

Diagnostic value of cerebrospinal fluid chemokine ligand 13 (CXCL13) levels for viral and autoimmune encephalitis

Karol Perlejewski^{1*}, Dominika Studnicka¹, Agnieszka Pawetczyk¹, Henryk Jurasz¹, Tomasz Dzieciatkowski², Marcin Paciorek³, Beata Zakrzewska-Pniewska⁴, Paulina Emmel¹, Andrzej Horban³, Tomasz Laskus³, Marek Radkowski¹

¹Department of Immunopathology of Infectious and Parasitic Diseases, Medical University of Warsaw, Poland

²Department of Microbiology, Medical University of Warsaw, Poland

³Department of Adults Infectious Diseases, Medical University of Warsaw, Poland

⁴Department of Neurology, Medical University of Warsaw, Poland

Submitted: 24 September 2024; **Accepted:** 2 July 2025

Online publication: 23 August 2025

Arch Med Sci 2026; 22 (2): 841–847

DOI: <https://doi.org/10.5114/aoms/207801>

Copyright © 2025 Termedia & Banach

***Corresponding author:**

Karol Perlejewski
Department of
Immunopathology of
Infectious and
Parasitic Diseases
Medical University
of Warsaw
Pawinskiego 3c
02-106 Warsaw, Poland
E-mail: kperlejewski@wum.edu.pl

Abstract

Introduction: Chemokine ligand 13 (CXCL13) has been reported to be a valuable diagnostic biomarker in Lyme neuroborreliosis (LNB). However, its utility in the diagnostics of viral and autoimmune (AE) encephalitis still remains unclear.

Material and methods: We measured CXCL13 concentrations in cerebrospinal fluid (CSF) samples collected from 21 patients with viral encephalitis (17 cases of herpes simplex viral (HSV) and 4 of enteroviral (EV) encephalitis) and 6 patients with AE (5 subjects with antibodies anti-NMDAR and 1 with anti-GABA) and compared them to those found in patients with LNB (7 subjects) and multiple sclerosis (8 cases) as well as ten control subjects without neuroinflammation.

Results: Patients with neuroinflammation had a mean level of CXCL13-CSF of 105 pg/ml compared to 29 pg/ml in controls. The highest mean level of CXCL13 in CSF was detected in LNB patients (233 pg/ml), and the lowest in controls (29 pg/ml). Significant upregulation of CXCL13-CSF levels in LNB patients was observed in comparison to viral encephalitis and MS patients as well as controls. A positive correlation between elevated chemokine levels and cell count in CSF was found in all patients ($r = 0.6496$; $p < 0.0001$), as well as in the LNB group when tested alone ($r = 0.8428$; $p = 0.0173$). A positive correlation with CSF protein levels was observed in all patients ($r = 0.7216$; $p < 0.0001$), and separately in LNB ($r = 0.8573$; $p = 0.0137$) and AE patients ($r = 0.8885$; $p = 0.0180$).

Conclusions: The findings support the utility of CXCL13 measurements in CSF for LNB diagnosis. No specific patterns in CXCL13-CSF levels were associated with viral or autoimmune encephalitis.

Key words: CXCL13, viral encephalitis, autoimmune encephalitis, Lyme neuroborreliosis.

Introduction

Despite recent improvements in clinical management of encephalitis, the syndrome still remains challenging due to its potentially severe

course and risk of mortality [1]. In more than 50% of cases, the etiologic factor of neuroinflammation remains undetermined due to the vast number of infectious and noninfectious causative agents, resulting in often time-consuming and complicated diagnostic procedures [2–5]. Moreover, different pathogens may generate similar clinical pictures, such as Lyme neuroborreliosis (LNB) and enteroviral (EV) encephalitis [6]. The leading causes of CNS inflammation are viral infections (45–69% of all cases), mostly triggered by human herpesvirus type 1 (HSV) and enteroviruses, which constitute up to 24% and 6–25% of confirmed encephalitis cases, respectively [5, 7, 8]. Up to 20% of encephalitis cases are immune-mediated, with the presence of anti-neuronal or anti-gial antibodies, in particular autoantibodies against N-methyl-D-aspartate receptor (anti-NMDAR) [9].

