Exploring the association between gut microbiota and venous thromboembolism using a Mendelian randomization analysis

Keywords

gut microbiota, venous thromboembolism, pulmonary embolism, deep vein thrombosis, mendelian randomization

Abstract

Introduction

Previous observational studies have suggested a potential association between gut microbiota (GM) and venous thromboembolism (VTE), including pulmonary embolism (PE) and deep vein thrombosis (DVT). However, the causal nature of this association remains uncertain due to potential confounding factors.

Material and methods

The summary statistics for VTE, PE, and DVT were obtained from the GWAS conducted by the FinnGen consortium R9. The genetic data for relevant GM SNPs were extracted from the metaanalysis of GWAS performed by the global MiBioGen consortium. Using SNPs as instrumental variables, the IVW method was primarily employed to assess the bidirectional causal relationship between GM and VTE, PE, and DVT.

Results

For the risk of VTE onset, Candidatus Solea ferrea, Ruminococcaceae UCG002, and Ruminococcaceae UCG004 were negatively correlated, while Eubacterium hallii group, Butyricimonas, and Dorea were positively correlated. For PE, Intestinimonas, Unknown genus, and Firmicutes were negatively correlated, while Veillonella, Erysipelatoclostridium, and Lentisphaerae were positively correlated. For DVT, Mollicutes, Actinobacteria, and Bifidobacteriaceae were negatively correlated, while Adlercreutzia, Collinsella, and Desulfovibrio were positively correlated. After multiple corrections using the Bonferroni method, a significant causal relationship was identified between Ruminococcaceae and VTE. Cochran's Q test was performed to evaluate instrumental variable heterogeneity (P > 0.05), MR-Egger regression analyses were performed to examine pleiotropy (P > 0.05), and Leave-one-out analysis was conducted to assess the impact of each SNP on the outcome.

Conclusions

Specific GM may have causal effects on VTE, PE, and DVT, potentially contributing to the development of microbiota-centered therapeutic approaches and the identification of novel biomarkers for targeted preventive strategies.

1	Exploring the association between gut microbiota and venous thromboembolism
2	using a Mendelian randomization analysis
3	Meijie Yuan ¹ , Weiran Li ¹ , Jian Sun ¹ , Hongshuo Shi ¹ *, Guobin Liu ¹ *
4	¹ Department of Peripheral Vascular Surgery, Shuguang Hospital Affiliated to Shanghai University of
5	Traditional Chinese Medicine, Shanghai, China.
6	*Corresponding author:
7	Guobin Liu, Department of Peripheral Vascular Surgery, Shuguang Hospital Affiliated to Shanghai
8	University of Traditional Chinese Medicine, No.528 Zhangheng Road, Pudong District, Shanghai
9	201203, China.
10	Email: 15800885533@163.com
11	Hongshuo Shi, Department of Peripheral Vascular Surgery, Shuguang Hospital Affiliated to Shanghai
12	University of Traditional Chinese Medicine, No.528 Zhangheng Road, Pudong District, Shanghai
13	201203, China.
14	Email: jf17510413109@163.com
15	
16	
17	
18	
19	
20	
	1

21 Abstract

Introduction: Previous observational studies have suggested a potential association between gut microbiota (GM) and venous thromboembolism (VTE), including pulmonary embolism (PE) and deep vein thrombosis (DVT). However, the causal nature of this association remains uncertain due to potential confounding factors.

Methods : The summary statistics for VTE, PE, and DVT were obtained from the meta-analysis of genome-wide association studies (GWAS) conducted by the FinnGen consortium R9. The genetic data for relevant GM single nucleotide polymorphisms (SNPs) were extracted from the meta-analysis of GWAS performed by the global MiBioGen consortium. Using SNPs as instrumental variables, the inverse variance weighting (IVW) method was primarily employed to assess the bidirectional causal relationship between GM and VTE, PE, and DVT.

32 Results: For the risk of VTE onset, Candidatus Solea ferrea, Ruminococcaceae UCG002, and Ruminococcaceae UCG004 were negatively correlated, while Eubacterium hallii group, 33 34 Butyricimonas, and Dorea were positively correlated. For PE, Intestinimonas, Unknown genus, and 35 Firmicutes were negatively correlated, while Veillonella, Erysipelatoclostridium, and Lentisphaerae were positively correlated. For DVT, Mollicutes, Actinobacteria, and Bifidobacteriaceae were 36 negatively correlated, while Adlercreutzia, Collinsella, and Desulfovibrio were positively correlated. 37 After multiple corrections using the Bonferroni method, a significant causal relationship was 38 39 identified between Ruminococcaceae and VTE. Cochran's Q test was performed to evaluate 40 instrumental variable heterogeneity (P > 0.05), MR-Egger regression analyses were performed to examine pleiotropy (P > 0.05), and Leave-one-out analysis was conducted to assess the impact of 41

42	each SNP on the outcome.
43	Conclusion: Specific GM may have causal effects on VTE, PE, and DVT, potentially contributing to
44	the development of microbiota-centered therapeutic approaches and the identification of novel
45	biomarkers for targeted preventive strategies.
46	Key words : gut microbiota, venous thromboembolism, pulmonary embolism, deep vein thrombosis,
47	mendelian randomization
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
59	
60	
61	
62	
63	
64	
65	
66	
67	

