Development and Validation of a Nomogram Prediction Model for Delayed Cerebral Edema After Intracerebral Hemorrhage Based on Serum Inflammatory Markers and Hemodynamics

Keywords

Hemodynamics, Delayed cerebral edema, Serum inflammatory factors, hypertensive cerebral hemorrhage

Abstract

Introduction

To explore the risk prediction model of delayed cerebral edema after hypertensive cerebral hemorrhage based on serum inflammatory factors and hemodynamics and its clinical application value.

Material and methods

Multivariate Logistic regression was used to analyze the risk factors of postoperative bleeding and build a nomogram prediction model, and ROC curve and calibration curve were drawn to evaluate the prediction efficiency of nomogram model, which was verified in the verification set.

Results

The results of univariate analysis showed that there were significant differences in L-6, TNF- α , IL-1 β , MIP-1 α , CBF, CVR, HMGB1 and CRP between patients with edema and those without edema in training (P<0.05). Logistic regression analysis showed that L-6, TNF- α , IL-1 β , CBF, CVR, HMGB1 and CRP were independent risk factors for bleeding (P<0.05). The nomogram model has good calibration and fitting degree between prediction and reality in training set and verification set (C-index index is 0.792 and 0.799). The results of HosmerLemeshow test are χ 2=16.582, P =0.035 and χ 2=7.472, P = 0.487. The ROC curve is shown in the training set and the validation set. The AUC of the nomogram model in predicting delayed cerebral edema is 0. 794 and 0. 796.

Conclusions

The nomogram prediction model of delayed cerebral edema after hypertensive cerebral hemorrhage based on serum inflammatory factors (IL-6, TNF- α , IL-1 β), hemodynamic parameters (CBF, CVR) and other related factors (HMGB1, CRP) was successfully established. The model can effectively predict the risk of delayed cerebral edema in patients with hypertensive cerebral hemorrhage.

1	Development and Validation of a Nomogram Prediction Model
2	for Delayed Cerebral Edema After Intracerebral Hemorrhage
3	Based on Serum Inflammatory Markers and Hemodynamics
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23 Abstract

Objective: To explore the risk prediction model of delayed cerebral edema after
hypertensive cerebral hemorrhage based on serum inflammatory factors and
hemodynamics and its clinical application value.

Methods: Multivariate Logistic regression was used to analyze the risk factors of postoperative bleeding and build a nomogram prediction model, and ROC curve and calibration curve were drawn to evaluate the prediction efficiency of nomogram model, which was verified in the verification set.

Results: The results of univariate analysis showed that there were significant 31 differences in L-6, TNF-a, IL-1β, MIP-1a, CBF, CVR, HMGB1 and CRP between 32 patients with edema and those without edema in training (P<0.05). Logistic regression 33 analysis showed that L-6, TNF-a, IL-1β, CBF, CVR, HMGB1 and CRP were 34 independent risk factors for bleeding (P<0.05). The nomogram model has good 35 calibration and fitting degree between prediction and reality in training set and 36 37 verification set (C-index index is 0.792 and 0.799). The results of HosmerLemeshow test are χ^2 =16.582, P =0.035 and χ^2 =7.472, P = 0.487. The ROC curve is shown in the 38 training set and the validation set. The AUC of the nomogram model in predicting 39 delayed cerebral edema is 0. 794 and 0. 796. 40

41 Conclusion: The nomogram prediction model of delayed cerebral edema after
42 hypertensive cerebral hemorrhage based on serum inflammatory factors (IL-6, TNF-α,
43 IL-1β), hemodynamic parameters (CBF, CVR) and other related factors (HMGB1,
44 CRP) was successfully established. The model can effectively predict the risk of

45 delayed cerebral edema in patients with hypertensive cerebral hemorrhage.

Keywords: Serum inflammatory factors; Hemodynamics; Delayed cerebral edema;
hypertensive cerebral hemorrhage

48

49 **1 Introduction**

With the acceleration of social aging, hypertensive cerebral hemorrhage, as a 50 common and dangerous cerebrovascular disease, has an increasing trend year by year, 51 posing a serious threat to human health on a global scale [1]. This disease has a 52 53 sudden onset and rapid progress, which often causes irreversible damage to the patient's brain tissue in a short time [2]. Hypertensive cerebral hemorrhage will not 54 only cause physical damage to local brain tissue, but also trigger a series of complex 55 56 pathophysiological changes [3]. Among them, delayed cerebral edema, as one of the common and extremely harmful complications after operation, seriously affects the 57 prognosis of patients. It usually develops gradually after cerebral hemorrhage for a 58 59 period of time, which makes the intracranial pressure continue to rise, then oppresses the surrounding brain tissue, affects cerebral perfusion, and may even lead to the 60 formation of cerebral hernia, which is an important cause of disability and death of 61 patients [4]. 62