Due to the variety of encephalitis types, it is vital to develop new tests and biomarkers which will allow for early diagnosis and their differentiation. Chemokine (C-X-C motif) ligand 13 (CXCL13) seems to be a new and promising biomarker, as its utility has already been confirmed in LNB diagnostics [2, 10, 11]. High chemokine concentrations in cerebrospinal fluid (CSF) were found to be strongly associated with the early phase of acute LNB and correlated with changes of typical CSF neuroinflammation markers (e.g. pleocytosis and CSF protein levels) [12]. Changes in CXCL13-CSF concentrations were also suggested to serve as a prognostic biomarker for multiple sclerosis (MS) progression [13], while CXCL13 serum levels were found to be associated with treatment response in MS [14, 15]. Among other immune diseases, CXCL13 was proposed as a possible marker and future therapeutic target [16] in systemic lupus erythematosus [17], rheumatoid arthritis [18], Sjögren's syndrome [19], myasthenia gravis [20], and inflammatory bowel disease (IBV) [21]. CXCL13-CSF levels as high as in LNB were also reported in neurosyphilis, and therefore it was proposed to use it as a diagnostic marker useful in patients also infected with human immunodeficiency virus (HIV), as the virus itself might trigger pleocytosis [22–24].

In this study, we analyzed CXCL13-CSF levels in viral and autoimmune encephalitis (AE), as well as in LNB and MS patients, to establish its possible diagnostic role in these types of encephalitis.

Material and methods

We analyzed 52 adult (≥ 18 years) patients who were admitted to two Warsaw hospitals: i) patients with encephalitis and LNB were hospitalized in the Hospital for Infectious Diseases in Warsaw; ii) MS patients and control subjects were hospitalized in the Department of Neurology,

Medical University of Warsaw. Written informed consent was obtained from all study participants or from their relatives if the participant was unable to provide consent. The study was approved by the Internal Review Board of the Medical University of Warsaw.

Encephalitis

Viral encephalitis patients consisted of subjects from a previously described surveillance study focused on the etiology of encephalitis [8]. Encephalitis was defined by the presence of an altered mental status, decreased level of consciousness, seizures or focal neurological signs, together with at least one abnormality in the cerebrospinal fluid (CSF): white blood cell count ≥ 4 cells/mm² and/or protein level ≥ 40 mg/dl [8]. Only patients with confirmed encephalitis caused by HSV and EV were included in the study. Viral etiology was determined based on positive viral detection performed in CSF using non-commercial PCRs [25, 26].

Lyme neuroborreliosis

Patients with LNB were part of a previously described study in which CXCL13-CSF levels were measured [6]; however, for the purpose of normalization, these measurements were performed once again in the same run with other samples analyzed in the present study.

LNB was diagnosed according to the guidelines recommended by the European Federation of Neurological Societies (EFNS) [27]. Besides clinical criteria, *B. burgdorferi* s.l. specific antibodies were detected in paired sera and CSF samples using *Borrelia* IgM/IgG ELISA tests (Biomedica, Vienna, Austria). Positive ELISA detections were confirmed by Western blot (recomLine *Borrelia* IgM and recomLine *Borrelia* IgG; Mikrogen Diagnostik, Neuried, Germany).

PCR detecting the *Borrelia* flagellin gene (flaB) was employed as described previously [28]. In short, DNA was extracted from 200 μ l of serum and CSF samples with the DNeasy Blood & Tissue Kit (Qiagen, Hilden, Germany) and then subjected to amplification using GeneProof *Borrelia* PCR Kit IVD (GeneProof, Brno, Czech Republic) with the following PCR conditions: initial denaturation at 95°C for 5 min followed by 35 cycles of denaturation at 95°C for 30 s, annealing at 52°C for 30 s, elongation at 72°C for 80 s, and a final elongation at 72°C for 7 min. The second round of PCR included 35 cycles of denaturation at 95°C for 20 s, followed by annealing at 55°C for 20 s, and elongation at 72°C for 60 s. Amplification was visualized on 1.5% agarose gel and, if positive, products were commercially sequenced and analyzed using MEGA ver.11 [29].

Among 7 patients with LNB included in the study, three were classified as confirmed and four as possible LNB based on the combination of clinical and diagnostics results described previously [6].