69	Venous thromboembolism (VTE), encompassing pulmonary embolism (PE) and deep vein
70	thrombosis (DVT), is a serious thromboembolic disorder that remains a substantial challenge for
71	global healthcare systems [1]. VTE is a multifactorial disease arising from the interplay between
72	genetic and acquired risk factors, with its heritability estimated at 40% to 60% based on studies
73	involving families, twins, and siblings [2]. A recent study indicated that the annual incidence of VTE
74	in the USA is 123 cases per 100,000 people, with a higher rate among the elderly [3]. PE is more
75	severe, often causing acute obstruction of the pulmonary vasculature, leading to hemodynamic
76	instability and even death [4]. Among cardiovascular complications worldwide, DVT is the third
77	leading cause of death and disability [5]. Therefore, ideveloping novel strategies for the prevention
78	and treatment of VTE is essential to mitigate the socioeconomic consequences associated with its
79	incidence and progression.
80	Anticoagulation therapy is the primary treatment for VTE, and delays in initiating or
81	maintaining therapeutic levels may lead to poorer outcomes [6]. Rivaroxaban, a non-vitamin K oral
82	anticoagulant (NOAC), is extensively employed for patients at heightened risk of thrombosis,
83	especially VTE [7]. Preventing VTE is a crucial goal for medical professionals, requiring a thorough
84	understanding of the risk factors to effectively mitigate the risk. A multicenter cohort study has
85	identified the neutrophil-to-lymphocyte ratio (NLR), lactate dehydrogenase (LDH), C-reactive
86	protein (CRP), and procalcitonin (PCT) as independent predictive factors for VTE [8]. Another
87	Mendelian randomization (MR) analysis found that, among the 41 inflammatory cytokines included,

88 only platelet-derived growth factor-BB (PDGF-BB) levels showed a causal relationship with an

increased risk of VTE, PE, and DVT [9]. Further research is needed to identify precise risk factors for VTE.

91	The human gut harbors trillions of microorganisms, which play a crucial role in maintaining
92	digestive health and immune homeostasis [10]. Research showed that alterations in the composition
93	of these GM are associated with various diseases, including gastrointestinal disorders, metabolic
94	issues, and cardiovascular conditions [11-13]. Environmental or genetic disturbances in GM can
95	trigger inflammatory reactions in blood vessels, platelets, and immune cells, potentially increasing
96	the risk of thrombosis [14]. Disruption of the intestinal epithelial barrier, caused by factors such as
97	inflammation, nutrition, and antibiotics, allowed microbial products and metabolites to enter the
98	systemic circulation via the portal vein, potentially leading to thrombosis [15]. Disturbances in GM
99	can activate pathways involving endothelial cells, platelets, and innate immune cells, leading to the
100	release of coagulation proteins and the development of a prethrombotic state [16]. However, the
101	precise role of GM in the development of thromboembolism is not yet fully understood.

MR uses natural variations in genetic variants across generations to determine causal relationships [17]. By employing single nucleotide polymorphisms (SNPs) associated with specific health conditions as proxies, MR helps identify causal relationships while avoiding the external biases often present in traditional epidemiological studies [18-20]. This approach provides a clearer understanding of the genetic influences on disease.

Our research investigates the influence of GM on venous thromboembolic conditions, including
 VTE, PE, and DVT, using two-sample summary MR with genetic variants associated with GM as
 instruments to explore these relationships.

110 Material and methods

111 Study design

112 This study was designed in accordance with the Strengthening the Reporting of Observational 113 Studies in Epidemiology Using Mendelian Randomization (STROBE-MR) . SNPs associated with the human GM were used as instrumental variables (IVs), with VTE, including PE and DVT, as the 114 115 outcome variable. The study satisfies the three core assumptions of MR analysis: (1) the relevance 116 assumption, indicating that the IVs are significantly associated with the exposure (GM); (2) the independence assumption, ensuring that the IVs are not associated with any confounding variables; 117 118 and (3) the exclusion restriction assumption, which states that the selected genetic variants influence 119 the outcome exclusively through the "IVs-exposure-outcome" pathway, without affecting the 120 outcome via alternative pathways (Figure 1). The data for this study were aggregated from 121 previously published research, for which participant consent and ethical clearance had been obtained.

122 Data resources

123 Forward MR data

The genetic data related to GM were obtained from the global MiBioGen consortium database (https://mibiogen.gcc.rug.nl/), which integrates data from 25 cohorts across multiple countries, including the USA and Italy, involving a total of 18,340 individuals. The primary objective of this study was to identify the relationship between autosomal human genetic variants and GM by analyzing the participants' 16S rRNA sequencing profiles [21]. The genome-wide association study (GWAS) data for VTE, PE, and DVT were derived from the FinnGen consortium R9 release dataset. This genetic dataset includes 19,372 VTE cases and 357,905 controls, 9,109 9,243 PE cases and

131 367,108 controls, as well as DVT cases and 324,121 controls (Table I).

132

Table I. GWAS summary data sources of outcomes.

	Phenotype (Trait)	Data source (Consortium)	Sample size (Case/Contro)	Ancestry	Covariates	Link
	VTE	FinnGen	19,372/357,905	European	sex, age, 10 genotyping batch.	PCs, FinnGen_ Access_Results
	PE		9,243/367,108			
	DVT		9,109/324,121			
133	VTE-venou	s thromboem	oolism, PE-pulm	ionary em	bolism, DVT-dee	ep vein thrombosis,
134	PCs-princip	al components.				
135	Rever	se MR data				
136	For rev	verse MR, we us	ed data similar to	that of the fo	orward MR approac	h. In this context, VTE,
137	PE, and D	VT are consid	ered variables of	interest, a	nd SNPs strongly	associated with these
138	conditions	$(P < 5 \times 10^{-8})$	are identified as	exposure v	ariables. This proc	ess involves removing
139	instances of	f linkage disequi	librium, palindron	nic sequence	es, and weakly corre	elated variables, as well
140	as SNPs inf	luenced by conf	ounding factors, ju	ist as in forw	vard MR.	

