

Exploring causal correlations between inflammatory response-related genes and osteoporosis: a Multi-Omics Mendelian Randomization Study

Ripeng Zhang, Jie Zhao*

Affiliated Hospital of Shandong University of Traditional Chinese Medicine, Jinan, China

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***Corresponding author:**

Jie Zhao

Affiliated Hospital of
Shandong University
of Traditional Chinese
Medicine
Jinan, China

E-mail:

jee18765800627@163.com

Abstract

Introduction: The relationship between the inflammatory response (IR) and osteoporosis (OP) has been the subject of extensive research; however, their genetic link remains unclear. This study used IR-related genes as instrumental variables (IVs) to represent IR, while summary data of OP served as the outcome to explore their genetic relationship.

Material and methods: IR-related genes were retrieved from the GeneCards database. OP transcriptome datasets were collected from the Gene Expression Omnibus (GEO) database and meta-analyzed to identify differentially expressed genes (DEGs) related to IR in OP. Genetic proxy instruments for IR-related genes were derived from studies of corresponding gene expression ($n = 31,684$) and DNA methylation ($n = 1,980$) quantitative trait loci (eQTLs and mQTLs), respectively. Aggregated data for OP (1,351 OP cases and 209,313 controls) were extracted from the largest genome-wide association study (GWAS) of OP. We integrated QTL data with OP GWAS data to estimate their genetic associations using summary data-based Mendelian randomization analysis (SMR). Additionally, Bayesian colocalization analysis was employed to reveal the potential relationships between IR gene expression and inflammatory factors, as well as various hormones. Finally, to further validate whether the statistical evidence provided in GWAS comprised true-positive findings, a replication study (1,955 cases and 278,169 controls) was conducted here through genotype-phenotype associations.

Results: A meta-analysis of four datasets identified 115 IR-related DEGs in OP out of 612 IR-related genes. Through SMR analysis, we found that the expression levels of two IR-related genes were associated with OP risk. Specifically, elevated gene expression levels of FAS (odds ratio (OR) = 1.094; 95% confidence interval (CI) = 0.892–1.341; false discovery rate (FDR) = 0.034) increased the risk of OP. Conversely, increased expression levels of CHUK decreased the risk of OP (OR = 0.518; 95% CI = 0.424–0.637; FDR = 0.039). Colocalization analysis identified potential interactions between the FAS gene and estradiol (PP.H4 = 0.95) as well as interleukin-1 α (IL-1 α) (PP.H4 = 0.65). Potential interactions were also observed between the CHUK gene and growth hormone (PP.H4 = 0.59) as well as macrophage inflammatory protein-1 α (MIP-1 α) (PP.H4 = 0.62). In addition, consistent results were observed in the replication study, further demonstrating the reliability of our findings.

Conclusions: This multi-omics integration study revealed a genetic link between IR and OP, as represented by IR-related genes, and provided new insights into the potential pathogenic mechanisms of OP. Additionally, these identified candidate genes offer avenues for future targeted functional studies aimed at developing appropriate therapeutic interventions and preventing OP.

Key words: inflammatory response, osteoporosis, integrative omics, Mendelian randomization.

Introduction

Osteoporosis (OP) is a chronic systemic disease characterized by a decrease in bone density and mass, increasing the risk of fractures and significantly impacting patients' quality of life while also elevating the socioeconomic burden [1]. The number of individuals at high risk for osteoporotic fractures is rising worldwide, with over 9 million people experiencing osteoporotic fractures each year [2]. Although previous studies have investigated various aspects of the pathogenesis of OP, such as environmental factors, immune responses, nutritional status, medications, and epigenetic changes [3], the underlying mechanism is not fully understood. Therefore, further research on the genetic mechanisms is necessary to deepen our understanding of the pathogenesis of OP.

Bone is a tissue that undergoes constant remodeling, involving both formation and destruction. This process is tightly regulated and can be disrupted by various factors, including hormonal changes. Furthermore, inflammation disrupts bone metabolism and promotes bone resorption. An increasing body of research on bone remodeling processes suggests that IR plays a significant role in the development of OP [4]. Inflammation is characterized by the overproduction of cytokines such as tumor necrosis factor (TNF), IL-1, IL-6, and IL-17. The overexpression of these cytokines during inflammation leads to excessive bone resorption, primarily through the overactivation of osteoclasts. Additionally, certain cytokines can impair osteoblast function. In inflammatory conditions, bone resorption is enhanced by the activation of inflammatory cells and mediators, leading to compromised balance in bone remodeling [5]. This condition may also adversely affect the skeletal system, leading to OP, fractures, and other bone-related diseases. In concrete terms, activated T cells specifically secrete a significant amount of the osteoclastogenic factor, which is encoded by the threonine synthase-like 2 gene homolog [6]. This factor is released and directly stimulates the formation of osteoclasts, resulting in bone resorption and damage, ultimately accelerating the progression of OP. Amarasekara *et al.* found that the IR gene regulates the metabolism of osteocyte populations [7]. Zhang *et al.* discovered that inhibiting inflammatory pathways can increase bone mass by preventing osteoclast-mediated bone resorption [8]. Moreover, numerous studies have shown that IR-related genes, such as IL-10 and nucleotide-binding oligomerization domain 2 (NOD2), are related to OP [9, 10]. IR is a complex physiologic process that involves multiple cytokine responses and gene expression. Although the promotion of OP by IR has been extensively studied, the specific mechanism remains unclear, necessitating further research.

Mendelian randomization (MR) is a powerful method for determining causal relationships between exposures and outcomes, using genetic variation as IVs [11]. This methodology is notably accurate and dependable, as it effectively mitigates the influence of confounding factors and minimizes the risks of reverse causation [12]. However, the intricate structure of linkage disequilibrium (LD) within the genome may lead to closely related variants that are not necessarily causally associated [13]. Furthermore, these genetic variants have the potential to influence DNA methylation (DNAm), gene expression, hormone levels, and the levels of inflammatory factors. Consequently, multi-omics integration has emerged as a promising approach in the post-GWAS era for identifying critical regulators and exploring therapeutic targets in OP. For instance, building on the MR analysis, the SMR method was introduced to enhance the inference of genetic associations between various molecular traits (such as gene expression, DNAm, or protein abundance) and relevant diseases [14]. Compared with traditional MR analyses, SMR, using the top cis-QTL as the exposure, can achieve greater statistical power when the exposure and outcome are derived from two independent samples with substantial sample sizes [14]. Up to now, limited MR studies have identified the potential genetic association between IR and OP. Therefore, this study aimed to explore the genetic relationship between IR proxied by IR-related genes and OP using multi-omics data (including eQTL data and mQTL data).

Material and methods

Ethical considerations

Data used in our study were publicly available, and further ethical approval was not required.

Study design

Figure 1 presents the design and workflow of this study. Initially, 612 genes with a relevance score of 15 or higher were identified as IR genes by retrieval from the GeneCards database (v5.10, <https://www.genecards.org>) using “inflammatory response” as the keyword. Four OP transcriptome datasets were found in the GEO database [15–17] and meta-analyzed to identify DEGs related to IR in OP. Then, IVs of the IR-related genes extracted from eQTLs and mQTLs were used as proxies for IR. Their association with OP was investigated through SMR analysis. Furthermore, to enhance the reliability of our findings, we conducted the heterogeneity in dependent instruments (HEIDI) test and Bayesian colocalization analysis. Only IR-related genes that passed both the HEIDI test and colocalization analysis were deemed candi-