Autoimmune encephalitis

The study group included previously described subjects tested for the presence of AE autoantibodies in CSF: NMDAR, contactin-associated protein 2 (CASPR2), glutamate receptors (type AMPA1/2), leucine-rich glioma-inactivated protein 1 (LGI1), dipeptidyl aminopeptidase-like protein 6 (DPPX) and GABA B receptor [30]. Antibodies were identified using the Autoimmune Encephalitis Mosaic 6 diagnostic kit (Euroimmun, Germany) supported by the Nikon Eclipse 80i (Nikon, Japan) microscope, employed to read fluorescence under the magnifications of $\times 20$ and $\times 40$.

Autoimmune-triggered encephalitis was confirmed in 6 patients. In 5 (83.33%) of them, antibodies against NMDAR were detected in CSF, whereas one had anti-GABA.

Multiple sclerosis and control patients

Eighty-three patients admitted to the hospital with initial diagnosis of MS after further examination were divided into two groups: patients with

confirmed MS and the control group defined as subjects without CNS inflammation. MS patients were diagnosed according to the revised McDonald criteria introduced in 2017 [31], with lumbar puncture being performed as part of their routine diagnostics.

Eight patients were diagnosed with MS and 10 were classified as controls in the study. Among control patients, one remained undiagnosed, while 4 cases had peripheral neuropathy, 3 had peripheral vertigo, and 2 patients had retinopathy.

CXCL13 measurements

CXCL13 in CSF was measured according to the instructions provided by the manufacturer using CXCL13 ELISA (Euroimmun, Lübeck, Germany). Optical density (OD) was analyzed on a Multiskan FC (Thermo Fisher Scientific, Waltham, MA, USA) photometer. Based on the manufacturer's recommendations, the following levels of CXCL13-CSF corresponded to a specific interpretation: < 20 pg/ml – normal values, values from 20 to 29 pg/ml – borderline range, values 30–100 pg/ml – elevated, and > 100 pg/ml – LNB if corresponding clinical symptoms are present. Statistical comparisons between groups were performed using analysis of variance (ANOVA), while potential correlations were evaluated using Pearson's test.

Table I. Clinical and laboratory data in patients with viral encephalitis

Parameter	Viral encephalitis (n = 21)	AE (n = 6)	LNB (n = 7)	MS (n = 8)	All patients with neuroinflammation (n = 42)	Controls (n = 10)
Age [years] median (range)	36 (25–80)	41 (20–63)	39 (25–64)	34 (23–59)	36 (20–80)	30 (20–56)
CSF analysis:						
Cytosis in 1 μ l; mean (range)	87 (1–1225)	67 (2–208)	116 (1–602)	7 (3–13)	76 (1–1225)	2 (1–6)
% of lymphocytes; mean (range)	77 (43–97)	72 (12–100)	93 (83–100)	87 (84–91)	81 (12–100)	39 (14–80)
Protein [mg/dl]; mean (range)	55 (16–178)	61 (19–142)	102 (23–232)	30 (18–48)	60 (16–232)	34 (18–53)
Detected viruses, n (viral load in copies/ml)						
HSV	17 (60–341)	–	–	–	–	–
EV	4 (12–60)	–	1 (270)	–	–	–
AE antibody in CSF; n (%)						
Anti-NMDAR	–	5 (83.33)	–	–	–	–
Anti-GABA	–	1 (16.67)	–	–	–	–
CXCL13 levels in CSF [pg/ml]; mean (range)						
	72 (12–377)	153 (23–289)	233 (23–657)	46 (23–76)	105 (38–657)	29 (17–45)
Number of patients with CXCL13 levels in CSF above 100 pg/ml (%)						
	4 (19)	3 (50)	4 (57)	0	11 (26)	0

LNB – Lyme neuroborreliosis, MS – multiple sclerosis, AE – autoimmune encephalitis and controls – subjects without neuroinflammation.

Results

Overall, 52 patients were included in the study, consisting of 18 women and 34 men with a mean age of 39 years. Subjects were divided into two major groups: patients with neuroinflammation (42 patients) and controls – 10 subjects without neuroinflammation, neuroinfection, or demyelination.