At present, in clinical practice, the prediction method of delayed cerebral edema after hypertensive cerebral hemorrhage is not perfect. Doctors mostly rely on their own clinical experience and some conventional imaging examination methods to make judgments, but these methods have obvious shortcomings. Although clinical experience is valuable, it is subjective, and there may be differences among different
doctors. However, traditional imaging examination can only be found when brain
edema has occurred or developed to a certain extent, which has limited value for early
prediction, which makes us often in a passive situation in the face of delayed cerebral
edema and unable to take timely and effective preventive measures [5].

In recent years, more and more studies show that serum inflammatory factors 72 and hemodynamic changes play a key role in the pathophysiological process after 73 cerebral hemorrhage. As an important marker of inflammatory reaction, serum 74 75 inflammatory factors have changed significantly during the initiation and development of local inflammatory reaction after cerebral hemorrhage [6-7]. They can 76 affect the function of cerebral vascular endothelial cells, the permeability of 77 78 blood-brain barrier and the survival state of neurons in many ways. The abnormal expression of inflammatory factors may induce the chemotaxis and activation of 79 white blood cells, release more inflammatory mediators, and further aggravate the 80 damage and edema of brain tissue [8-9]. At the same time, the change of 81 hemodynamics is also a factor that can not be ignored. The change of hemodynamic 82 parameters such as cerebral blood flow and cerebrovascular resistance is directly 83 related to the perfusion of brain tissue [10]. After the occurrence of cerebral 84 hemorrhage, the automatic regulation mechanism of cerebral blood vessels may be 85 damaged, leading to the decrease of cerebral blood flow and the increase of cerebral 86 vascular resistance. This abnormal blood flow state will lead to ischemia and hypoxia 87 of brain tissue, which will further promote a series of pathophysiological chain 88

reactions and create conditions for the occurrence of delayed cerebral edema [11-13].

90 Therefore, it is of great significance to conduct in - depth research on the 91 relationship between the changes in serum inflammatory factors, hemodynamics, and 92 delayed cerebral edema. The aim of this study is to construct an accurate and effective 93 prediction model, provide a strong basis for clinical treatment and intervention, and

94 improve the prognosis of patients with hypertensive intracerebral hemorrhage.

95 **2 Data and methods**

96 2.1 Research object

From January 2022 to January 2024, 300 patients with hypertensive intracerebral hemorrhage in our hospital were selected as the research subjects, and all of them informed and agreed to participate in this study voluntarily, which was approved by the ethics Committee of our hospital. Patients were randomly divided into training set (n=210) and verification set (n=90) according to the ratio of 7:3, and general data were collected simultaneously. Data can be provided upon request. The follow-up period was 10 months.

104 **2.2 Inclusion exclusion criteria**

105 2.2.1 Inclusion criteria

Patients should clearly meet the diagnostic criteria of hypertensive cerebral hemorrhage; The age is between 40 and 80 years old; The time from onset to admission is within 24 hours; The patient or family members sign the informed consent form. Ethical Approval Date: December 20, 2021, Ethical Approval Number: KY2023126, Research Protocol (Version Number: Y1.0; Version Date: December 6,

111 2023), Informed Consent Form (Version Number: V1.0; Date: December 6, 2023).

112 **2.2.2 Exclusion criteria**

Exclude patients with other serious craniocerebral trauma, brain tumor and other diseases; Patients with severe heart, liver and renal insufficiency; The clinical data is incomplete.

116 2.3 Detection methods

117 2.3.1 Detection method of serum inflammatory factors (enzyme-linked 118 immunosorbent assay, ELISA)

Collect patient's venous blood in a vacuum blood collection tube without 119 anticoagulant, and centrifuge at the speed of 3000-3500rpm for 15 minutes after the 120 blood naturally coagulates to obtain serum. Transfer the serum to a clean EP tube, and 121 122 store the sample at -20°C or lower if it is not detected immediately. Before testing, thaw the sample at room temperature and mix it gently. Select the corresponding 123 ELISA kit according to the measured serum inflammatory factors, and check the 124 status of each reagent in the kit. According to the instructions, the standard is prepared 125 into a suitable concentration gradient with diluent. Add 50µL of standard solution and 126 50µL of serum sample to the microplate coated with specific antibody in turn, and set 127 up multiple wells for each sample and standard. Gently shake the microplate to make 128 the liquid fully mixed, then seal the microplate with a sealing film and incubate at 129 room temperature for 1.5 hours. After the incubation, the liquid in the well was 130 discarded, and the microplate was washed with washing liquid for 5 times, soaking 131 for 2 minutes each time to remove unbound substances. Then, enzyme-labeled 132