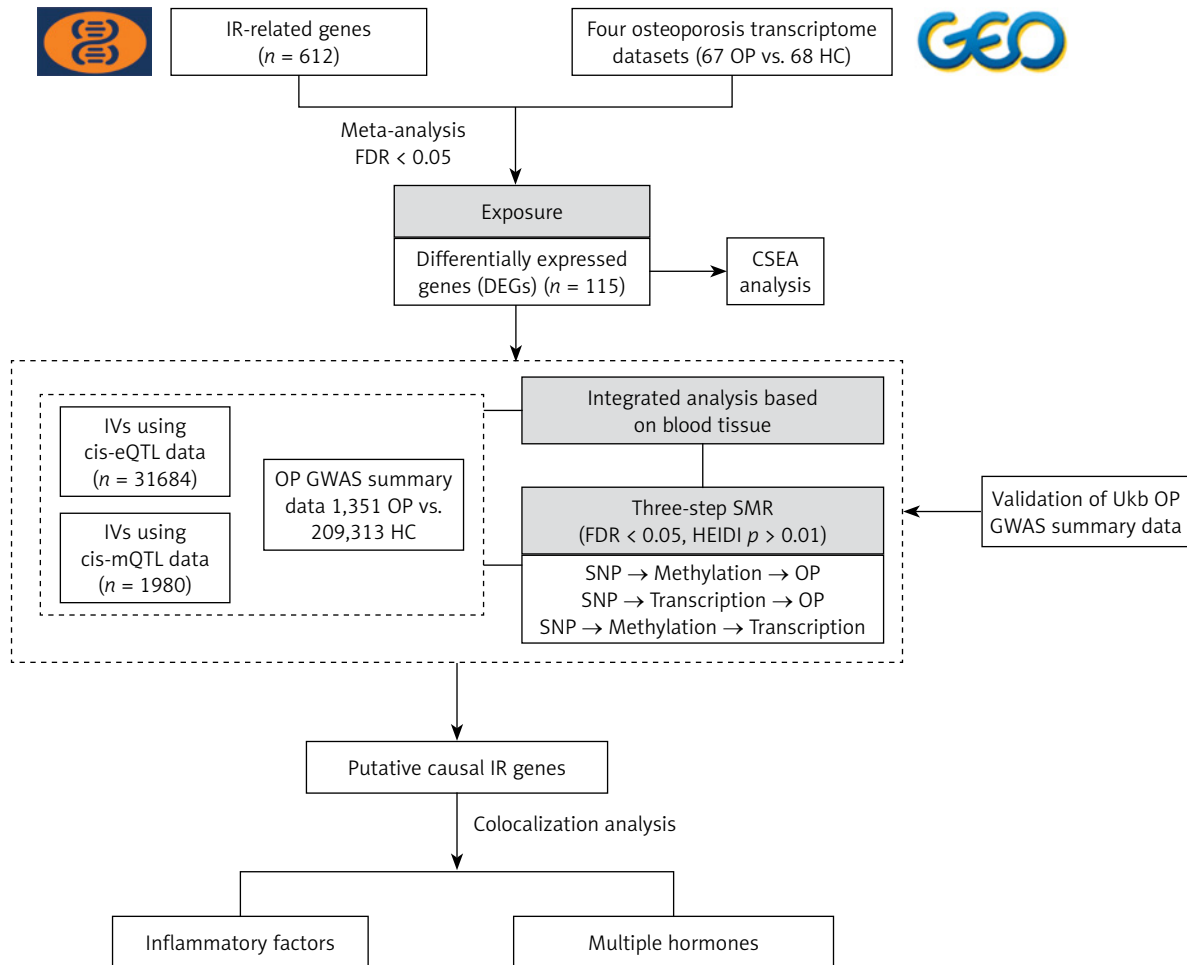


Figure 1. Workflow of the study. A series of analyses were performed to identify candidate causal IR genes associated with OP onset. IR-related genes were sourced from the GeneCards database. Transcriptome datasets including OP cases and controls (HC) were obtained from the GEO database, and meta-analysis was conducted to identify DEGs, followed by cell type-specific expression analysis (CSEA). Integration of GWAS summary and cis-eQTLs/cis-mQTLs data from the blood used three-step SMR methods, which prioritized putative blood IR genes and their regulatory elements associated with the risk of OP ($FDR < 0.05$; $P_{HEIDI} > 0.01$). The UK Biobank database was used to validate the aforementioned genes and their regulatory components. Additionally, colocalization analysis was conducted to reveal potential interactions between the gene expression of presumed causal genes and inflammatory factors as well as various hormones ($PP.H4 > 0.5$).

IR – inflammatory response, OP – osteoporosis, GEO – Gene Expression Omnibus, DEG – differentially expressed genes, eQTL – Gene expression quantitative trait locus, mQTL – DNA methylation quantitative trait locus, GWAS – genome-wide association study, SMR – summary data-based Mendelian randomization analysis, FDR – false discovery rate, HEIDI – heterogeneity in dependent instruments.

date genes for OP. In the present study, summarized GWAS data of OP from both the FinnGen and the UK Biobank were used as the primary discovery dataset, while summarized GWAS data of OP from the UK Biobank were used as the replication dataset to validate our results. Finally, colocalization analysis was performed to evaluate the shared genetic effects between blood gene expression and inflammatory factors as well as various hormones.

Data sources

The eQTL IVs of IR-related genes were sourced from eQTLGen and included the genetic data of

blood gene expression in 31,684 individuals derived from 37 datasets [18]. The mQTL IVs of IR-related genes were generated from a meta-analysis of two European cohorts: the Brisbane Systems Genetics Study ($n = 614$) and the Lothian Birth Cohorts ($n = 1366$) [13]. The current study focused only on cis-eQTLs and cis-mQTLs, which constituted single-nucleotide polymorphisms (SNPs) within a 1 Mb distance from the start and end of the gene ($FDR < 0.05$).

GWAS data of OP for the discovery study were obtained by integrating two databases: FinnGen and UK Biobank. This summary data for OP included 8 international cohorts and 210,664 individu-

als (1,351 cases and 209,313 controls) [19, 20]. To validate our findings, we used different OP GWAS data (1,955 cases and 278,169 controls) provided by the UK Biobank for the replication study [19]. More data information is presented in Supplementary Table SXIII).

Statistical analysis

We used SMR analysis to investigate the genetic relationship between IR, as represented by IR-related genes, and OP. In the current SMR analysis, the most significantly associated top cis-QTLs of IR-related genes were chosen as IVs. Importantly, for specific IR-related genes that presented multiple significant cis-QTLs with identical *p*-values, we selected the cis-QTL with the highest absolute Z-score as the primary cis-QTL. Additionally, we checked the consistency of allele frequency of each SNP between pairwise datasets, including QTL data, GWAS data, and LD reference data. SNPs with allele frequency differences < 0.20 between any pair of the data sets were included. Furthermore, to minimize bias caused by heterogeneity, the HEIDI test was rationally employed. Of note, a *p*-value of HEIDI > 0.01 suggested no heterogeneity, implying that the result was reliable [21]. Finally, we used *FDR* correction to avoid false genetic association. Only IR-related cis-QTLs that passed *FDR* correction (P_{SMR} corrected by $FDR < 0.05$) and the HEIDI test ($P_{\text{HEIDI}} > 0.01$) were allowed to undergo further colocalization analyses. Both SMR analysis and HEIDI tests were performed using SMR software for Windows version 1.3.1.

Meta-analysis of DEGs

Four transcriptome datasets (GSE56815, GSE13850, GSE35958, and GSE7158) (67 cases and 68 controls) were obtained from the GEO database. A linear regression model was employed to identify DEGs. Confounding factors such as age, sex, and menopause may affect the results [22], so we chose them as covariables for correction. DEGs were analyzed individually in the four gene expression datasets and subsequently integrated through a fixed-effects meta-analysis using the R package metafor.

SMR and colocalization analysis

Multiple SMR tools were developed to determine whether the effects of SNPs on phenotypes are mediated by molecular features such as gene expression or DNAm. Colocalization analyses were performed to examine common variants that may be responsible for multiple traits; integration of GWAS data with other molecular QTL data through SMR and colocalization improved identification of causal SNPs via specific pathways.

Blood tissue analysis used the SMR multi-tool to determine the causal inference of IR genes and the 1000 Genomes European reference to calculate LD. A three-step SMR analysis was performed: (1) SNPs were instruments, blood gene expression was exposure, and OP was the outcome; (2) SNPs were instruments, blood DNAm was exposure, and OP was the outcome; (3) SNPs were instruments, blood DNAm was exposure, and blood gene expression was the outcome. The third step included only significant signals from steps 1 and 2. The final candidate signals were defined as those that (1) passed the three-step SMR analysis ($FDR < 0.05$); (2) were suggestively significant genome-wide ($p < 1 \times 10^{-5}$) in all eQTLs and mQTLs; and (3) had no HEIDI test results, with $p > 0.01$.