141 IVs selection

Based on the three core assumptions of MR analysis, we first selected SNPs significantly associated with GM as IVs. SNPs with *P*-values less than the genome-wide significance threshold (*P* $< 5 \times 10^{-8}$) were chosen as the initial IVs. Secondly, to eliminate linkage disequilibrium (LD), SNPs within an LD region defined by a distance of 10,000 kb and an LD r² < 0.001 were excluded. Subsequently, SNPs associated with known confounding factors, such as cancer, prolonged bed rest, and fractures [22], were removed (http://www.phenoscanner.medschl.cam.ac.uk/). Finally, to ensure allele alignment accuracy, harmonization of SNPs was performed [23]. The F-statistic was calculated using the formula: $F = R^2 (N - K - 1) / [K(1 - R^2)]$, where R^2 represents the proportion of variance explained by the SNPs, N is the number of participants in the exposure group, and K is the total number of SNPs included in the final analysis. An F-statistic ranging from 26.604 to 26.992 indicates a low risk of weak instrument bias. These screening steps ensure the robustness and reliability of the study results.

154 MR Analysis

155 In this study, the inverse variance weighted (IVW) method was employed as the primary 156 analytical approach. In the forward MR analysis, GM served as the exposure variable, with SNPs associated with GM used as IVs to evaluate the causal relationships between VTE, PE, and DVT. 157 Cochran's Q test was conducted to assess heterogeneity across genetic instruments. When P < 0.05, 158 the IVW random-effects model was applied to estimate the causal effect; when the $P \ge 0.05$, a 159 160 fixed-effects model was used [24]. To further validate and complement the causal inference results, 161 additional methods were applied, including MR-Egger regression, weighted median estimator 162 (WME), simple mode (SM), and weighted mode (WM) approaches. The MR-Egger method 163 incorporates an intercept into the regression model to detect and adjust for horizontal pleiotropy in 164 IVs, thereby enhancing the robustness of causal effect estimates [25]. The WME method calculates 165 the weighted median of IV effects, thereby reducing the influence of outliers or biased IVs on the overall results [26]. The SM method estimates the mode of the causal effect distribution, providing 166 reliable estimates when most IVs exhibit similar effects [27]. The WM method calculates the 167 weighted mode of effect results, offering more robust estimates in the presence of effect 168

heterogeneity, particularly when data points have different weights or when outliers are present [28].
By integrating these methods, this study aims to enhance the accuracy and robustness of causal effect
estimates.

172 Statistical analysis

173 In this study, sensitivity analyses included Cochran's Q test, MR-Egger intercept test, 174 MR-PRESSO method, and Leave-one-out analysis. Cochran's Q test was conducted to assess the 175 heterogeneity of the effects of the included IVs or SNPs, aiming to determine whether significant 176 differences exist between data and estimates from different sources. P < 0.05 indicates significant 177 heterogeneity, in which case the IVW random-effects model should be applied; if no significant 178 heterogeneity is detected, the IVW fixed-effects model is used. The MR-Egger intercept test was 179 employed to evaluate and quantify the horizontal pleiotropy of the instrumental variables, aiming to 180 detect and correct potential bias in causal effect estimates. The MR-PRESSO method improves the accuracy of causal inference by identifying and removing outlier SNPs, thereby detecting and 181 182 adjusting for horizontal pleiotropy. The Leave-one-out analysis assesses the robustness and reliability of the results by sequentially removing each SNP and re-evaluating the causal estimates using the 183 184 remaining SNPs. MR analyses were reported using P values, odds ratios (ORs), and 95% confidence 185 intervals (CIs). A significance level of $\alpha = 0.05$ was applied for causal inference. $\beta > 0$ indicates a positive association between the microbiota and the disease, whereas $\beta < 0$ suggests a negative 186 association. Similarly, an OR > 1 indicates a positive association, while an OR < 1 suggests a 187 188 negative association between the microbiota and the disease. All MR analyses were performed using R software (version 4.3.1) with the "Two Sample MR" package (version 0.5.6). 189

190 **Results**

191 IVs selection

According to the selection criteria for IVs, we identified 2,779 eligible SNPs. This dataset 192 193 includes 211 GM taxa, comprising 131 genera, 35 families, 20 orders, 16 classes, and 9 phyla. 194 F-statistics were calculated for each of the 2,779 SNPs, and no SNPs with F < 10 were identified, suggesting a low likelihood of weak instrument bias affecting the causal associations. Based on the 195 IVW analysis, we found that most of the 211 GM taxa were not associated with VTE, whereas only a 196 197 few taxa showed significant associations. Moreover, different types of thrombotic diseases were 198 associated with specific bacterial taxa (Figure 2). In total, we identified 17 SNPs causally related to 199 VTE (Supplementary Table 1), 14 SNPs associated with PE (Supplementary Table 2), and 17 SNPs 200 associated with DVT (Supplementary Table 3).

201 Forward-direction MR analyses

A total of 14 gut bacterial taxa were found to have significant associations with VTE. Among them, *Candidatus Soleaferrea* (id.11350, OR = 0.92, 95% *CI*: 0.85-1.00, P = 0.047), *Ruminococcaceae UCG002* (id.11360, OR = 0.92, 95% *CI*: 0.85-1.00, P = 0.046), and *Ruminococcaceae UCG004* (id.11362, OR = 0.92, 95% *CI*: 0.85-1.00, P = 0.046) were negatively associated with VTE, while the *Eubacterium hallii group* (id.11338, OR = 1.12, 95% *CI*: 1.01-1.23, P = 0.025), *Butyricimonas* (id.945, OR = 1.11, 95% CI: 1.01-1.22, P = 0.027), and *Dorea* (id.1997,

208 OR = 1.14, 95% CI: 1.00-1.31, P = 0.047) were positively associated with VTE.

A total of 9 gut bacterial taxa were found to have significant associations with PE. Among them,