secondary antibody (100µL per well) was added, and the plate was sealed again and 133 incubated at room temperature for 60 minutes. After that, repeat the washing step, and 134 then add the substrate solution, 100µL per hole, and develop color in the dark. When 135 the color is developed to an appropriate degree, the reaction is terminated by adding a 136 termination solution. Finally, the absorbance of each hole was measured by 137 enzyme-labeled instrument at the wavelength of 450nm, Each sample is measured 138 three times. Based on the average value of the absorbance obtained from the three 139 measurements, the concentration of inflammatory factors in the sample is calculated 140 141 in combination with the standard curve.

142 **2.3.2 Detection method of hemodynamic parameters (TCD)**

Patients take supine position or sitting position, and keep quiet and relaxed. 143 144 Examiners apply a proper amount of coupling agent to the transcranial Doppler ultrasound probe to ensure good contact between the probe and the skin and reduce air 145 interference. Firstly, the temporal window is found in the temporal part, which is a 146 common part of transcranial Doppler ultrasound examination. Generally, the 147 appropriate ultrasonic signal is found by adjusting the angle and depth of the probe. 148 When clear blood flow signals are obtained, the blood flow spectra of main cerebral 149 vessels such as middle cerebral artery (MCA), anterior cerebral artery (ACA) and 150 posterior cerebral artery (PCA) are identified. For the middle cerebral artery, the 151 probe is usually placed in the temporal window, and the depth is adjusted to about 152 30-65mm to find signals with typical blood flow spectrum characteristics, that is, the 153 systolic blood flow velocity is relatively fast, the diastolic blood flow velocity is 154

relatively low, and the blood flow spectrum is three peaks (S1, S2 and D peaks). The 155 blood flow velocity parameters of each major cerebral blood vessel need to be 156 measured repeatedly 5 times at different time points. The blood flow velocity 157 parameters of major cerebral vessels, including peak systolic blood flow velocity (Vs), 158 end diastolic blood flow velocity (Vd) and average blood flow velocity (Vm), can be 159 directly read on the instrument or calculated by measuring spectral envelope line. At 160 the same time, the blood pressure was recorded, and combined with the measured 161 blood flow velocity, the hemodynamic parameters such as cerebrovascular resistance 162 163 (CVR) and cerebral blood flow (CBF) were calculated according to the formula (such as CVR = (mean arterial pressure-intracranial pressure)/CBF, in which CBF can be 164 calculated from blood flow velocity and other parameters through a specific formula). 165 166 During the inspection, it is necessary to adjust the angle and depth of the probe to obtain the best signal quality, and measure the average value for many times at 167 different time points to improve the accuracy of the measurement. If the detection of 168 temporal window is not ideal, we can try to check it through other parts such as 169 orbital window or occipital window. 170

2.3.3 Challenges Encountered During the Measurement Process and Their
Solutions During the measurement process, some patients are unable to remain
quiet and relaxed. This can lead to head movement or vasospasm, causing the position
of the probe during measurement to change. As a result, the measured blood flow
velocity values fluctuate significantly, failing to accurately reflect the true
hemodynamic state and affecting the accuracy of subsequent parameter calculations.

Before the examination, fully explain the examination process and precautions to the patients to relieve their nervousness. For patients who have difficulty cooperating, appropriate psychological comfort can be provided or sedative medications can be used (subject to the doctor's assessment and permission). At the same time, during the measurement, use a fixation device to assist in fixing the patient's head to reduce errors caused by head movement.

183 If the TCD instrument is not calibrated regularly, there may be errors in the basic 184 data it measures, directly affecting the measured values of blood flow velocity. 185 Consequently, the hemodynamic parameters calculated based on these data lose their 186 reliability. Moreover, if unstable conditions such as signal drift occur during the use of 187 the instrument, it will also render the measurement results unreliable.

We have established a strict instrument calibration system. In accordance with the requirements of the instrument operation manual, the TCD instrument is calibrated and maintained regularly. Before each use, warm up and self - check the instrument to ensure it is in a stable working condition. If abnormal conditions such as signal drift are detected during the measurement, stop the measurement immediately, check the instrument connections and settings, and if necessary, recalibrate the instrument before resuming the measurement.