We selected colocalization analysis to assess the potential interactions between the expression of IR genes and inflammatory factors as well as various hormones in blood tissues. This method is used to assess the presence of a shared causal variant in the region for two traits. Analysis was performed using the coloc R package with $PP_{H4} > 0.5$ as the threshold for the shared genetic effects between the two traits. The sources of GWAS data for inflammatory factors and hormones can be found in the Supplementary File.

Cell type-specific enrichment and regulatory component annotation

The Cell type-Specific Enrichment Analysis Database (CSEA-DB, <https://bioinformatics.fo.uth.edu/CSEADB/>) was used to investigate whether DEGs are specific to any cell type. We focused on cell types present in blood, bone, and muscles.

The regulatory signature enrichment of DNAm sites was assessed using eFORGE (<http://eforge.cs.ucl.ac.uk/>), including chromatin status (active and inactive) and histone marker (H3K4me1) annotation. The regions of individual DNAm sites were annotated at <http://grch37.ensembl.org/>.

Results

Meta-analysis of differentially expressed IR-related genes between patients with OP and HC

Using the linear regression model, we individually analyzed four OP transcriptome datasets (GSE56815, GSE13850, GSE35958, and GSE7158) (67 OP cases and 68 controls) (Supplementary Table SI) to identify DEGs in patients with OP. Meanwhile, 612 IR genes with a correlation score ≥ 15 were obtained from the GeneCards database (Methods section; Supplementary Table SII). A meta-analysis was conducted to identify 115 differentially expressed IR-related genes in patients with OP ($p < 0.05$; Figure 2 A, Supplementary Ta-

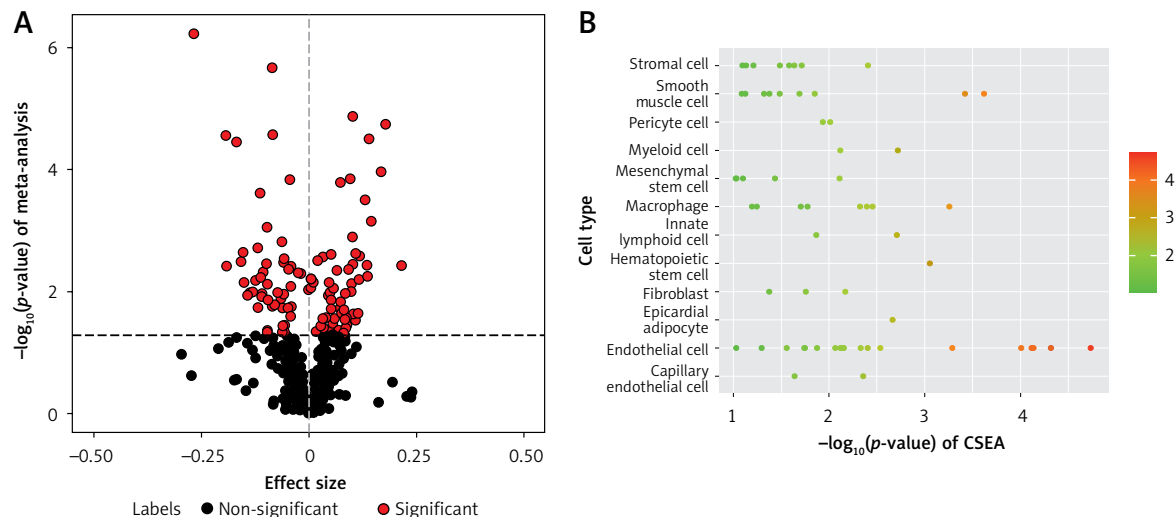


Figure 2. Meta-analysis of four gene expression datasets between patients with OP and HC. **A** – In total, 115 of 612 genes presented in all four OP transcriptome data were assessed for expression differences between patients with OP and HC. The volcano plot shows the meta effect sizes on the x-axis, while the y-axis indicates the $-\log_{10}$ -transformed meta p -values. Red dots represent 116 significant DEGs, and black dots represent non-significantly expressed genes. The dashed line indicates the significance threshold with $p < 0.05$ corrected for the number of gene tests. **B** – The Cell type-Specific Enrichment Analysis Database was used to investigate whether DEGs were specific to any cell type. The y-axis indicates the cell types derived from the blood. Dots represent 12 cell types. The dashed line is the significance threshold with $p < 0.05$.

OP – osteoporosis, HC – healthy controls, DEG – differentially expressed genes.

ble SIII). In addition, we conducted CSEA for 115 DEGs. Among the 12 enriched cell types, we found significant enrichment of IR genes in smooth muscle cells (Figure 2 B, Supplementary Table SIV).

Integration of OP GWAS and IR-related eQTL/mQTL data from the blood

A three-step SMR method was used, and only the significant results in all three SMR analyses that passed sensitivity checks were interpreted as suggestive causal genes (“Methods” section). A total of 115 IR-related DEG cis-eQTLs and their cis-mQTLs were integrated with the largest available GWAS summary statistics for OP.

In the first step, 13 IR-related genes ($FDR < 0.05$, $P_{\text{HEIDI}} > 0.01$) were identified by integrating blood eQTLGen data ($n = 31,684$) with OP GWAS data (Supplementary Table SV). In the second step, we identified 1638 DNAm probes by integrating the same OP GWAS results and mQTL summary statistics from the meta-analysis of Brisbane Systems Genetics Study and Lothian Birth Cohorts ($n = 1980$) (Supplementary Table SVI). In the third step, further integration analysis of putative OP-causal cis-eQTL and cis-mQTL data prioritized 7 DNAm probes potentially regulating 5 neighboring genes, namely, ARPC1B, FAS, CHUK, TRPC6, and TRPV1 ($FDR < 0.05$, $P_{\text{HEIDI}} > 0.01$) (Supplementary Table SVII). These CpG sites are significantly enriched in the transcription start sites of peripheral blood cells. Examples are (1) primary monocytes from peripheral blood ($p = 9.3 \times 10^{-3}$),

(2) primary B cells from peripheral blood ($p = 1.42 \times 10^{-3}$), and (3) primary T cells from cord blood ($p = 6.81 \times 10^{-3}$) (Supplementary Table SVIII). Evident enrichment of the transcription start sites was found in the three cell types.

Putative OP-causal genes mediated by blood methylation regulation of gene expression

Through a three-step SMR analysis, we first considered the FAS gene as a potential pathogenic gene. Significant SNP signals associated with the FAS gene were identified during OP GWAS, eQTL, and mQTL data analysis. The DNAm probe cg09221159 is located in the region 718 kbp upstream of the FAS gene. The methylation level in this region is negatively correlated with FAS gene expression ($\beta_{\text{SMR}} = -0.162$; $FDR = 0.002$; $P_{\text{HEIDI}} = 0.19$) and OP incidence ($\beta_{\text{SMR}} = -0.579$; $FDR = 0.021$; $P_{\text{HEIDI}} = 0.98$), while FAS gene expression is positively correlated with OP incidence ($\beta_{\text{SMR}} = 0.089$; $FDR = 0.034$; $P_{\text{HEIDI}} = 0.24$). Overall, the findings suggest that low DNAm levels of the FAS gene may upregulate its expression, thereby increasing the risk of OP (Figures 3 A, B).