Among 42 cases of patients with neuroinflammation, 21 were classified as viral encephalitis; in 17 (81%) and in 4 (19%), the causative agent of inflammation was determined to be HSV and EV, respectively. Viral loads of HSV in CSF ranged from 60 to 341 copies per milliliter and were higher than those found for EV (12–60 copies/ml) (Table I). In 1 patient with confirmed LNB, EV was detected with viral load of 220 copies/ml.

Among all analyzed subgroups, LNB patients had the highest mean values of cytosis (116 cells/ μ l) and protein (102 mg/dl) in CSF, as well as the percentage of detected lymphocytes (93%). Based on routine CSF analysis, the mean cytosis was 7 and 2 cells/ μ l in the MS group and controls, respectively.

Patients with neuroinflammation had a mean level of CXCL13-CSF of 105 pg/ml compared to a value of 29 pg/ml in controls (Table I). The highest mean level of CXCL13 in CSF was detected in LNB patients (233 pg/ml), followed by AE (153 pg/ml), viral encephalitis cases (72 pg/ml), patients with MS (46 pg/ml), and controls (29 pg/ml).

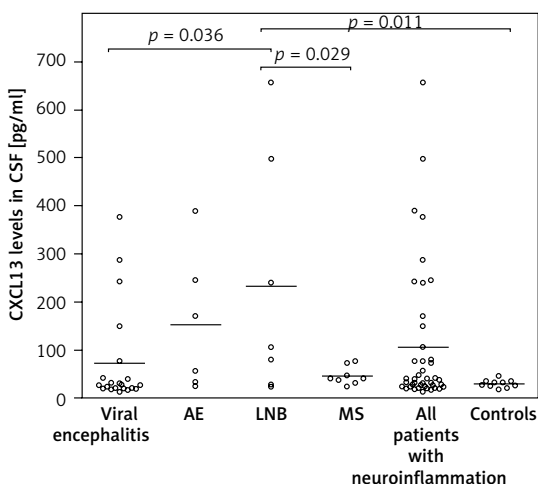


Figure 1. Concentration of CXCL13 in CSF (pg/ml) in patients with neuroinflammation ($n=42$) and controls (10 subjects without neuroinflammation). Patients were divided into the following groups: viral encephalitis ($n = 21$), autoimmune encephalitis (AE; $n = 6$), Lyme neuroborreliosis (LNB; $n = 7$), and multiple sclerosis (MS; $n = 8$) cases. ANOVA ($p = 0.0093$) was used to analyze the differences between the groups, and all significant results are shown ($p < 0.05$ in Tukey-Kramer test). Horizontal lines indicate mean CXCL13-CSF levels in each group

The threshold criterion of > 100 pg/ml for the chemokine in CSF was met for four LNB and viral encephalitis patients each, as well as in three AE cases. Significant ($p < 0.05$) upregulation of CXCL13-CSF levels in LNB patients was observed in comparison to viral encephalitis and MS patients as well as controls (Figure 1).

A positive correlation between elevated chemokine levels and cell count in CSF was found in all patients ($r = 0.6496$; $p < 0.0001$), as well as in the LNB group when tested alone ($r = 0.8428$; $p = 0.0173$). Similar results were obtained when CXCL13 levels were analyzed together with CSF protein levels. A positive correlation was observed in all patients ($r = 0.7216$; $p < 0.0001$), and separately in LNB ($r = 0.8573$; $p = 0.0137$) and AE patients ($r = 0.8885$; $p = 0.0180$) (Table I).

Discussion

CXCL13, previously known as B cell attracting chemokine 1 (BCA-1) [32] is constitutively expressed by dendritic cells (DC), monocytes, macrophages, and stromal cells in secondary lymphoid organs such as spleen, lymph nodes, and Peyer's patches [33, 34]. The chemokine acts via the cognate CXCR5 receptor (Burkitt's lymphoma receptor; CD185), which is expressed on mature B lymphocytes, DC, and T follicular helper cells (Tfh) [35]. CXCL13 plays a primary role in the development and organization of lymphoid tissues, as it regulates migrations of B cells and subsets of T cells to lymphoid follicles [36]. Furthermore, the CXCL13/CXCR5 axis seems to play a role in the formation of ectopic lymphoid tissues within the CNS, and therefore it is involved in the development of neuroinflammation [10].