210 Intestinimonas (id.2062, OR = 0.88, 95% CI: 0.78-0.99, P = 0.036), Unknown genus (id.2041, OR =

211	0.87, 95% CI: 0.77-0.97, $P = 0.015$), and Firmicutes (id.1672, $OR = 0.86, 95%$ CI: 0.74-0.99, $P = 0.87, 95%$ CI: 0.77-0.97, $P = 0.015$), and Firmicutes (id.1672, $OR = 0.86, 95%$ CI: 0.74-0.99, $P = 0.87, 95%$ CI: 0.74-0.99, $P = 0.015$), and Firmicutes (id.1672, $OR = 0.86, 95%$ CI: 0.74-0.99, $P = 0.015$), and Firmicutes (id.1672, $OR = 0.86, 95%$ CI: 0.74-0.99, $P = 0.85, 95%$ CI: 0.74-0.99, $P = 0.85, 95%$ CI: 0.74-0.99, $P = 0.015$), and Firmicutes (id.1672, $OR = 0.86, 95%$ CI: 0.74-0.99, $P = 0.85, 95%$ CI: 0.75, $P = 0.85, 95%$
212	0.036) were negatively associated with PE, while Veillonella (id.2198, OR = 1.22, 95% CI: 1.01-1.48,
213	P = 0.043), Erysipelatoclostridium (id.11381, $OR = 1.16$, 95% CI: 1.04-1.30, $P = 0.007$), and
214	<i>Lentisphaerae</i> (id.2250, $OR = 1.14$, 95% CI: 1.03-1.26, $P = 0.023$) were positively associated with
215	PE.

216	A total of 13 gut bacterial taxa were found to have significant associations with DVT. Among
217	them, <i>Mollicutes</i> (id.3920, <i>OR</i> = 0.96, 95% <i>CI</i> : 0.84-1.09, <i>P</i> = 0.004), <i>Actinobacteria</i> (id.419, <i>OR</i> =
218	0.85, 95% CI: 0.74-0.97, P = 0.017), and Bifidobacteriaceae (id.433, OR = 0.85, 95% CI: 0.74-0.99,
219	P = 0.031) were negatively associated with the disease, while <i>Adlercreutzia</i> (id.812, $OR = 1.80, 95%$
220	CI: 0.93-3.50, $P = 0.046$), Collinsella (id.815, $OR = 1.31$, 95% CI: 1.08-1.59, $P = 0.006$), and
221	<i>Desulfovibrio</i> (id.3173, $OR = 1.16$, 95% CI: 1.01-1.33, $P = 0.006$) were positively associated with the
222	disease (Table II).

223	Table II. MR analysis of the causal relationship between GM and VTE, PE, and DVT

Outcome	Exposure	nSNP	P value	OR (95% CI)
VTE	Candidatus Soleaferrea	9	0.047	0.92 (0.85-1.00)
	Ruminococcaceae UCG002	20	0.046	0.92 (0.85-1.00)
	Ruminococcaceae UCG004	11	0.045	0.91 (0.83-1.00)
	Eubacterium hallii group	13	0.025	1.12 (1.01-1.23)
	Butyricimonas	13	0.027	1.11 (1.01-1.22)
	Dorea	10	0.047	1.14 (1.00-1.31)
PE	Intestinimonas	16	0.036	0.88 (0.78-0.99)
	Unknown genus	12	0.015	0.87 (0.77-0.97)
	Firmicutes	14	0.036	0.86 (0.74-0.99)
	Veillonella	5	0.043	1.22 (1.01-1.48)
	Erysipelatoclostridium	15	0.007	1.16 (1.04-1.30)
	Lentisphaeria	8	0.023	1.13 (1.02-1.26)
DVT	Mollicutes	11	0.004	0.82 (0.71-0.94)
	Actinobacteria	14	0.017	0.85 (0.74-0.97)
	Bifidobacteriaceae	11	0.031	0.85 (0.74-0.99)
	Adlercreutzia	8	0.046	1.16 (1.00-1.35)

Collinsella	9	0.006	1.31 (1.08-1.59)
Desulfovibrio	10	0.034	1.16 (1.01-1.33)

224	MR-Mendelian	randomization,	GM-gut	microbion	ne, VTE-ven	ious thrombo	oembolism,
225	PE-encompassing pu	lmonary embolis	sm, DVT-	deep vein	thrombosis, 1	nSNP-number	of single
226	nucleotide polymorph	isms.					

This MR analysis revealed a positive causal relationship between certain GM and the risk levels of VTE, PE, and DVT. After multiple testing correction using the Bonferroni method, a significant causal association was identified between *Ruminococcaceae* and VTE. This suggests that changes in GM may have corresponding effects on thrombotic diseases such as VTE, PE, and DVT. Additionally, the results of supplementary analyses, including MR-Egger, WME, and ML methods, were consistent with the direction of the IVW method. The statistically significant associations are illustrated by scatter plots and forest plots (Figure 3 and Figure 4).

234 **Reverse-direction MR analyses**

In the reverse MR analysis, we applied the same analytical procedures, setting GM as the outcome and VTE, PE, and DVT as the exposure factors. The results showed no causal relationships between GM and VTE (P > 0.05), PE (P > 0.05), or DVT (P > 0.05) (Supplementary Table 4-6).

238 Sensitivity analysis

The funnel plot generated from the MR-Egger regression (Figure 5) indicated no evidence of heterogeneity or horizontal pleiotropy in the forward MR results. Additionally, no outliers were detected in the MR-PRESSO analysis or Cochran's Q test. A Leave-one-out analysis was subsequently performed, and the corresponding forest plot (Figure 6) further confirmed the stability of these results.