Another gene deemed to be crucial is the CHUK gene (Figures 3 C, D). The DNAm probe cg03679755 is located in the region 73 kbp upstream of the CHUK gene. A higher level of methylation at this site would inhibit the transcription of the CHUK gene ($\beta_{\text{SMR}} = -0.031$; $FDR = 8.23 \times 10^{-5}$; $P_{\text{HEIDI}} = 0.16$), thereby increasing the risk

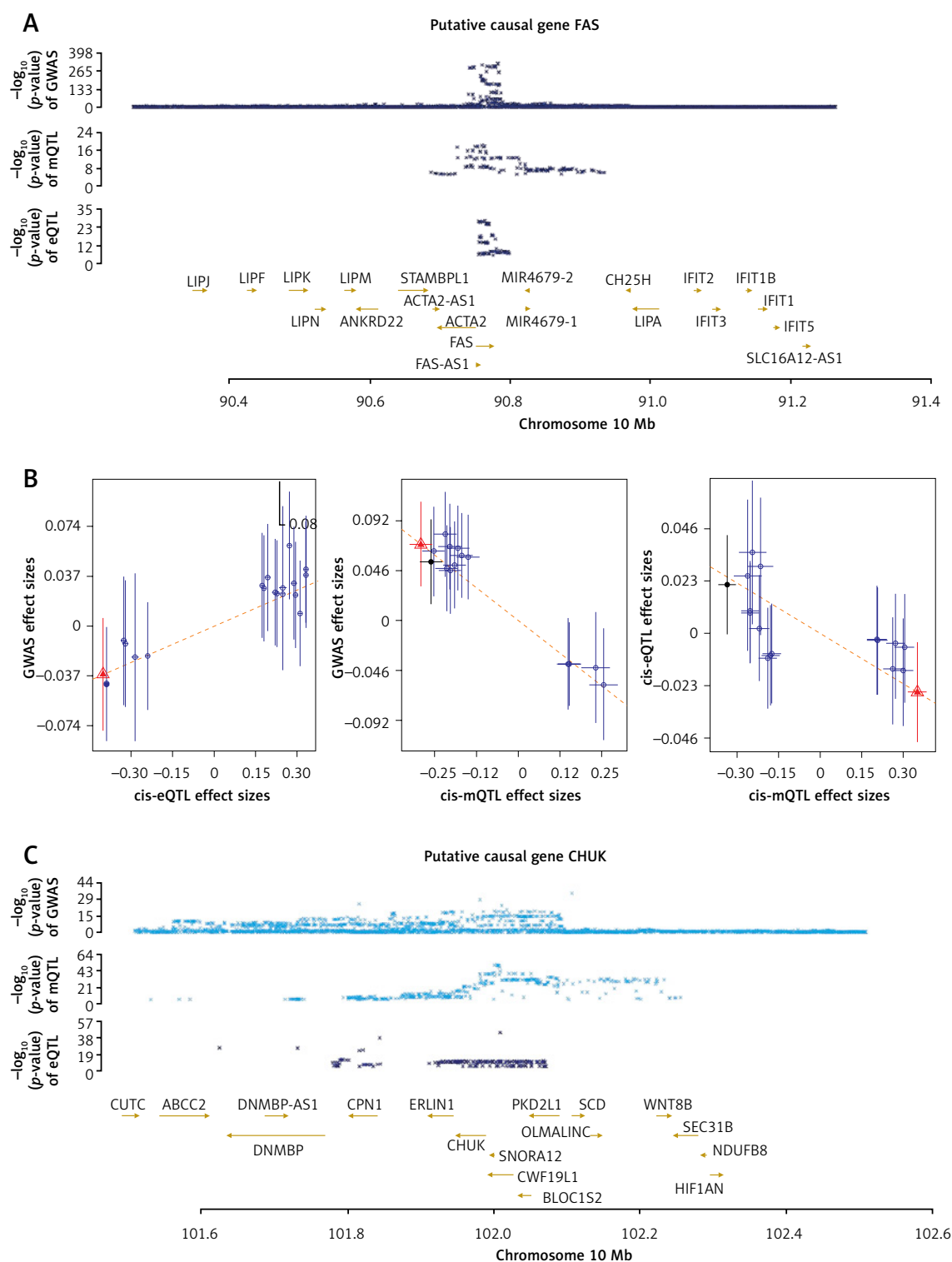


Figure 3. Three-step SMR analysis prioritized putative causal IR genes and mechanisms in OP using blood tissue. **A, C** – Locus zoom plots showing the consistent genetic effects from OP GWAS, cis-mQTL, and cis-eQTL near FAS and CHUK (from upper to lower panels, all minimum $p < 1 \times 10^{-5}$). **B, D** – Three-step SMR indicating significant causal relationships between gene expression and OP onset mediated by methylation ($FDR < 0.05$, $P_{HEIDI} > 0.01$). From left to right: SMR between gene expression and OP GWAS, SMR between gene methylation and OP GWAS, and SMR between gene methylation and expression

IR – inflammatory response, OP – osteoporosis, eQTL – gene expression quantitative trait locus, mQTL – DNA methylation quantitative trait locus, GWAS – genome-wide association study, SMR – summary data-based Mendelian randomization analysis, FDR – false discovery rate, HEIDI – heterogeneity in dependent instruments.

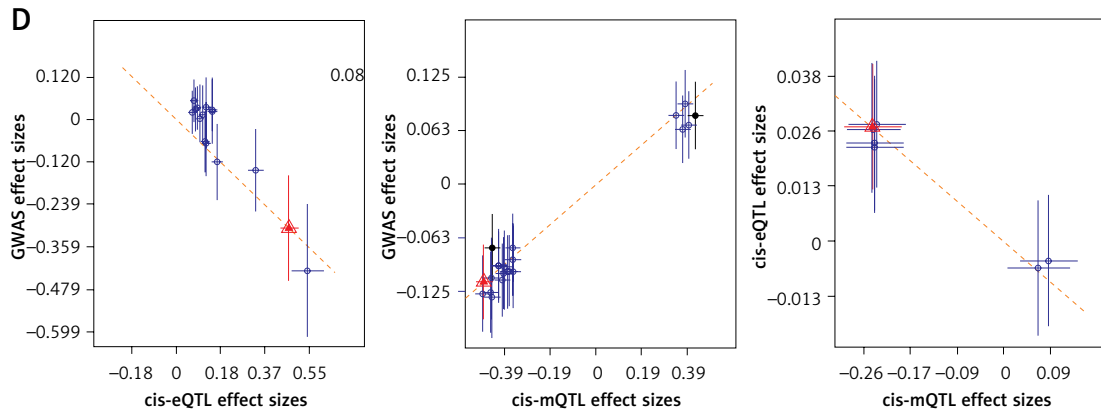


Figure 3. Cont.

IR – inflammatory response, OP – osteoporosis, eQTL – gene expression quantitative trait locus, mQTL – DNA methylation quantitative trait locus, GWAS – genome-wide association study, SMR – summary data-based Mendelian randomization analysis, FDR – false discovery rate, HEIDI – heterogeneity in dependent instruments.