195 **2.4 Statistical methods**

All data were imported into SPSS 26.0 software for analysis. Categorical data
were described by frequency and percentage, and the chi-square test was used.
Measurement data with a normal distribution were expressed as mean±standard

deviation, and the independent-samples t-test was applied. A model was established 199 through Logistic regression analysis, and the rms package in R language (R4.0.0) was 200 used to visualize the model. The Receiver Operating Characteristic (ROC) curve of 201 the model was plotted to determine the sensitivity, specificity, Youden index, and 202 optimal cut-off value of the model. The discrimination of the model was evaluated by 203 the Area Under the Curve (AUC) of the ROC curve. The larger the value, the better 204 the discrimination. The calibration of the model was evaluated using the calibration 205 curve or Hosmer-Lemeshow test. The Decision Curve Analysis (DCA) was plotted to 206 207 test the practical application efficiency of the model. **3 Results** 208 3.1 Comparison of delayed cerebral edema rate and clinical characteristics 209 210 between training set and verification set. 100 cases (47.62%) of 175 patients in the training set developed delayed cerebral 211 edema, and 42 cases (46.67%) of 90 patients in the verification set developed delayed 212 cerebral edema. There was no significant difference in the incidence and clinical 213 characteristics between the training set and the verification set (P > 0.05), as shown in 214 Table 1. 215 3.2 Comparison of clinical features between edema group and non-edema group 216 217 In the training concentration, the results of univariate analysis showed that there were significant differences in L-6, TNF-a, IL-1β, MIP-1a, CBF, CVR, HMGB1 and 218 CRP between the patients with edema and those without edema (P < 0.05), as shown 219 in Table 2. 220

3.3 Risk factors analysis of delayed cerebral edema after cerebral hemorrhage

The occurrence of edema was taken as the dependent variable (0= none, 1= yes), and the factors in univariate analysis (P < 0.05) were taken as the covariate. Further multivariate Logistic regression analysis showed that L-6, TNF- α , IL-1 β , CBF, CVR, HMGB1 and CRP were independent risk factors for bleeding (P < 0.05), as shown in Table 3.

3.4 Establishment of nomogram prediction model for delayed cerebral edema after cerebral hemorrhage

Based on the independent risk factors identified by multivariate Logistic regression analysis, the nomogram prediction model of delayed cerebral edema after cerebral hemorrhage was constructed, and the independent risk factors in the model were assigned, and the total score for predicting delayed cerebral edema after cerebral hemorrhage was calculated, which was reflected by the prediction probability of bleeding. The higher the total score, the higher the accuracy of predicting delayed cerebral edema after cerebral hemorrhage. See figure 1.

3.5 Evaluation and validation of the prediction model of delayed cerebral edema

237 after cerebral hemorrhage

In the training set and verification set, the C-index index of nomogram model is 0.792 and 0.799, respectively. The average absolute error of the coincidence degree between the predicted value and the real value in the calibration curve is 0.188 and 0.182, respectively, and the results of HosmerLemeshow test are χ^2 =16.582, P =0.035 and χ^2 =7.472, P = 0.487, respectively. The ROC curve is shown in the training set

243	and the validation set. The AUC of the nomogram model in predicting delayed
244	cerebral edema is 0. 794(95% CI: 0. 722-0. 865) and 0. 796(95% CI: 0. 688-0.904),
245	and the sensitivity and specificity are 0.629 and 0. 804, respectively. See Figure 2 for
246	the calibration curve and Figure 3 for the ROC curve.
247	26 Analysis of desision survey of non-servery prediction model for deleved

3.6 Analysis of decision curve of nomogram prediction model for delayed
 cerebral edema after cerebral hemorrhage

The decision curve shows that when the threshold probability is between 0. 05 and 0. 95, the nomogram model constructed in this study has more clinical benefits in predicting postoperative bleeding. See Figure 4.

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253 **4 Discussion**

254 Delayed cerebral edema after hypertensive cerebral hemorrhage seriously affects the prognosis of patients, and there is no effective prediction method in clinic at 255 present. The nomogram prediction model constructed in this study is of great 256 significance. Serum inflammatory factors play a key role in the inflammatory 257 response after cerebral hemorrhage. As important proinflammatory factors, IL-6, 258 TNF- α and IL-1 β can activate a series of inflammatory cascade reactions [14-15]. IL-6 259 can induce the activation and proliferation of immune cells, promote the production of 260 adhesion molecules by vascular endothelial cells, increase vascular permeability, and 261 make plasma components seep into the brain tissue space, leading to brain edema [16]. 262 TNF- α can directly damage vascular endothelial cells, destroy the integrity of 263 blood-brain barrier and induce chemotaxis and infiltration of inflammatory cells [17]. 264