244 **DISCUSSION**

To gain a more comprehensive understanding of the impact of GM on VTE development, this 245 246 study conducted bidirectional two-sample MR analyses using summary statistics for VTE, PE, and 247 DVT from the FinnGen Consortium R9 data, along with GM meta-analysis data from the global 248 MiBioGen Consortium. A total of 211 GM taxa were analyzed for causal associations with VTE, PE, 249 and DVT. The analysis identified causal associations between 18 GM taxa and the incidence of VTE, Candidatus Soleaferrea, Ruminococcaceae UCG002, 250 PE. and DVT. Specifically, and 251 Ruminococcaceae UCG004 were negatively associated with VTE, while Eubacterium hallii group, Butyricimonas, and Dorea showed positive associations with VTE (P < 0.05 and OR > 1). For PE 252 253 risk, Intestinimonas, Unknown genus, and Firmicutes were negatively associated, whereas Veillonella, *Erysipelatoclostridium*, and *Lentisphaerae* showed positive associations (P < 0.05 and OR > 1). 254 255 Regarding DVT risk, Mollicutes, Actinobacteria, and Bifidobacteriaceae were negatively associated, 256 while *Adlercreutzia*, *Collinsella*, and *Desulfovibrio* were positively associated (P < 0.05 and OR > 1). These findings enhance the understanding of GM's role in VTE pathogenesis and highlight specific 257 258 taxa that may contribute to or protect against VTE progression.

The pathogenesis of VTE is highly complex. Thrombosis is a pathological process characterized by the abnormal aggregation and solidification of blood components within blood vessels, leading to vascular obstruction, driven by endothelial injury, altered hemodynamics, and a hypercoagulable state [29]. While thrombosis is essential for hemostasis in damaged vessels, it can also result in adverse events such as vascular occlusion, embolism, and pathological clot formation. Conversely, impaired thrombosis may lead to excessive bleeding [30]. The regulatory role of GM in VTE

265	development has increasingly become a research focus, with different bacterial taxa potentially
266	influencing VTE risk through metabolic products, inflammation modulation, and gut barrier function
267	Ruminococcaceae is a key GM family closely associated with various metabolic processes and
268	diseases. Although direct studies on the association between Ruminococcaceae and VTE are limited,
269	metabolites produced by GM, particularly metabolite trimethylamine-N-oxide (TMAO), have been
270	shown to be associated with VTE [31]. One study reported that, compared to low TMAO levels,
271	patients with moderate and high TMAO had a 38% and 44% increased risk of VTE recurrence,
272	respectively. However, the results were not statistically significant [32]. Papa et al. reported that
273	TMAO is a risk factor for inflammatory bowel disease, with <i>Ruminococcaceae</i> playing a crucial role
274	in its production and potentially influencing the thrombotic process [33]. Ruminococcaceae
275	metabolize dietary lipids, including choline, phosphatidylcholine, and L-alpha glyceryl
276	phosphorylcholine, into trimethylamine (TMA), which is subsequently oxidized to TMAO in the
277	liver. Elevated TMAO levels have been linked to endothelial dysfunction, platelet hyperreactivity,
278	and increased thrombosis risk [34]. Notably, Ruminococcaceae abundance is negatively correlated
279	with thrombosis formation, suggesting that a higher abundance of Ruminococcaceae may be
280	associated with a lower risk of VTE.
281	Additionally, a study by Huang et al. involving 33 patients with liver cirrhosis found a positive
282	correlation between Eubacterium hallii group and the occurrence of VTE. This bacterial group was
283	significantly enriched in patients with both liver cirrhosis and VTE, and its abundance was positively
284	associated with coagulation factor parameters [35]. These findings align with the results of our study.
285	Butyricimonas is a GM primarily known for producing butyrate, a key short-chain fatty acid with

286	anti-inflammatory properties that plays a crucial role in maintaining intestinal barrier function.
287	Disruption of inflammation and intestinal barrier integrity has been identified as a potential trigger
288	for VTE [36]. Therefore, it has been hypothesized that <i>Butyricimonas</i> may indirectly influence VTE
289	risk by modulating inflammation or preserving gut health. A reduction in <i>Butyricimonas</i> abundance
290	could theoretically promote thrombosis through inflammatory pathways. However, our findings
291	contradict this assumption, suggesting that the association between Butyricimonas and VTE may
292	involve more complex underlying mechanisms. Dorea belongs to the Firmicutesphylum and is
293	frequently associated with metabolic diseases such as obesity and diabetes, as well as inflammatory
294	conditions, all of which are important risk factors for VTE [37]. In cardiovascular disease-related
295	studies, changes in Dorea abundance have been linked to inflammation and metabolic dysregulation,
296	potentially influencing VTE risk by activating coagulation pathways or increasing blood viscosity
297	[38]. If an increase in <i>Dorea</i> abundance is associated with exacerbated inflammation or metabolic
298	disturbances, it may indirectly elevate the risk of VTE through these mechanisms. However, Dorea
299	may also contribute to reducing VTE risk by maintaining GM stability or supporting
300	anti-inflammatory effects. Therefore, the precise association between Dorea and VTE (whether
301	positive or negative) requires further investigation and validation. Currently, no clear research has
302	explored the association between Candidatus Soleaferrea and VTE risk, highlighting a potential
303	direction for future studies.
304	The GM may play a crucial role in the occurrence and progression of PE through the regulation
305	of the "gut-lung axis". Wu et al. demonstrated that certain drugs could alleviate pulmonary

306 inflammation in mice by modulating Intestinimonas, and this inflammatory response is closely