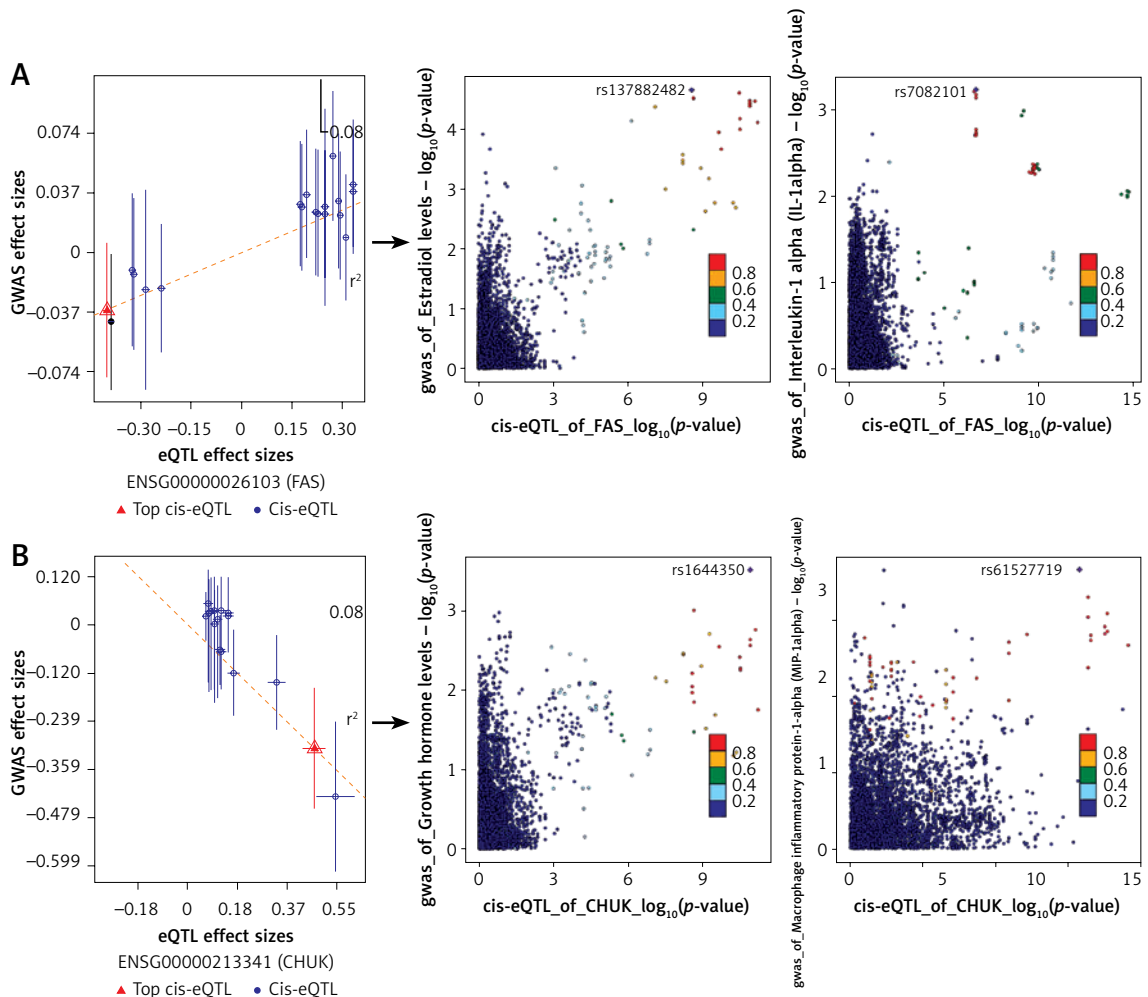


Figure 4. SMR and colocalization analyses prioritized blood causal IR genes and interactions with inflammatory factors and various hormones in OP. The left panels indicate the SMR between gene expression and OP GWAS (all SMR $FDR < 0.05$, $P_{HEIDI} > 0.01$), while the right panels show the locus comparisons between cis-eQTLs and mbQTLs by colocalization analysis (all $PP.H4 > 0.5$). The r^2 value indicates the LD between the variants and the top SNPs. **A, B** – the FAS and CHUK genes, respectively

IR – inflammatory response, OP – osteoporosis, eQTL – gene expression quantitative trait locus, mQTL – DNA methylation quantitative trait locus, GWAS – genome-wide association study, SMR – summary data-based Mendelian randomization analysis, FDR – false discovery rate, LD – linkage disequilibrium, HEIDI – heterogeneity in dependent instruments, SNPs – single-nucleotide polymorphisms.

of developing OP ($\beta_{\text{SMR}} = 0.011$; $FDR = 0.049$; $P_{\text{HEIDI}} = 0.07$). Conversely, a higher level of transcription of the CHUK gene would decrease the risk of developing the disease ($\beta_{\text{SMR}} = -0.656$; $FDR = 0.039$; $P_{\text{HEIDI}} = 0.16$).

Putative OP-causal genes involved in inflammatory factors and multiple hormone interactions

We classify $0.5 < PP.H4 < 0.8$ as moderate-strength colocalization evidence, and $PP.H4 > 0.8$ as high-strength colocalization evidence. Factors that meet the criteria for moderate-strength evidence include TNF-related activation cytokine, IL-1 α , interleukin-12 subunit beta, growth hormone levels, osteoprotegerin levels, free androgen index, and C-X-C motif chemokine 9. Factors that meet the criteria for high-strength evidence include T-cell surface glycoprotein CD5 and estradiol levels (to be interpreted with caution).

Based on SMR and colocalization analysis, we prioritized FAS as a candidate IR causal gene in OP associated with inflammatory factors and several hormones (Figure 4 A) (Supplementary Table SIX, SXI). Our findings indicate that increased expression of FAS likely contributes to the onset of OP ($\beta_{\text{SMR}} = 0.089$; $FDR = 0.034$; $P_{\text{HEIDI}} = 0.24$). Additionally, SNPs that regulate FAS expression may influence the levels of inflammatory cytokines and hormones, as observed in the colocalization analysis. The expression of the FAS gene facilitates the onset and progression of OP. After excluding hormones and inflammatory factors having weaker associations with bone metabolism, we found that both estrogen ($PP.H4 = 0.95$) and IL-1 α ($PP.H4 = 0.65$) exert a shared genetic influence on FAS gene expression (Supplementary Tables SX, SXII).

The CHUK gene (Figure 4 B) (Supplementary Tables SIX, SXI) is another significant gene that is closely associated with the risk of OP ($\beta_{\text{SMR}} = -0.656$; $FDR = 0.039$; $P_{\text{HEIDI}} = 0.16$). After excluding hormones and inflammatory factors having weaker associations with bone metabolism, we found that CHUK exhibits a shared genetic effect with growth hormone ($PP.H4 = 0.59$) and MIP-1 α ($PP.H4 = 0.62$) (Supplementary Table SX, SXII). Consequently, genetic variations in the CHUK gene may modulate its expression, influence hormone levels, and alter inflammatory factor levels, all of which are closely associated with the risk of OP.

Discussion

To our knowledge, this is the first study to identify the putative IR-causing genes and underlying mechanisms in OP from blood tissue samples using a multi-omics integrated approach. We identified 115 IR-related DEGs in OP from a pool of

612 potential IR-related genes, based on a comprehensive meta-analysis of transcriptome data. Using large-scale QTL data and GWAS summary data, we conducted SMR analysis to investigate the association between IR genes and the risk of OP. By integrating the SMR analysis with the HEIDI test, we found that the expression levels of two IR-related genes were associated with the risk of OP. Specifically, elevated expression levels of the FAS gene were associated with an increased risk of OP, while higher expression levels of the CHUK gene were linked to a decreased risk of OP. Additionally, colocalization analysis demonstrated that FAS shares genetic variants with estrogen and IL-1 α , whereas CHUK shares variants with growth hormone and MIP-1 α . Consequently, this study enhances our understanding of the causal relationship between IR genes and OP, elucidating the underlying genetic interaction mechanisms.

The human FAS gene/CD95 is located on chromosomal region 10q23.31 and encodes a mature protein composed of 319 amino acids [23]. As a member of the TNF receptor family, it plays a critical role in the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signaling pathway [24]. Previous studies have confirmed its significant role in human physiology. In recent decades, genetic variants of FAS have been associated with autoimmune diseases [25], osteoarthritis [26], and acute lung injury [27]. However, limited reports exist on the relationship between FAS and OP. In the present study, we found that high expression of the FAS gene is associated with an increased risk of OP. Additionally, we discovered that DNAm in the upstream region of FAS negatively regulates FAS expression, suggesting a connection between DNAm, FAS expression, and the risk of OP. The association can be reasonably explained through two possible mechanisms. First, FAS plays a key role in the receptor activator of nuclear factor kappaB (RANK)/RANK ligand (RANKL) signaling pathway [28], which promotes osteoclast differentiation and induces bone resorption [29]. Notably, bone loss is primarily regulated by the RANK/RANKL system, the main mechanism responsible for the induction, activation, and survival of osteoclasts [30, 31]. Some studies have demonstrated that inhibiting the RANKL/RANK axis can enhance bone mass by preventing osteoclast-mediated bone resorption [8]. AS2690168, a RANK/RANKL signaling pathway inhibitor, has been shown to alleviate pain associated with OP and slow its progression [32]. Furthermore, several tests have revealed that stimulating RANKL expression in osteocytes can increase bone resorption, resulting in bone loss in mice [33]. Secondly, FAS induces the process of apoptosis, which may be influenced by oxidative stress (OS) [29, 34]. OS leads to dysfunction of bone marrow mesenchy-

mal stem cells, resulting in decreased osteogenic ability and increased differentiation into adipose tissue [35]. Osteoblasts can produce antioxidants, such as glutathione peroxidase, to protect cells from damage caused by reactive oxygen species [36]. During bone resorption, osteoclasts generate superoxide, and OS enhances the differentiation and function of osteoclasts [37]. Melatonin is an antioxidant that has been shown to inhibit the generation of osteoclasts [38]. In *in vivo* experiments, melatonin supplementation has a positive effect on the bone mass of ovariectomized mice [39]. Other studies have shown that the FAS gene may promote the differentiation of bone marrow mesenchymal stem cells by regulating the expression of peroxisome proliferator-activated receptor gamma and fatty acid-binding protein 4, and inhibit lipid accumulation during the differentiation process, thereby increasing bone mass and maintaining the health of the bone microenvironment [40, 41]. In summary, most studies suggest that FAS may accelerate OP progression by promoting the RANK/RANKL signaling pathway or by inducing OS. Other studies indicate that the FAS gene could regulate the expression of peroxisome proliferator-activated receptor gamma and fatty acid-binding protein 4, promoting the differentiation of bone marrow mesenchymal stem cells and inhibiting lipid accumulation during the differentiation process, thereby increasing bone mass and slowing the progression of OP. This may explain why, despite the causal relationship between FAS and osteoporosis being statistically significant ($FDR < 0.05$), the 95% confidence interval of the OR still includes 1.