IL-1 β activates endothelial cells in the early stage of inflammation, promotes the 265 expression of inflammatory cell adhesion molecules, and further intensifies the 266 inflammatory reaction. These inflammatory factors interact and jointly promote the 267 development of brain edema [18]. The changes of hemodynamic parameters CBF and 268 CVR are also closely related to delayed cerebral edema [19]. The decrease of CBF 269 means insufficient perfusion of brain tissue, which will cause ischemia and hypoxia of 270 brain tissue, energy metabolism disorder of cells, increase anaerobic metabolism, 271 produce a large amount of lactic acid accumulation, lead to intracellular acidosis, and 272 then cause imbalance of ion balance inside and outside cells and edema of cells [20]. 273 At the same time, ischemia and hypoxia can also activate inflammation-related signal 274 pathways, promote inflammatory cells to release more inflammatory mediators, and 275 276 aggravate brain edema. The increase of CVR reflects the spasm or stenosis of cerebral vessels, which will hinder the normal blood perfusion, further reduce CBF, and 277 interact with CBF, becoming an important hemodynamic factor in the occurrence of 278 cerebral edema [21]. As an injury-related molecular model molecule, HMGB1 is 279 released from injured cells after cerebral hemorrhage. It can activate the signal 280 pathways of various immune cells, induce inflammatory cells to produce more 281 inflammatory factors, such as IL-6 and TNF-a, and form positive feedback, 282 continuously amplify the inflammatory response, increase the permeability of 283 blood-brain barrier and promote the formation of brain edema [22-23]. As an acute 284 phase reaction protein, the increase of CRP level reflects the activation degree of 285 inflammatory reaction, which can activate the complement system, enhance the 286

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function of phagocytes, lead to vascular endothelial damage, promote the destruction of blood-brain barrier and create conditions for brain edema[24].

289 The nomogram model constructed in this study is excellent in forecasting efficiency. From the point of calibration and fitting degree, the C- index index is 0.792 290 and 0.799, respectively, which shows that the model has good prediction accuracy and 291 discrimination ability. The average absolute error of the calibration curve is small, 292 which is 0.188 and 0.182 respectively, indicating that the predicted value is in high 293 agreement with the real value. Hosmer-Lemeshow test results further support the 294 goodness of fit of the model in the training set (P = 0.035) and the verification set (P =295 0.487). ROC curve analysis shows that the AUC of the training set and the validation 296 set are 0.794 and 0.796, respectively. This result shows that the model has good 297 discrimination and can effectively distinguish patients with and without delayed 298 cerebral edema. The sensitivity and specificity of the model are 0.629, 0.805, 0.633 299 and 0.786, respectively, which shows that the model is reliable in accurately 300 identifying whether patients will have delayed cerebral edema. Compared with 301 traditional forecasting methods, this model has obvious advantages. Traditional 302 methods rely on doctors' experience or single examination index, which is subjective 303 and limited. However, this model comprehensively considers many factors closely 304 related to the occurrence of cerebral edema, organically combines serum 305 inflammatory factors and hemodynamic parameters, and more comprehensively 306 reflects the complex pathophysiological state after cerebral hemorrhage. The form of 307 nomogram is intuitive and easy to understand. Clinicians can easily calculate scores 308

according to patients' specific indicators, quickly assess the risk of patients with
delayed cerebral edema, and help to identify high-risk patients early in clinical
practice.

Although some achievements have been made in this study, there are still some 312 limitations. First of all, although the sample size has reached 300 cases, it may be 313 necessary to further expand the sample size for this complex clinical situation to 314 improve the stability and accuracy of the model. A larger sample size can better cover 315 patients with different severity and individual differences, and reduce the errors 316 317 caused by sample deviation. Secondly, there may be other factors that have not been taken into account in the process of model construction, which have an impact on 318 delayed cerebral edema. For example, the genetic polymorphism of patients may play 319 320 a role in the inflammatory reaction and the development of brain edema after cerebral hemorrhage, but this study does not involve this aspect. In addition, factors such as 321 patients' lifestyle and drug use may also be related to the occurrence of brain edema. 322 323 Future research can further improve the model and include these potential factors. In terms of research design, this study is a retrospective study. Although the relationship 324 between related factors and delayed cerebral edema can be analyzed to some extent, 325 there may be problems such as information bias. Prospective research can be carried 326 out in the future to further verify the application value of the model in clinical practice, 327 and at the same time, the whole process from cerebral hemorrhage to delayed cerebral 328 edema can be observed more accurately, and more comprehensive data can be 329 collected. In this study, the specific application processes, application scenarios of the 330

model in clinical practice, as well as the adaptation methods to the existing clinical workflow have not been clarified yet. One of the key focuses of future research work lies in developing the nomogram into software or clinical tools. This can provide intuitive and efficient support for clinical decision - making, enhance the accessibility and practicality of the model in clinical practice, promote its true integration into the clinical decision - making process, and improve the quality of medical services.