In the present study we analyzed the CXCL13-CSF level in patients with viral and autoimmune encephalitis, as well as in those with LNB and MS in whom they were previously reported to be elevated, to verify its diagnostic utility for these types of neuroinflammation.

The highest chemokine levels in CSF have been detected in LNB patients but widely ranged from 23 to 657 pg/ml. Although elevated, CXCL13-CSF concentrations found in our study were lower than those described for untreated acute LNB (mean value of 15,149 pg/ml) [37] or observed by Erhart *et al.*, where the median CXCL13 level was 3,920 pg/ml [38]. The most likely explanation is that CXCL13 levels are high when tested immediately after sample collection and they decrease with the CSF storage time [22, 37].

Although higher CXCL13-CSF mean concentrations were found in 42 patients with neuroinflammation compared to controls, the difference was not significant. However, significant differences in CXCL13-CSF levels were observed between LNB

patients and controls, MS patients, and viral encephalitis patients. Our data are consistent with the results published by other research groups [37, 39–41], and confirm the utility of CXCL13-CSF testing as a valuable biomarker of LNB. Our results also support the claim that CXCL13-CSF can be helpful to differentiate LNB from other types of neuroinflammation, at the same time showing that its upregulation cannot be ruled out in case of single encephalitis patients [38]. Furthermore, we observed that the CSF chemokine levels were above recommended thresholds in 19% and 50% of viral and autoimmune encephalitis cases, respectively.

Viral encephalitis subjects included in the study encompassed patients with confirmed HSV and EV neuroinfection. Only a few previous studies have analyzed CXCL13-CSF in viral encephalitis cases separately; most have instead combined viral and bacterial cases together as “infectious encephalitis” [42].

Current data on CXCL13 activity in CNS in viral encephalitis are inconsistent. In our study, CXCL13-CSF levels did not significantly differ from controls. In contrast, Smíšková *et al.* reported that median CSF levels of the chemokine were significantly higher in patients with viral encephalitis, including HSV, EV, and tick-borne encephalitis (TBE) cases, compared to controls [43]. In another study, significantly higher CXCL13-CSF levels were also reported in patients with encephalitis caused by HSV and varicella zoster virus (VZV) than in controls consisting of subjects with non-inflammatory neurological diseases [38]. In contrast, viral encephalitis patients infected by human herpesvirus 6, VZV, EV, and HSV were found to have similar CXCL13-CSF levels as controls [44].

A high proportion (50%) of AE patients had CXCL13-CSF upregulation; however, chemokine concentrations did not significantly differ from those in controls, which could be explained by the small AE sample size used in the study. In one of the largest retrospective studies (conducted on 167 patients with anti-NMDAR encephalitis), elevated ($p < 0.001$) CXCL13 levels in CSF were reported when compared to 25 patients with non-inflammatory disorders [45]. Significant upregulation was also described by Kothur *et al.* in pediatric patients with anti-NMDAR encephalitis, acute disseminated encephalomyelitis (ADEM), or EV encephalitis in comparison to 20 non-inflammatory neurological controls mostly consisting of cases with cerebral palsy monogenic movement disorders [46]. It was found that CXCL13 levels in CSF were the highest at an early stage of anti-NMDAR encephalitis and they gradually decreased over time, which could explain the outcome of our investigation, as some of our patients had chronic AE [47].

Although specific diagnostic patterns were not identified in our study, the previously reported characteristic changes of CXCL13-CSF levels in autoimmune and viral encephalitis may indicate a meaningful marker of CNS immune activation, still warranting continued research [46, 48]. In AE, this could help identify patients experiencing immunologically active phases of disease and potentially guide decisions regarding immunotherapy [9, 49]. Similarly, in viral encephalitis, early CXCL13 increase in serum/CSF may assist in confirming a robust inflammatory response and help monitor the resolution of infection, though the overlap in levels across conditions underscores the need for cautious interpretation and combination with other diagnostic modalities [50, 51].