307	associated with thrombosis risk. It has been hypothesized that an increase in Intestinimonas
308	abundance may exert a protective effect against PE [39]. Further research has revealed that
309	Intestinimonas influences pulmonary inflammation and immune responses through the "gut-lung
310	axis", contributing to immune homeostasis in the lungs [40]. Conversely, a decrease in
311	Intestinimonas abundance may exacerbate systemic inflammation, thereby increasing the risk of PE.
312	Our findings are consistent with this evidence. <i>Veillonella</i> is an anaerobic bacterium that may play a
313	crucial role in PE formation. A case report described a 38-year-old female who developed Lemierre's
314	syndrome following a throat infection, with imaging revealing thrombosis in the jugular and
315	subclavian veins, accompanied by systemic complications. Blood culture identified Veillonella
316	<i>parvula</i> , suggesting that this bacterium may contribute to the formation of infectious thrombosis [41]
317	This finding also supports our study conclusions. Additionally, recent studies have found that the GM
318	composition in patients with idiopathic pulmonary arterial hypertension differs from that of healthy
319	controls, with <i>Firmicutes</i> exhibiting the highest abundance at 53.16% and 57.08%, respectively
320	[42].Another study demonstrated that cryptotanshinone alleviates pulmonary fibrosis by modulating
321	GM and bile acid metabolism, significantly reducing the proportion of Erysipelatoclostridium,
322	suggesting its potential involvement in pulmonary diseases [43]. However, the specific association
323	between Erysipelatoclostridium and PE requires further investigation. Regarding Unknown genus
324	and Lentisphaerae, there is a lack of research on their association with PE in the existing literature,
325	and no direct evidence currently supports their association with PE.
326	Research on the role of GM in DVT remains in its early stages. A metabolomics study identified
327	altered metabolic profiles in DVT patients, suggesting that GM may contribute to DVT pathogenesis

328 [44]. Studies indicated that *Collinsella* is enriched in DVT patients with myelofibrosis, producing

- 329 short-chain fatty acids and other metabolites that influence host metabolism and immune function
- 330 [45]. This underscores the significance of GM in DVT progression and its potential as a diagnostic
- and therapeutic target. However, no studies have directly reported associations between *Mollicutes*,
- 332 *Actinobacteria, Bifidobacteriaceae, Adlercreutzia, Desulfovibrio, and DVT.*

333 With aging, GM diversity increases, and its composition and function tend to stabilize. The 334 dominant bacterial phyla in the adult gut include Bacteroidetes, Firmicutes, Actinobacteria, and 335 Proteobacteria. These active microbial communities play a crucial role in carbohydrate metabolism, energy production, cell component synthesis, nutrient processing, and immune system development 336 337 [46]. However, the vast diversity of GM poses significant challenges for comprehensive 338 measurement and quality control. Additionally, the effects of the same GM taxa may vary across 339 different diseases. Therefore, this study employs MR analysis to investigate the association between GM and the risk of VTE, PE, and DVT. The strategy of targeting the GM as a host regulatory factor 340 341 has gradually attracted attention in emerging therapeutic approaches for chronic diseases. These 342 approaches include fecal microbiota transplantation, probiotic supplementation, dietary interventions, targeted use of antibiotics, and inhibition of specific microbial enzymes [47-51]. GM is also closely 343 344 linked to intestinal inflammation. However, most supporting evidence for these associations is 345 indirect, necessitating further direct experiments, modeling studies, and comprehensive investigations for validation. 346

Our study offers several advantages. It is among the few MR analyses that investigate the causal
association between GM and VTE. We utilized extensive GWAS data from multiple databases,

ensuring high-quality instrumental variables with F > 10, thereby reducing the risk of weak instrumental bias and enhancing explanatory power. However, certain limitations should be acknowledged. The GWAS data primarily involve individuals of European descent, which may restrict the generalizability of our findings to other ethnic groups. Additionally, most GWAS studies employ 16S rRNA gene sequencing at the genus level, preventing precise association of specific strains or species with our findings. Moreover, due to limitations in the outcomes database, the phenotypes discussed do not completely encompass all types of VTE, PE, and DVT, thereby somewhat reducing the clinical relevance and interpretative depth of our conclusions. Future large-scale clinical trials and cohort studies are necessary to further validate our findings.

373 Conclusion

In summary, specific GM exhibit a clear causal relationship with the development of VTE, PE, and DVT. Ruminococcaceae was found to significantly reduce the risk of VTE. This study enhances the understanding of the role of GM in VTE pathogenesis, particularly in the potential mechanisms of the microbiota-immune-coagulation network. Moreover, it provides important theoretical support for the development of innovative therapeutic approaches based on probiotics or microbiota transplantation.

- 380
- 381
- 382
- 383

384 Figure Legends



385 386

Figure1. Two-sample MR directed acyclic graph of the 3 key assumptions



Figure2. The results of MR analysis reveal the association between GM and VTE, PE, and DVT





Figure4. Leave-one-out assessing the impact of individual SNPs on the overall causal estimate





Figure 5. Funnel Plots of MR Analysis among VTE, PE, and DVT



Figure6. Forest Plots of MR Analysis between GM and VTE, PE, and DVT

400 **References**

- 401 [1] Al Raizah A, Alrizah M. Artificial intelligence in thrombosis: transformative
- 402 potential and emerging challenges. Thromb J,2025,23(1):2.
- 403 [2] Xia Y Q, Tang L, Hu Y. [Advances in the genetics of venous thromboembolic
- 404 disease]. Zhonghua Xue Ye Xue Za Zhi,2024,45(12):1144-1147.
- 405 [3] Saad M, Batool R M, Waqas S A, et al. Unveiling the trends: Growing cancer and
- 406 venous thromboembolism mortality in older adults in the United States, 1999-2020.
- 407 Thromb Res,2025,247:109259.
- 408 [4] Opitz C F, Meyer F J. Pulmonary Embolism: An Update Based on the Revised
- 409 AWMF-S2k Guideline. Hamostaseologie,2024,44(2):111-118.
- 410 [5] Navarrete S, Solar C, Tapia R, et al. Pathophysiology of deep vein thrombosis.
- 411 Clin Exp Med,2023,23(3):645-654.
- 412 [6] Szymanski K, Weber C, Daugherty K, et al. A review of venous
- 413 thromboembolism for the hospitalist. Postgrad Med,2025:1-8.
- 414 [7] Wang X, Zhang C, Pan M, et al. Design and rationale of the multicenter
- 415 randomized clinical trial (REVERSE): Efficacy and safety of rivaroxaban in the early
- 416 postoperative period for patients with bioprosthetic valve replacement or valve repair.
- 417 Int J Cardiol,2025:133023.
- 418 [8] Zeng J, Feng J, Luo Y, et al. Inflammatory Biomarkers as Predictors of
- 419 Symptomatic Venous Thromboembolism in Hospitalized Patients with AECOPD: A
- 420 Multicenter Cohort Study. J Atheroscler Thromb, 2024.