In a word, the nomogram prediction model based on serum inflammatory factors and hemodynamics provides a new and effective tool for predicting delayed cerebral edema after hypertensive cerebral hemorrhage. Through further research and improvement, it is expected to continuously optimize the model, better serve the clinic and improve the prognosis of patients with hypertensive cerebral hemorrhage.

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347 Authors' contributions

Conception and design: Fuqiang Zhai and Jixin Shou. Method: Haidong Gao and Jianye Wang. Data Collection: Haidong Gao and Jianye Wang. Manuscript Writing: Fuqiang Zhai. Manuscript revision: Jixin Shou and Peng Yang. Research supervision: Jixin Shou and Peng Yang. All authors contributed to the article and approved the submitted version.

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357	
358	Data Availability
359	The simulation experiment data used to support the findings of this study are available
360	from the corresponding author upon request.
361	
362	Ethics approval and consent to participate
363	The study was approved by the Ethics Committee of the Fifth Affiliated Hospital of
364	Zhengzhou University (No. KY2023126), and informed consent was obtained from
365	all patients. This study was conducted in accordance with the Declaration of Helsinki.
366	
367	Consent for publication
368	Not applicable.
369	
370	Competing interests
371	The authors declare that they have no competing interests.
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375 **References**

- 376 [1] Chen F, Zhang S, Li B, et al. A review of invasive intracranial pressure monitoring
- following surgery for hypertensive cerebral hemorrhage[J]. Front Neurol.
 2023 ,14:1108722.
- [2] Xiao W, Jiang Z, Wan W, et al. miR-145-5p targets MMP2 to protect brain
 injury in hypertensive intracerebral hemorrhage via inactivation of the Wnt/β-catenin
 signaling pathway[J]. Ann Transl Med. 2022 ,10(10):571.
- 382 [3] Wu J, Zhang S. Analysis of the Therapeutic Effect and Prognostic Factors of 126
- 383 Patients With Hypertensive Cerebral Hemorrhage Treated by Soft-Channel Minim
- ally Invasive Puncture and Drainage[J]. Front Surg. 2022, 9:885580.
- [4] Chen Y, Qin C, Chang J, et al. Defining Delayed Perihematomal Edema
 Expansion in Intracerebral Hemorrhage: Segmentation, Time Course, Risk Factors
- and Clinical Outcome[J]. Front Immunol. 2022, 13:911207.
- 388 [5] Otomo M, Kanamori M, Sato S, et al. A Case of Haemorrhagic-Onset
- Glioblastoma With Delayed Diagnosis[J]. Cureus. 2023, 15(2):e34672.
- 390 [6] Dicpinigaitis AJ, Galea VP, Sursal T, et al. Low serum albumin as a risk factor
- 391 for delayed cerebral ischemia following aneurysmal subarachnoid hemorrhage: eICU
- collaborative research data base analysis[J]. J Neurosurg Sci. 2024,68(3):287-293.
- 393 [7] Liu Y, Qiu T, Fu Z, et al. Systemic immune-inflammation index and serum
- 394 glucose-potassium ratio predict poor prognosis in patients with spontaneous cerebral
- hemorrhage: A n observational study[J]. Medicine (Baltimore). 2024,103(29):e39041.
- 396 [8] Henein MY, Vancheri S, Longo G, et al. The Role of Inflammation in

- 397 Cardiovascular Disease[J]. Int J Mol Sci. 2022,23(21):12906.
- 398 [9] Han C, Zhai L, Shen H, et al. Advanced Glycation End-Products (AGEs)
- 399 Promote Endothelial Cell Pyroptosis Under Cerebral Ischemia and Hypoxia via
- 400 HIF-1α-RAGE-NLRP3[J]. Mol Neurobiol. 2023 ,60(5):2355-2366.
- 401 [10] Li X, Hui Y, Shi H, et al. Association of blood pressure with brain perfusion and
- 402 structure: A population-based prospective study[J]. Eur J Radiol. 2023,165:110889.
- 403 [11] Webb AJS, Werring DJ. New Insights Into Cerebrovascular Pathophysiology and
- 404 Hypertension[J]. Stroke. 2022 ,53(4):1054-1064.
- [12] Wan Y, Holste KG, Hua Y, et al. Brain edema formation and therapy after
 intracerebral hemorrhage[J]. Neurobiol Dis. 2023 ,176:105948.
- 407 [13] Lauzier DC, Jayaraman K, Yuan JY, et al. Early Brain Injury After Subarachnoid
- 408 Hemorrhage: Incidence and Mechanisms[J]. Stroke. 2023,54(5):1426-1440.
- 409 [14] Hong G. Enoxolone suppresses apoptosis in chondrocytes and progression of
- 410 osteoarthritis via modulating the ERK1/2 signaling pathway[J]. Archives of Medical
- 411 Science. 2024 ,20(3):947-961.
- 412 [15] Rafaqat S, Patoulias D, Behnoush AH, et al. Interleukins: pathophysiological role
- in acute pancreatitis[J]. Archives of Medical Science. 2024,20(1):138-156.
- 414 [16] Jing L, Wu N, Zhang J, et al. Protective effect of 5,6,7,8-Tetrahydroxyflavone on
- 415 high altitude cerebral edema in rats[J]. Eur J Pharmacol. 2022,928:175121.
- 416 [17] Su Y, Zhang W, Zhang R, et al. Activation of Cholinergic Anti-Inflammatory
- 417 Pathway Ameliorates Cerebral and Cardiac Dysfunction After Intracerebral
- Hemorrhage Through Autophagy[J]. Front Immunol. 2022,13:870174.