Finally, in our study we have observed a strong positive correlation between pleocytosis and CXCL13-CSF levels, both in LNB patients and in all analyzed patients with neuroinflammation, which is similar to other data [38, 43, 44]. No such association was present in viral and autoimmune encephalitis, while these were reported in some other studies [38, 48]. On the other hand, a positive correlation was found between CXCL13 and protein levels in CSF when all patients were analyzed, as well as in LNB and AE cases separately. This correlation was reported in the past in relation to LNB; however, to our knowledge, there have been no such reports for AE [12].

The major limitation of the study is the relatively small study group. However, all analyzed patients are clinically well defined and represent diverse types of different neuroinflammation. A strength of the study is the well-selected control group, which includes a comprehensive set of general CSF analysis results. The small sample size also makes it challenging to reliably interpret CXCL13 levels in CSF, especially given the lack of disease specificity, absence of standardized cut-off values, variability in testing methods, and overlap with other neuroinflammatory or non-inflammatory conditions [22, 38, 52]. Addressing these limitations in the future will require larger studies and efforts to standardize assays and diagnostic thresholds.

In conclusion, we have confirmed the utility of CXCL13 measurements in CSF for LNB diagnosis. No specific patterns in CXCL13-CSF levels were associated with viral and autoimmune encephalitis. Nevertheless, CXCL13-CSF upregulation cannot be excluded in individual cases. Therefore the parameter should be used with caution and only as a supplementary diagnostic marker.

Funding

No external funding.

Ethical approval

Approval number: KB/8/2015.

Conflict of interest

The authors declare no conflict of interest.