The CHUK gene encodes the inhibitor of kappa B kinase complex, a direct upstream activator of the NF- κ B signaling pathway. During mitochondrial stress, the NF- κ B essential modulator is recruited to the mitochondria. This recruitment triggers two processes: mitochondrial autophagy and reverse recruitment of the inhibitor of kappa B kinase complex, leading to activation of the NF- κ B signaling pathway [42]. Thus, CHUK may trigger an IR while maintaining bone homeostasis and reducing the risk of OP through the mitochondrial autophagy pathway. Previous research on CHUK has predominantly focused on tumors [43], developmental disorders [44] and inflammation-related diseases [45]. However, studies examining the relationship between CHUK and OP remain limited. This study suggests that elevated CHUK expression may correlate with a reduced risk of OP, while DNAm inhibits CHUK expression. Mitochondrial autophagy is widely observed in bone tissue. Mitochondria play a critical role in regulating intracellular calcium transport and ion homeostasis by providing adenosine triphosphate and facilitating calcium signaling [46]. In osteoblasts, mitochondria

secrete vesicles that promote bone formation, thereby influencing the osteogenic process [47]. Mitochondrial autophagy is essential for maintaining mitochondrial function. Recent studies have indicated that under conditions of chronic inflammation, inhibition of mitochondrial autophagy can activate the wntless-integrated/ β -catenin signaling pathway, which, in conjunction with Runt-related transcription factor 2 and bone morphogenetic proteins, triggers the differentiation of osteoblasts [48, 49]. Thus, mitophagy plays a pivotal role in bone remodeling. Research has demonstrated that activation of mitochondrial autophagy via inhibition of the mechanistic target of rapamycin/phosphoinositide 3-kinase pathway suppresses senescence in bone marrow mesenchymal stem cells and enhances their osteogenic function [50–52]. Additionally, research has shown that resveratrol protects osteoblast function by activating the sir-tuin 1-mediated mitochondrial autophagy pathway [53]. Furthermore, mitophagic cell death in osteoclasts may be associated with the expression of bone resorption-related proteins [54]. In conclusion, CHUK may impede the progression of osteoporotic conditions by regulating mitochondrial autophagy; however, further research is required to elucidate the specific mechanisms through which CHUK decelerates the development of OP.

By integrating multi-omics studies from various tissues, researchers can analyze GWAS signals to prioritize relevant genes and disease mechanisms. Our study identified the colocalization of genetic regulations of FAS expression and hormones, as well as inflammatory factors. Estradiol, IL-1 α , and FAS gene expression exhibit common genetic effects, indicating a potential interaction among this gene, hormones, and inflammatory factors. Estradiol can induce the apoptosis of pre-osteoclasts by regulating Fas ligand in osteoblasts, thereby maintaining bone homeostasis [55]. Additionally, estradiol intake has been shown to stimulate osteoblast proliferation and collagen gene expression, thereby slowing down the progression of OP in postmenopausal women [56, 57]. IL-1 α stimulates bone turnover by increasing bone resorption and blood calcium levels, exerting long-term local effects mediated by prostaglandins [58]. CHUK has common genetic effects with growth hormone and MIP-1 α . Growth hormone increases bone density, promotes bone formation, and enhances the biomechanical strength of fracture healing by stimulating osteoblast proliferation and activity, showing significant effects in the treatment of osteoporotic fractures [59]. MIP-1 α plays a critical role in bone remodeling and metabolism by promoting the formation and activity of osteoclasts [60]. The high expression in various bone destruction-related diseases makes it a potential target for bone disease treatment.

Tissue- and cell-specific gene expression has been shown to yield valuable insights into regulatory mechanisms and biological processes underlying disease. The site of onset for OP is in the bone, and studying the genetic influence on gene expression using eQTL data from bone tissue may be more meaningful. However, there is currently a lack of public databases related to bone tissue. This study conducted CSEA on 115 DEGs related to IR. Among the 12 enriched cell types, smooth muscle cells were significantly enriched. This finding suggests that smooth muscle cells may be involved in the regulation of skeletal inflammation. Previous studies have found that smooth muscle cells may influence bone metabolism by altering the bone microenvironment [61]. Further exploration is needed into the mechanisms between IR-related genes and OP, and future studies could utilize public databases on smooth muscle tissue for in-depth analysis.

Some limitations of this study are worth acknowledging. First, the dataset used in our research integrates four OP transcriptome datasets, and the quality and measurement errors of different data sources may not be consistent, which could potentially affect the reliability of the results. Therefore, we conducted linear regression model analysis on each dataset separately and adjusted for potential confounding factors that could affect gene expression levels, such as age, gender, and menopausal status, to ensure the authenticity of the research data. Although we adjusted for available covariates in the datasets, including age, gender, and menopausal status, some potential confounding factors, such as medication use, could not be accounted for due to their absence in the original studies. A common challenge in the field is the varying degree of standardization of phenotype information across public databases such as GEO. Second, although the OP outcome indicators in this study encompass populations from various ethnic groups, the blood eQTL and mQTL data used as exposure factors are predominantly derived from European populations. To enhance the robustness and generalizability of the study's findings, we chose to rely on data from European populations. However, this approach may introduce issues such as insufficient sample size and population stratification bias. Therefore, future research should consider including more data from diverse population groups to produce more reliable conclusions. Third, although peripheral blood samples are convenient and non-invasive, they may not fully capture tissue-specific gene expression and epigenetic changes that occur in bone cells, which are directly related to the pathophysiology of OP. Bone tissue, with its unique cellular composition and microenvironment, may contain specific eQTL and mQTL that are more closely re-

lated to the processes driving bone metabolism and remodeling. We reviewed the literature, but unfortunately, we were unable to find data related to bone tissue. Nevertheless, there are several reasons why peripheral blood data were chosen. First, obtaining bone tissue samples from OP patients presents challenges and is highly invasive, especially in large clinical cohorts where such tissue-specific data are more difficult to obtain. Second, peripheral blood can reflect systemic biological changes, including factors that may influence bone health, such as inflammation, endocrine signals, and metabolic alterations. Therefore, although peripheral blood may not provide direct insights into bone-specific molecular pathways, it still offers valuable information regarding the broader systemic factors contributing to the risk of OP. We acknowledge that the lack of bone tissue data represents a significant gap in this study. In future research, the use of bone biopsies or imaging techniques to analyze gene expression and DNAm in bone tissue holds the potential to provide more targeted efforts in directly understanding the molecular mechanisms of OP. Third, there is currently no universally accepted threshold standard for colocalization analysis. Different studies set thresholds based on their specific objectives to balance sensitivity and specificity. For instance, Chen *et al.* defined $PP.H4 > 0.5$ as “moderate evidence” for the preliminary identification of potential colocalization signals [62]. This threshold is particularly useful in multi-tissue or cross-omics analyses, as it helps reduce the risk of false negatives [63–66]. Similarly, Xu *et al.*, in a comparable multi-omics study, directly adopted $PP.H4 > 0.5$ as the criterion for colocalization evidence [67]. Most studies tend to use higher thresholds (e.g., 0.75 or 0.8) to increase specificity. However, such an approach may miss true signals, especially in the presence of heterogeneity or weak effect sizes. Our study integrates multi-omics data (mQTL, eQTL, pQTL, and GWAS), and thus requires a balance between discovery potential and false positive control during the exploratory phase. Therefore, we retained signals with moderate confidence to guide subsequent experimental validation. Finally, trans-regulatory elements, including transcription factors, enhancers, and noncoding RNA, play key roles in regulating gene expression and may influence susceptibility to OP by affecting key biological pathways. However, considering that cis-regulatory elements have more direct and specific biological effects than trans-regulatory elements [68], we used cis-eQTL/cis-mQTL data from human blood as the exposure factor. Future studies could explore the interaction of these genes with trans-regulatory elements by using transcriptomics data, chromatin immunoprecipitation sequencing, or CRISPR-based tech-

niques to identify enhancer regions, transcription factors, and noncoding RNA involved in their regulation. These studies will provide greater insight into the complex regulation of gene expression in OP and identify potential therapeutic targets.