- [18] Hu S, Lee H, Zhao H, et al. Inflammation and Severe Cerebral Venous
 Thrombosis[J]. Front Neurol. 2022 ,13:873802.
- 421 [19] Trofimov AO, Trofimova SY, Agarkova DI, et al. Intracranial dynamics
 422 biomarkers at traumatic cerebral vasospasm[J]. Brain Spine. 2023, 4:102727.
- 423 [20] Zadka Y, Doron O, Rosenthal G, et al. Mechanisms of reduced cerebral blood
- flow in cerebral edema and elevated intracranial pressure[J]. J Appl Physiol (1985).
 2023 ,134(2):444-454.
- 426 [21] Zhu P, Cheng K, He M, et al. Diagnostic value of congenital pulmonary airway
- 427 malformation volume ratio for fetal hydrops due to congenital lung malformations: a
- 428 systematic review and meta-analysis[J]. Orphanet J Rare Dis. 2022, 17(1):213.
- [22] Zhang Y, Ye P, Zhu H, et al. Neutral polysaccharide from Gastrodia elata
 alleviates cerebral ischemia-reperfusion injury by inhibiting ferroptosis-mediated
 neuroinflammatio n via the NRF2/HO-1 signaling pathway[J]. CNS Neurosci Ther.
 2024,30(3):e14456.
- [23] Zhang W, Dong Y, Sun C. Gedunin induces apoptosis and inhibits
 HMBG1/PI3K/AKT signaling pathways in a rat model of gastric carcinogenesis
 induced by methylnitronitrosoguanidine[J]. Archives of Medical Science.
 2024,20(2):691-697.
- [24] Hou J, Chen X, Xia J, et al. Down-regulation of PM20D1 is associated with
 clinical outcomes and prognosis of pregnant patients with diabetes
 mellitus[J]. Archives of Medical Science. 2023 ,19(6):1701-1708.
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clinical features		Training set	Training set Verification set statistica		Р
		(n=210)	(n=90)	values	value
Age (y	Age (years)		62.31±6.87	0.682	0.496
.	man	118 (56.19)	49 (54.44)	0.079	0 790
gender	woman	92 (43.81)	41 (45.56)	0.078	0./80
BMI (k	g/m2)	23.00±3.08	22.41±2.86	1.568	0.118
Course of hy	pertension	7 00 1 4 0 1	7.0(+4.21	1 500	0 112
(yea	ar)	/.88±4.01	/.00±4.21	1.390	0.113
Smoking	have	79 (37.62)	36 (40.00)	0.151	0.000
history	without	131 (62.38)	54 (60.00)	0.151	0.698
Drinking	have	68 (32.38)	27 (30.00)	0.165	0.685
history	without	142 (67.62)	63 (70.00)	0.165	
IL-6 (pg	IL-6 (pg/mL)		26.48±9.46	0.735	0.463
TNF-a (1	ng/mL)	1.13±0.49	1.04±0.37	1.825	0.069
IL-1 β (p	IL-1 β (pg/mL)		14.04±5.21	1.014	0.311
IL-8 (pg/mL)		17.41±6.05	17.25±6.11	0.201	0.840
IL-10 (pg/mL)		14.73±3.84	14.07±2.72	1.704	0.090
MCP-1 (ng/mL)		4.82±2.09	4.57±2.11	0.973	0.331
MIP-1a (ng/mL)		3.43±1.59	3.71±1.68	1.379	0.169
CBF (mL/(100g·min))		38.49±8.19	37.88±7.26	0.613	0.540
CVR ((mmHg·min)/mL)		1.31±0.40	1.34 ± 0.37	0.511	0.610
HMGB1 (ng/mL)	11.09±3.87	11.86±3.75	1.610	0.108
CRP (n	ng/L)	7.06±2.78	7.21±2.44	0.468	0.640