- 421 [9] Liu Q, Yang F, Kong K, et al. Potential causal relationships between blood
 422 metabolites, inflammatory cytokines, and venous thromboembolism. Front
 423 Immunol,2024,15:1445790.
- 424 [10] Dong Y, Zhang K, Wei J, et al. Gut microbiota-derived short-chain fatty acids
- 425 regulate gastrointestinal tumor immunity: a novel therapeutic strategy? Front426 Immunol,2023,14:1158200.
- 427 [11] Larsson S C, Butterworth A S, Burgess S. Mendelian randomization for
 428 cardiovascular diseases: principles and applications. Eur Heart
 429 J,2023,44(47):4913-4924.
- [12] Wu Q, Li J, Sun X, et al. Multi-stage metabolomics and genetic analyses
 identified metabolite biomarkers of metabolic syndrome and their genetic
 determinants. EBioMedicine,2021,74:103707.
- [13] Xu S, Li X, Zhang S, et al. Oxidative stress gene expression, DNA methylation,
 and gut microbiota interaction trigger Crohn's disease: a multi-omics Mendelian
 randomization study. BMC Med,2023,21(1):179.
- 436 [14] Pasqualini J, Facchin S, Rinaldo A, et al. Emergent ecological patterns and
- 437 modelling of gut microbiomes in health and in disease. PLoS Comput438 Biol,2024,20(9):e1012482.
- 439 [15] Gong F, Zheng X, Zhao S, et al. Disseminated intravascular coagulation: cause,
- 440 molecular mechanism, diagnosis, and therapy. MedComm (2020),2025,6(2):e70058.
- 441 [16] Johnson T A, Mukhopadhyay S, Buzza M S, et al. Regulation of macrophage
- 442 fibrinolysis during venous thrombus resolution. Thromb Res,2024,243:109149.

- 443 [17] Birney E. Mendelian Randomization. Cold Spring Harb Perspect444 Med,2022,12(4).
- 445 [18] Song Q, Huang T, Song J, et al. Causal associations of body mass index and
- 446 waist-to-hip ratio with cardiometabolic traits among Chinese children: A Mendelian
- 447 randomization study. Nutr Metab Cardiovasc Dis,2020,30(9):1554-1563.
- 448 [19] Zhou J, Li Y, Lin Y, et al. The genetic causal association between hip or knee
- 449 osteoarthritis and frailty: a two-sample Mendelian randomization analysis. Arch Med
- 450 Sci,2024,20(3):938-946.
- 451 [20] Jiang R, Qu Q, Wang Z, et al. Association between air pollution and bone mineral
- 452 density: a Mendelian randomization study. Arch Med Sci,2024,20(4):1334-1338.
- 453 [21] Kurilshikov A, Medina-Gomez C, Bacigalupe R, et al. Large-scale association
- 454 analyses identify host factors influencing human gut microbiome composition. Nat
- 455 Genet,2021,53(2):156-165.
- 456 [22] Kamat M A, Blackshaw J A, Young R, et al. PhenoScanner V2: an expanded tool
- 457 for searching human genotype-phenotype associations.
 458 Bioinformatics,2019,35(22):4851-4853.
- 459 [23] Emdin C A, Khera A V, Kathiresan S. Mendelian Randomization.
 460 JAMA,2017,318(19):1925-1926.
- 461 [24] Bowden J, Del Greco M F, Minelli C, et al. A framework for the investigation of
- 462 pleiotropy in two-sample summary data Mendelian randomization. Stat463 Med,2017,36(11):1783-1802.
- 464 [25] Zhang Y, Li D, Zhu Z, et al. Evaluating the impact of metformin targets on the

- 465 risk of osteoarthritis: a mendelian randomization study. Osteoarthritis
 466 Cartilage,2022,30(11):1506-1514.
- 467 [26] Zhao J V, Schooling C M. Using Mendelian randomization study to assess the
- 468 renal effects of antihypertensive drugs. BMC Med,2021,19(1):79.
- 469 [27] Liu Z, Zhang H, Sun X, et al. Causal association between metabolites and
 470 age-related macular degeneration: a bidirectional two-sample mendelian
 471 randomization study. Hereditas,2024,161(1):51.
- 472 [28] Wu Y, Shen Z, Chen B, et al. Investigation of bidirectional causal association
- between temporomandibular disorders and five mental disorders. Arch OralBiol,2024,171:106169.
- 475 [29] Zhou C, Zhou Y, Ma W, et al. Revisiting Virchow's triad: exploring the cellular
- and molecular alterations in cerebral venous congestion. Cell Biosci,2024,14(1):131.
- 477 [30] Lv K, Chen S, Xu X, et al. Protein disulfide isomerase cleaves allosteric
 478 disulfides in histidine-rich glycoprotein to regulate thrombosis. Nat
 479 Commun,2024,15(1):3129.
- 480 [31] Gong D, Zhang L, Zhang Y, et al. Gut Microbial Metabolite Trimethylamine
- 481 N-Oxide Is Related to Thrombus Formation in Atrial Fibrillation Patients. Am J Med
 482 Sci,2019,358(6):422-428.
- 483 [32] Reiner M F, Muller D, Gobbato S, et al. Gut microbiota-dependent
- 484 trimethylamine-N-oxide (TMAO) shows a U-shaped association with mortality but
- 485 not with recurrent venous thromboembolism. Thromb Res,2019,174:40-47.
- 486 [33] Papa A, Santini P, De Lucia S S, et al. Gut dysbiosis-related thrombosis in

