MV. Meta-analysis and Mendelian randomization: a review. Research Synthesis Methods 2019; 10: 486-96.
- 19. Lopera-Maya EA, Kurilshikov A, van der Graaf A, et al. Effect of host genetics on the gut microbiome in 7,738 participants of the Dutch Microbiome Project. Nat Genet 2022; 54: 143-51.
- Ottensmann L, Tabassum R, Ruotsalainen SE, et al. Genome-wide association analysis of plasma lipidome identifies 495 genetic associations. Nat Commun 2023; 14: 6934.
- 21. Chia R, Sabir MS, Bandres-Ciga S, et al. Genome sequencing analysis identifies new loci associated with Lewy body dementia and provides insights into its genetic architecture. Nat Genet 2021; 53: 294-303.
- 22. Cohen JF, Chalumeau M, Cohen R, Korevaar DA, Khoshnood B, Bossuyt PM. Cochran's Q test was useful to assess heterogeneity in likelihood ratios in studies of diagnostic accuracy. J Clin Epidemiol 2015; 68: 299-306.
- 23. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. Nat Genet 2018; 50: 693-8.
- 24. Khan R, Di Gesù CM, Lee J, McCullough LD. The contribution of age-related changes in the gut-brain axis to neurological disorders. Gut Microbes 2024; 16: 2302801.
- 25. Kim JS, Park H, Lee JH, et al. Effect of altered gene expression in lipid metabolism on cognitive improvement in patients with Alzheimer's dementia following fecal microbiota transplantation: a preliminary study. Ther Adv Neurol Disord 2024; 17: 17562864231218181.

- 26. Li C, Wang N, Zheng G, Yang L. Oral administration of resveratrol-selenium-peptide nanocomposites alleviates Alzheimer's disease-like pathogenesis by inhibiting Aβ aggregation and regulating gut microbiota. ACS Appl Mater Interfaces 2021; 13: 46406-20.
- 27. Sun P, Zhu H, Li X, et al. Comparative metagenomics and metabolomes reveals abnormal metabolism activity is associated with gut microbiota in Alzheimer's disease mice. Int J Mol Sci 2022; 23: 11560.
- Hung CC, Chang CC, Huang CW, Nouchi R, Cheng CH. Gut microbiota in patients with Alzheimer's disease spectrum: a systematic review and meta-analysis. Aging 2022; 14: 477-96.
- 29. Jemimah S, Chabib CMM, Hadjileontiadis L, AlShehhi A. Gut microbiome dysbiosis in Alzheimer's disease and mild cognitive impairment: a systematic review and meta-analysis. PLoS One 2023; 18: e0285346.
- 30. Chiroma SM, Mohd Moklas MA, Mat Taib CN, Baharuldin MTH, Amon Z. d-galactose and aluminium chloride induced rat model with cognitive impairments. Biomed Pharmacother 2018; 103: 1602-8.
- Shwe T, Pratchayasakul W, Chattipakorn N, Chattipakorn SC. Role of D-galactose-induced brain aging and its potential used for therapeutic interventions. Exp Gerontol 2018; 101: 13-36.
- 32. Tynkkynen J, Chouraki V, van der Lee SJ, et al. Association of branched-chain amino acids and other circulating metabolites with risk of incident dementia and Alzheimer's disease: a prospective study in eight cohorts. Alzheimer's Dementia 2018; 14: 723-33.
- 33. Yang B, Sun T, Chen Y, Xiang H, Xiong J, Bao S. The role of gut microbiota in mice with bile duct ligation-evoked cholestatic liver disease-related cognitive dysfunction. Front Microbiol 2022; 3: 909461.
- 34. Krypotou E, Townsend GE, Gao X, et al. Bacteria require phase separation for fitness in the mammalian gut. Science 2023; 379: 1149-56.
- 35. Li Y, Chen Y, Fan Y, Chen Y, Chen Y. Dynamic network modeling of gut microbiota during Alzheimer's disease progression in mice. Gut Microbes 2023; 15: 2172672.
- 36. Stopa EG, Tanis KQ, Miller MC, et al. Comparative transcriptomics of choroid plexus in Alzheimer's disease, frontotemporal dementia and Huntington's disease: implications for CSF homeostasis. Fluids Barriers CNS 2018; 15: 18.
- 37. Klein RD, Shu Q, Cusumano ZT, et al. Structure-function analysis of the curli accessory protein CsgE defines surfaces essential for coordinating amyloid fiber formation. mBio 2018; 9: e01349-18.
- Chang CH, Lin CH, Lane HY. d-glutamate and Gut Microbiota in Alzheimer's Disease. Int J Mol Sci 2020; 21: 2676.
- Heravi FS, Naseri K, Hu H. Gut microbiota composition in patients with neurodegenerative disorders (Parkinson's and Alzheimer's) and healthy controls: a systematic review. Nutrients 2023; 15: 4365.
- 40. Un-Nisa A, Khan A, Zakria M, et al. Updates on the role of probiotics against different health issues: focus on Lactobacillus. Int J Mol Sci 2022; 24: 142.
- 41. Bedarf JR, Hildebrand F, Coelho LP, et al. Functional implications of microbial and viral gut metagenome changes in early stage L-DOPA-naïve Parkinson's disease patients. Genome Med 2017; 9: 39.
- 42. Aljumaah MR, Bhatia U, Roach J, Gunstad J, Azcarate Peril MA. The gut microbiome, mild cognitive impairment, and probiotics: a randomized clinical trial in middle-aged and older adults. Clin Nutr 2022; 41: 2565-76.

- Du Y, Li X, An Y, Song Y, Lu Y. Association of gut microbiota with sort-chain fatty acids and inflammatory cytokines in diabetic patients with cognitive impairment: a cross-sectional, non-controlled study. Front Nutr 2022; 9: 930626.
- 44. Wu PH, Tseng YF, Liu W, et al. Exploring the relationship between gut microbiome composition and blood indole-3-acetic acid in hemodialysis patients. Biomedicines 2024; 12: 148.
- 45. Xia Y, Xiao Y, Wang ZH, et al. Bacteroides Fragilis in the gut microbiomes of Alzheimer's disease activates microglia and triggers pathogenesis in neuronal C/ΕΒΡβ transgenic mice. Nat Commun 2023; 14: 5471.
- 46. Zhao Y, Jaber VR, Pogue AI, Sharfman NM, Taylor C, Lukiw WJ. Lipopolysaccharides (LPSs) as potent neurotoxic glycolipids in Alzheimer's disease (AD). Int J Mol Sci 2022; 23: 12671.
- 47. Custodia A, Romaus-Sanjurjo D, Aramburu-Núñez M, et al. Ceramide/sphingosine 1-phosphate axis as a key target for diagnosis and treatment in Alzheimer's disease and other neurodegenerative diseases. Int J Mol Sci 2022; 23: 8082.
- 48. Ylilauri MPT, Voutilainen S, Lönnroos E, et al. Associations of dietary choline intake with risk of incident dementia and with cognitive performance: the Kuopio Ischaemic Heart Disease Risk Factor Study. Am J Clin Nutr 2019; 110: 1416-23.