441 Table 1 Comparison of clinical characteristics between training set and verification set

442 Note:BMI: Body Mass Index; IL-6: Interleukin-6; TNF-α: Tumor Necrosis Factor-α;
443 IL-1β: Interleukin-1β; IL-8: Interleukin-8; IL-10: Interleukin-10; MCP-1: Monocyte
444 Chemoattractant Protein-1; MIP-1α: Macrophage Inflammatory Protein-1α; CBF:
445 Cerebral Blood Flow; CVR: Cerebrovascular Reactivity; HMGB1: High-Mobility
446 Group Box 1 Protein; CRP: C-reactive Protein

clinical features		Ed., (~ 100)	No edema	statistical	Р
		Edema (n=100)	(n=110)	values	value
Age (y	Age (years)		61.12±6.88	1.227	0.221
1	man	60 (60.00)	58 (52.73)	1 1 2 5	0.289
gender	woman	40 (40.00)	52 (47.27)	1.125	
BMI (k	g/m2)	22.88±3.21	23.12±2.98	0.563	0.574
Course of hy	pertension	0.0014.10	7 57 1 2 00	1 104	0.024
(yea	ar)	8.23±4.12	7.57±3.89	1.194	0.234
Smoking	have	44 (44.00)	35 (31.82)	2 2 1 2	0.070
history	without	56 (56.00)	75 (68.18)	3.312	0.069
Drinking	have	38 (38.00)	30 (27.27)	0.752	0.007
history	without	62 (62.00)	80 (72.73)	2.753	0.097
L-6 (pg/mL)		29.67±10.23	25.35±10.02	3.082	0.002
TNF- α (1	TNF- α (ng/mL)		1.02±0.42	3.307	0.001
IL-1 β (pg/mL)		14.56±5.12	12.34±4.56	3.320	0.001
IL-8 (pg/mL)		18.23±6.78	16.67±5.23	1.855	0.065
IL-10 (pg/mL)		14.19±3.45	15.23±4.12	1.972	0.050
MCP-1 (1	ng/mL)	5.13±2.12	4.56±2.03	1.955	0.052
MIP-1a (1	ng/mIL)	3.67±1.89	3.23±1.23	1.984	0.049
CBF (mL/(1	00g·min))	36.56±8.12	40.23±7.89	3.320	0.001
CVR ((mmHg·min)/mL)		$1.40{\pm}0.45$	1.24±0.32	3.075	0.002
HMGB1 (ng/mL)	12.03±4.32	10.23±3.21	3.423	0.001
CRP (n	ng/L)	7.63±3.12	6.56±2.34	2.795	0.006

447 Table 2 Comparison of clinical characteristics between edema group and non-edema448 group

cerebral hemorrhage

factor	В	S.E.	Wald	Р	OR	95%CI
 L-6	0.054	0.017	10.167	0.001	1.056	1.021-1.092
TNF-α	1.002	0.368	7.407	0.006	2.724	1.324-5.606
IL-1β	0.109	0.037	8.766	0.003	1.115	1.037-1.198
MIP-1a	0.205	0.111	3.427	0.064	1.228	0.988-1.525
CBF	-0.065	0.021	9.316	0.002	0.937	0.899-0.977
CVR	1.292	0.433	8.906	0.003	3.639	1.558-8.498
HMGB1	0.110	0.043	6.475	0.011	1.116	1.025-1.214
MIP-1a	0.156	0.062	6.241	0.012	1.169	1.034-1.320
 constant	-6.416	1.532	17.536	0.002		

-05

467	Figure Legends
468	Figure 1 Nomogram of delayed cerebral edema prediction model after cerebral
469	hemorrhage. Note: X1-X7 are L-6, TNF- α , IL-1 β , CBF, CVR, HMGB1 and CRP
470	respectively.
471	
472	Figure 2 Calibration curve of postoperative bleeding prediction model (A is training
473	set and B is verification set).
474	
475	Figure 3 ROC curve (A is training set and B is verification set).
476	
477	Figure 4 Decision Curve (A is training set and B is verification set).
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480	
481	



0.4 0.6 1-Specificity (FPR)

0.4

0.2

0.8

1.0



Figure 1 Nomogram of delayed cerebral edema prediction model after cerebral hemorrhage. Note: X1-X7 are L-6, TNF- α , IL-1 β , CBF, CVR, HMGB1 and CRP respectively.



Figure 2 Calibration curve of postoperative bleeding prediction model (A is training set and B is verification set).



Figure 3 ROC curve (A is training set and B is verification set).



Figure 4 Decision Curve (A is training set and B is verification set).