In conclusion, this study explored the genetic association between IR and OP based on a multi-omics MR approach. The findings highlighted the significant role of IR-related genes, such as FAS and CHUK, in the pathogenesis of OP. Moreover, the newly discovered IR-related genes interacting with multiple hormones and inflammatory factors warrant further functional-level investigation to elucidate the underlying biological mechanisms. This research contributes to the understanding of IR in OP and identifies potentially novel therapeutic targets for clinical application.

Availability of data and material

The standard tools used in this study (including SMR and MeCS) are available at <https://yanglab.westlake.edu.cn/software/smr>. Four publicly available transcriptome datasets were obtained from the GEO database (<https://www.ncbi.nlm.nih.gov/geo/>). GWAS summary statistics for OP were downloaded from the UK/FinnGen open GWAS project (<https://gwas.mrcieu.ac.uk/> and <https://www.finnngen.fi/en>). Publicly available summary statistics of blood eQTLs and mQTLs were obtained from <http://www.eqtlgen.org> and <https://yanglab.westlake.edu.cn/software/smr/#mQTL-summarydata>, respectively. The data that support the findings of this study are available from the corresponding author upon reasonable request.

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Ethical approval

Not applicable.

Conflict of interest

The authors declare no conflict of interest.

References

- Lane JM, Russell L, Khan SN. Osteoporosis. *Clin Orthop Relat Res* 2000; 372: 139-50.
- Odén A, McCloskey EV, Kanis JA, Harvey NC, Johansson H. Burden of high fracture probability worldwide: secular increases 2010-2040. *Osteoporos Int* 2015; 26: 2243-8.
- Sigurdsson G, Halldorsson BV, Styrkarsdottir U, Kristjansson K, Stefansson K. Impact of genetics on low bone mass in adults. *J Bone Miner Res* 2008; 23: 1584-90.
- Teitelbaum SL. Bone resorption by osteoclasts. *Science* 2000; 289: 1504-8.
- Xue Y, Zhou Y, Li C, Zhang J, Liu F, Shi R. Causal relationship between Interleukin-27 expression levels and osteoporosis: a bidirectional mendelian randomization study. *BMC Musculoskelet Disord* 2024; 25: 680.
- Rifas L, Weitzmann MN. A novel T cell cytokine, secreted osteoclastogenic factor of activated T cells, induces osteoclast formation in a RANKL-independent manner. *Arthritis Rheum* 2009; 60: 3324-35.
- Amarasekara DS, Kim S, Rho J. Regulation of osteoblast differentiation by cytokine networks. *Int J Mol Sci* 2021; 22: 2851.
- Zhang R, Peng S, Zhu G. The role of secreted osteoclastogenic factor of activated T cells in bone remodeling. *Jpn Dent Sci Rev* 2022; 58: 227-32.
- Rong K, Lang Y, Zhou Y, et al. Risk genetic variants (IL-10) for osteoporosis in han population from Northwest China. *J Inflamm Res* 2023; 16: 1091-102.
- Ke K, Sul OJ, Chung SW, Suh JH, Choi HS. Lack of NOD2 attenuates ovariectomy-induced bone loss via inhibition of osteoclasts. *J Endocrinol* 2017; 235: 85-96.
- Xie J, Ma R, Xu X, et al. Identification of genetic association between mitochondrial dysfunction and knee osteoarthritis through integrating multi-omics: a summary data-based Mendelian randomization study. *Clin Rheumatol* 2024; 43: 3487-96.
- Xie N, Xie J, Wang Z, et al. The role of calcium, 25-hydroxyvitamin D, and parathyroid hormone in irritable bowel syndrome: a bidirectional two-sample mendelian randomization study. *Nutrients* 2022; 14: 5109.
- Wu Y, Zeng J, Zhang F, et al. Integrative analysis of omics summary data reveals putative mechanisms underlying complex traits. *Nat Commun* 2018; 9: 918.
- Zhu Z, Zhang F, Hu H, et al. Integration of summary data from GWAS and eQTL studies predicts complex trait gene targets. *Nat Genet* 2016; 48: 481-7.
- Lei SF, Wu S, Li LM, et al. An in vivo genome wide gene expression study of circulating monocytes suggested GBP1, STAT1 and CXCL10 as novel risk genes for the differentiation of peak bone mass. *Bone* 2009; 44: 1010-4.
- Zhou Y, Gao Y, Xu C, Shen H, Tian Q, Deng HW. A novel approach for correction of crosstalk effects in pathway analysis and its application in osteoporosis research. *Sci Rep* 2018; 8: 668.
- Benisch P, Schilling T, Klein-Hitpass L, et al. The transcriptional profile of mesenchymal stem cell populations in primary osteoporosis is distinct and shows overexpression of osteogenic inhibitors. *PLoS One* 2012; 7: e45142.
- Vösa U, Claringbould A, Westra HJ, et al. Large-scale cis- and trans-eQTL analyses identify thousands of genetic loci and polygenic scores that regulate blood gene expression. *Nat Genet* 2021; 53: 1300-10.
- Ollier W, Sprosen T, Peakman T. UK Biobank: from concept to reality. *Pharmacogenomics* 2005; 6: 639-46.
- Kurki MI, Karjalainen J, Palta P, et al. FinnGen provides genetic insights from a well-phenotyped isolated population. *Nature* 2023; 613: 508-18.