- 487 inflammatory bowel disease: Potential disease mechanisms and emerging therapeutic488 strategies. Thromb Res,2023,232:77-88.
- 489 [34] Jonsson A L, Backhed F. Role of gut microbiota in atherosclerosis. Nat Rev
- 490 Cardiol,2017,14(2):79-87.
- 491 [35] Huang X, Zhang Y, Yi S, et al. Potential contribution of the gut microbiota to the
- 492 development of portal vein thrombosis in liver cirrhosis. Front493 Microbiol,2023,14:1217338.
- 494 [36] Lee H, An J, Kim J, et al. A Novel Bacterium, Butyricimonas virosa, Preventing
- 495 HFD-Induced Diabetes and Metabolic Disorders in Mice via GLP-1 Receptor. Front
- 496 Microbiol,2022,13:858192.
- 497 [37] Tsai Y, Tai W, Liang C, et al. Alternations of the gut microbiota and the
- 498 Firmicutes/Bacteroidetes ratio after biologic treatment in inflammatory bowel disease.
- 499 J Microbiol Immunol Infect,2024.
- 500 [38] Mi H T N, Chaiyasarn S, Kim H, et al. C-Glycoside-Metabolizing Human Gut
- 501 Bacterium, Dorea sp. MRG-IFC3. J Microbiol Biotechnol, 2023, 33(12):1606-1614.
- 502 [39] Wu Y, Chen Y, Li Q, et al. Tetrahydrocurcumin alleviates allergic airway
- 503 inflammation in asthmatic mice by modulating the gut microbiota. Food504 Funct,2021,12(15):6830-6840.
- 505 [40] Wang L, Cai Y, Garssen J, et al. The Bidirectional Gut-Lung Axis in Chronic
- 506 Obstructive Pulmonary Disease. Am J Respir Crit Care Med, 2023, 207(9):1145-1160.
- 507 [41] Montatore M, Zagaria A, Masino F, et al. A Rare Case of Lemierre's Syndrome
- 508 due to Veillonella Parvula: A Dangerous and Forgotten Complication of a Septic

- 509 Condition. Indian J Otolaryngol Head Neck Surg, 2024, 76(4): 3570-3575.
- 510 [42] Li J, Liu C, Xu Y, et al. Gut Microbiota Alterations in Adolescent Idiopathic
- 511 Scoliosis Are Associated with Aberrant Bone Homeostasis. Orthop512 Surg,2024,16(4):965-975.
- 513 [43] Li T, Chu Y, Yan K, et al. Simultaneous determination of tanshinol, 514 protocatechuic aldehyde, protocatechuic acid, notoginsenoside R1, ginsenoside Rg1 515 and Rb1 in rat plasma by LC-MS/MS and its application. Biomed 516 Chromatogr,2017,31(6).
- 517 [44] Cao J, An G, Li R, et al. Novel Strategy for Human Deep Vein Thrombosis
- 518 Diagnosis Based on Metabolomics and Stacking Machine Learning. Anal 519 Chem,2024,96(36):14560-14570.
- 520 [45] Barone M, Barone M, Ricci F, et al. A Specific Host/Microbial Signature of
 521 Plasma-Derived Extracellular Vesicles Is Associated to Thrombosis and Marrow
- 522 Fibrosis in Polycythemia Vera. Cancers (Basel),2021,13(19).
- 523 [46] An J, Kwon H, Kim Y J. The Firmicutes/Bacteroidetes Ratio as a Risk Factor of
 524 Breast Cancer. J Clin Med,2023,12(6).
- 525 [47] Rafie E, Zugman M, Pal S K, et al. What Is the Role of Fecal Microbiota
- 526 Transplantation in Immunotherapy Trials? Current Perspectives and Future Directions.
- 527 Eur Urol Focus,2025.
- 528 [48] Tiwari S, Paramanik V. Role of Probiotics in Depression: Connecting Dots of
- 529 Gut-Brain-Axis Through Hypothalamic-Pituitary Adrenal Axis and
- 530 Tryptophan/Kynurenic Pathway involving Indoleamine-2,3-dioxygenase. Mol

- 531 Neurobiol,2025.
- 532 [49] Yu J, Wu Y, Zhu Z, et al. The impact of dietary patterns on gut microbiota for the
- 533 primary and secondary prevention of cardiovascular disease: a systematic review.
- 534 Nutr J,2025,24(1):17.
- 535 [50] Almeida-Santos A C, Duarte B, Tedim A P, et al. The healthy human gut can take
- 536 it all: vancomycin-variable, linezolid-resistant strains and specific bacteriocin-species
- 537 interplay in Enterococcus spp. Appl Environ Microbiol,2025,91(1):e169924.
- 538 [51] Abdullah, Ahmad N, Xiao J, et al. Gingerols: Preparation, encapsulation, and
- 539 bioactivities focusing gut microbiome modulation and attenuation of disease
- 540 symptoms. Phytomedicine,2025,136:156352.
- 541





FIGURE 1: Two-sample MR directed acyclic graph of the three key assumption.



Figure2. The results of MR analysis reveal the association between GM and VTE, PE, and DVT



Figure3. Scatter Plots of MR Effect size for causal association between gut microbiota and VTE, PE, and DVT



Figure4. Leave-one-out assessing the impact of individual SNPs on the overall causal estimate



Figure5. Funnel Plots of MR Analysis among VTE, PE, and DVT



Figure6. Forest Plots of MR Analysis between GM and VTE, PE, and DVT