21. Xu S, Li X, Zhang S, et al. Oxidative stress gene expression, DNA methylation, and gut microbiota interaction trigger Crohn's disease: a multi-omics Mendelian randomization study. *BMC Med* 2023; 21: 179.
22. Walker MD, Shane E. Postmenopausal osteoporosis. *N Engl J Med* 2023; 389: 1979-91.
23. Yamada A, Arakaki R, Saito M, Kudo Y, Ishimaru N. Dual role of Fas/FasL-mediated signal in peripheral immune tolerance. *Front Immunol* 2017; 8: 403.
24. Galli G, Vacher P, Ryffel B, Blanco P, Legembre P. Fas/CD95 signaling pathway in damage-associated molecular pattern (DAMP)-sensing receptors. *Cells* 2022; 11: 1438.
25. Yan H, Hong Y, Cai Y. Association between FAS gene -670 A/G and -1377 G/A polymorphisms and the risk of autoimmune diseases: a meta-analysis. *Biosci Rep* 2020; 40: BSR20191197.
26. Huang D, Xiao J, Deng X, et al. Association between Fas/FasL gene polymorphism and musculoskeletal degenerative diseases: a meta-analysis. *BMC Musculoskelet Disord* 2018; 19: 137.
27. Glavan BJ, Holden TD, Goss CH, et al. Genetic variation in the FAS gene and associations with acute lung injury. *Am J Respir Crit Care Med* 2011; 183: 356-63.
28. Zhang L, Cheng J, Su H, Wang Z, Dai W. Diagnostic value of circulating bone turnover markers osteocalcin, cathepsin K, and osteoprotegerin for osteoporosis in middle-aged and elderly postmenopausal women. *Arch Med Sci* 2024; 20: 1727-30.
29. Wu X, Pan G, McKenna MA, Zayzafoon M, Xiong WC, McDonald JM. RANKL regulates Fas expression and Fas-mediated apoptosis in osteoclasts. *J Bone Miner Res* 2005; 20: 107-16.
30. Udagawa N, Koide M, Nakamura M, et al. Osteoclast differentiation by RANKL and OPG signaling pathways. *J Bone Miner Metab* 2021; 39: 19-26.
31. Zhao G, Lyu J, Huang X, et al. The role and underlying mechanism of dental pulp stem cell-derived exosomal miR-31 in the treatment of osteoarthritis by targeting mTOR to enhance chondrocyte autophagy levels. *Arch Med Sci* 2024; 20: 1680-94.
32. Morikawa N, Kato Y, Takeshita N, Shimizu Y. Pharmacological characterization of AS2690168, a novel small molecule RANKL signal transduction inhibitor. *Eur J Pharmacol* 2022; 924: 174941.
33. Xiong J, Piemontese M, Thostenson JD, Weinstein RS, Manolagas SC, O'Brien CA. Osteocyte-derived RANKL is a critical mediator of the increased bone resorption caused by dietary calcium deficiency. *Bone* 2014; 66: 146-54.
34. Chen T, Wang H, Jiang C, Lu Y. PKD1 alleviates oxidative stress-inhibited osteogenesis of rat bone marrow-derived mesenchymal stem cells through TAZ activation. *J Cell Biochem* 2021; 122: 1715-25.
35. Astudillo P, Ríos S, Pastenes L, Pino AM, Rodríguez JP. Increased adipogenesis of osteoporotic human-mesenchymal stem cells (MSCs) characterizes by impaired leptin action. *J Cell Biochem* 2008; 103: 1054-65.
36. Chen K, Qiu P, Yuan Y, et al. Pseurotin A inhibits osteoclastogenesis and prevents ovariectomized-induced bone loss by suppressing reactive oxygen species. *Theranostics* 2019; 9: 1634-50.
37. Agidigbi TS, Kim C. Reactive oxygen species in osteoclast differentiation and possible pharmaceutical targets of ROS-mediated osteoclast diseases. *Int J Mol Sci* 2019; 20: 3576.
38. Liu HD, Ren MX, Li Y, et al. Melatonin alleviates hydrogen peroxide induced oxidative damage in MC3T3-E1 cells and promotes osteogenesis by activating SIRT1. *Free Radic Res* 2022; 56: 63-76.
39. Gürler EB, Çilingir-Kaya ÖT, Peker Eyüboğlu I, et al. Melatonin supports alendronate in preserving bone matrix and prevents gastric inflammation in ovariectomized rats. *Cell Biochem Funct* 2019; 37: 102-12.
40. Rippo MR, Babini L, Prattichizzo F, et al. Low FasL levels promote proliferation of human bone marrow-derived mesenchymal stem cells, higher levels inhibit their differentiation into adipocytes. *Cell Death Dis* 2013; 4: e594.
41. Wang L, Liu S, Zhao Y, et al. Osteoblast-induced osteoclast apoptosis by fas ligand/FAS pathway is required for maintenance of bone mass. *Cell Death Differ* 2015; 22: 1654-64.
42. Harding O, Holzer E, Riley JF, Martens S, Holzbaur ELF. Damaged mitochondria recruit the effector NEMO to activate NF- κ B signaling. *Mol Cell* 2023; 83: 3188-204.e7.
43. Li X, Hu Y. Attribution of NF- κ B activity to CHUK/IKK α -involved carcinogenesis. *Cancers* 2021; 13: 1411.
44. Takeda K, Takeuchi O, Tsujimura T, et al. Limb and skin abnormalities in mice lacking IKK α . *Science* 1999; 284: 313-6.
45. Antonia RJ, Hagan RS, Baldwin AS. Expanding the view of IKK: new substrates and new biology. *Trends Cell Biol* 2021; 31: 166-78.
46. Walkon LL, Strubbe-Rivera JO, Bazil JN. Calcium overload and mitochondrial metabolism. *Biomolecules* 2022; 12: 1891.
47. Suh J, Kim NK, Shim W, et al. Mitochondrial fragmentation and donut formation enhance mitochondrial secretion to promote osteogenesis. *Cell Metab* 2023; 35: 345-60.e7.
48. Zhai Q, Chen X, Fei D, et al. Nanorepairers rescue inflammation-induced mitochondrial dysfunction in mesenchymal stem cells. *Adv Sci* 2022; 9: e2103839.
49. Eijken M, Meijer IMJ, Westbroek I, et al. Wnt signaling acts and is regulated in a human osteoblast differentiation dependent manner. *J Cell Biochem* 2008; 104: 568-79.
50. Liu F, Yuan Y, Bai L, et al. LRRc17 controls BMSC senescence via mitophagy and inhibits the therapeutic effect of BMSCs on ovariectomy-induced bone loss. *Redox Biol* 2021; 43: 101963.
51. Ma W, Li C. Enhancing postmenopausal osteoporosis: a study of KLF2 transcription factor secretion and PI3K-Akt signaling pathway activation by PIK3CA in bone marrow mesenchymal stem cells. *Arch Med Sci* 2024; 20: 918-37.
52. Liang Y, Li W, Li X, Nan J. Involvement of PI3K/Akt/ β -catenin signaling in schisandrin B-mitigated bone deterioration in an experimental rat model of estrogen deficiency. *Arch Med Sci* 2023; 19: 1520-9.
53. Yang X, Jiang T, Wang Y, Guo L. The role and mechanism of SIRT1 in resveratrol-regulated osteoblast autophagy in osteoporosis rats. *Sci Rep* 2019; 9: 18424.
54. Zheng Z, Zhang X, Huang B, et al. Site-1 protease controls osteoclastogenesis by mediating LC3 transcription. *Cell Death Differ* 2021; 28: 2001-18.
55. Shao B, Fu X, Yu Y, Yang D. Regulatory effects of miR-NA-181a on FasL expression in bone marrow mesenchymal stem cells and its effect on CD4+T lymphocyte apoptosis in estrogen deficiency-induced osteoporosis. *Mol Med Rep* 2018; 18: 920-30.
56. Tella SH, Gallagher JC. Prevention and treatment of postmenopausal osteoporosis. *J Steroid Biochem Mol Biol* 2014; 142: 155-70.

57. Yang S, Zhu L, Luo F, Jiang R, Yu L. Association between female reproductive characteristics and bone mineral density: evidence from Mendelian randomization. *Arch Med Sci* 2024; 20: 2063-6.
58. Boyce BF, Aufdemorte TB, Garrett IR, Yates AJ, Mundy GR. Effects of interleukin-1 on bone turnover in normal mice. *Endocrinology* 1989; 125: 1142-50.
59. Landin-Wilhelmsen K, Nilsson A, Bosaeus I, Bengtsson BA. Growth hormone increases bone mineral content in postmenopausal osteoporosis: a randomized placebo-controlled trial. *J Bone Miner Res* 2003; 18: 393-405.
60. Kukita T, Nomiya H, Ohmoto Y, et al. Macrophage inflammatory protein-1 alpha (LD78) expressed in human bone marrow: its role in regulation of hematopoiesis and osteoclast recruitment. *Lab Invest J Tech Methods Pathol* 1997; 76: 399-406.
61. Kawao N, Kaji H. Interactions between muscle tissues and bone metabolism. *J Cell Biochem* 2015; 116: 687-95.
62. Chen J, Xu F, Ruan X, et al. Therapeutic targets for inflammatory bowel disease: proteome-wide Mendelian randomization and colocalization analyses. *EBioMedicine* 2023; 89: 104494.
63. Gudjonsson A, Gudmundsdottir V, Axelsson GT, et al. A genome-wide association study of serum proteins reveals shared loci with common diseases. *Nat Commun* 2022; 13: 480.
64. Shi J, Wang Z, Yi M, et al. Evidence based on Mendelian randomization and colocalization analysis strengthens causal relationships between structural changes in specific brain regions and risk of amyotrophic lateral sclerosis. *Front Neurosci* 2024; 18: 1333782.
65. Rasooly D, Peloso GM, Pereira AC, et al. Genome-wide association analysis and Mendelian randomization proteomics identify drug targets for heart failure. *Nat Commun* 2023; 14: 3826.
66. Arvanitis M, Tayeb K, Strober BJ, Battle A. Redefining tissue specificity of genetic regulation of gene expression in the presence of allelic heterogeneity. *Am J Hum Genet* 2022; 109: 223-39.
67. Xu S, Li X, Zhang S, et al. Oxidative stress gene expression, DNA methylation, and gut microbiota interaction trigger Crohn's disease: a multi-omics Mendelian randomization study. *BMC Med* 2023; 21: 179.
68. Emilsson V, Ilkov M, Lamb JR, et al. Co-regulatory networks of human serum proteins link genetics to disease. *Science* 2018; 361: 769-73.