References

1. Feng G, Zhou L, Li F et al. Predictors of outcome in clinically diagnosed viral encephalitis patients: a 5-year prospective study. *Biomed Res Int* 2020; 2020: 2832418.
2. Du FZ, Zhang X, Zheng XL et al. Cerebrospinal fluid CXCL13 concentration for diagnosis of neurosyphilis: a systematic review and meta-analysis. *BMJ Open* 2024; 14: e078527.
3. Cellucci T, Van Mater H, Graus F, et al. Clinical approach to the diagnosis of autoimmune encephalitis in the pediatric patient. *Neurol Neuroimmunol Neuroinflamm* 2020; 7: e663.
4. Alam AM, Easton A, Nicholson TR, et al. Encephalitis: diagnosis, management and recent advances in the field of encephalitides. *Postgrad Med J* 2023; 99: 815-25.
5. Glaser CA, Honarmand S, Anderson LJ, et al. Beyond viruses: clinical profiles and etiologies associated with encephalitis. *Clin Infect Dis* 2006; 43: 1565-77.
6. Perlejewski K, Radkowski M, Pawelczyk A, et al. Enteroviral central nervous system infections in patients with Lyme neuroborreliosis. *Ticks Tick Borne Dis* 2023; 14: 102253.
7. Granerod J, Ambrose HE, Davies NW, et al. Causes of encephalitis and differences in their clinical presentations in England: a multicentre, population-based prospective study. *Lancet Infect Dis* 2010; 10: 835-44.
8. Popiel M, Perlejewski K, Bednarska A, et al. Viral etiologies in adult patients with encephalitis in Poland: a prospective single center study. *PLoS One* 2017; 12: e0178481.
9. Dutra LA, Abrantes F, Toso FF, et al. Autoimmune encephalitis: a review of diagnosis and treatment. *Arq Neuropsiquiatr* 2018; 76: 41-9.
10. Huber AK, Irani DN. Targeting CXCL13 during neuroinflammation. *Adv Neuroimmune Biol* 2015; 6: 1-8.
11. Lintner H, Hochgatterer-Rechberger P, Pischinger B, et al. Sensitivity and specificity of cerebrospinal fluid CXCL13 for diagnosing Lyme neuroborreliosis – a study on 1410 patients and review of the literature. *J Neurol Sci* 2020; 414: 116843.
12. Barstad B, Tveitnes D, Dalen I, et al. The B-lymphocyte chemokine CXCL13 in the cerebrospinal fluid of children with Lyme neuroborreliosis: associations with clinical and laboratory variables. *Infect Dis (Lond)* 2019; 51: 856-63.
13. Sellebjerg F, Bornsen L, Khademi M, et al. Increased cerebrospinal fluid concentrations of the chemokine CXCL13 in active MS. *Neurology* 2009; 73: 2003-10.
14. Fissolo N, Pappolla A, Rio J, et al. Serum levels of CXCL13 are associated with teriflunomide response in patients with multiple sclerosis. *Neurol Neuroimmunol Neuroinflamm* 2023; 10: e200050.
15. Karaaslan Z, Kurtuncu M, Akcay HI, et al. CXCL13 levels indicate treatment responsiveness to fingolimod in MS patients. *Eur Neurol* 2022; 85: 69-71.
16. Pan Z, Zhu T, Liu Y, Zhang N. Role of the CXCL13/CXCR5 axis in autoimmune diseases. *Front Immunol* 2022; 13: 850998.
17. Da Z, Li L, Zhu J, et al. CXCL13 promotes proliferation of mesangial cells by combination with CXCR5 in SLE. *J Immunol Res* 2016; 2016: 2063985.
18. Zhao J, Ye X, Zhang Z. The predictive value of serum soluble ICAM-1 and CXCL13 in the therapeutic response to TNF inhibitor in rheumatoid arthritis patients who are refractory to csDMARDs. *Clin Rheumatol* 2020; 39: 2573-81.
19. Nocturne G, Seror R, Fogel O, et al. CXCL13 and CCL11 serum levels and lymphoma and disease activity in primary Sjogren's syndrome. *Arthritis Rheumatol* 2015; 67: 3226-33.
20. Zhang M, Guo J, Li H, et al. Expression of immune molecules CD25 and CXCL13 correlated with clinical severity of myasthenia gravis. *J Mol Neurosci* 2013; 50: 317-23.
21. Singh UP, Singh NP, Murphy EA, et al. Chemokine and cytokine levels in inflammatory bowel disease patients. *Cytokine* 2016; 77: 44-9.
22. Rupprecht TA, Manz KM, Fingerle V, et al. Diagnostic value of cerebrospinal fluid CXCL13 for acute Lyme neuroborreliosis. a systematic review and meta-analysis. *Clin Microbiol Infect* 2018; 24: 1234-40.
23. Dersch R, Hottenrott T, Senel M, et al. The chemokine CXCL13 is elevated in the cerebrospinal fluid of patients with neurosyphilis. *Fluids Barriers CNS* 2015; 12: 12.
24. Marra CM, Tantalos LC, Sahi SK, et al. CXCL13 as a cerebrospinal fluid marker for neurosyphilis in HIV-infected patients with syphilis. *Sex Trans Dis* 2010; 37: 283-7.
25. Machura P, Gorka E, Mlynarczyk-Bonikowska B, et al. [Novel multiplex real-time PCR assay for detection and differentiation of herpes simplex virus type 1 and 2 DNA]. *Med Dosw Mikrobiol* 2015; 67: 125-32.
26. Les K, Przybylski M, Dzieciatkowski T, Mlynarczyk G. Detection of human enteroviruses with real-time PCR assay using TaqMan fluorescent probe. *Med Dosw Mikrobiol* 2010; 62: 245-53.
27. Mygland A, Ljostad U, Fingerle V, et al. EFNS guidelines on the diagnosis and management of European Lyme neuroborreliosis. *Eur J Neurol* 2010; 17: 8-16, e11-14.
28. Wodecka B. flaB gene as a molecular marker for distinct identification of *Borrelia* species in environmental samples by the PCR-restriction fragment length polymorphism method. *Appl Environ Microbiol* 2011; 77: 7088-92.
29. Tamura K, Stecher G, Kumar S. MEGA11: molecular evolutionary genetics analysis version 11. *Mol Biol Evol* 2021; 38: 3022-7.
30. Perlejewski K, Pawelczyk A, Bukowska-Osko I, et al. Search for Viral infections in cerebrospinal fluid from patients with autoimmune encephalitis. *Open Forum Infect Dis* 2020; 7: ofaa468.
31. Thompson AJ, Banwell BL, Barkhof F, et al. Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. *Lancet Neurol* 2018; 17: 162-73.
32. Sivina M, Xiao L, Kim E, et al. CXCL13 plasma levels function as a biomarker for disease activity in patients with chronic lymphocytic leukemia. *Leukemia* 2021; 35: 1610-20.
33. Legler DF, Loetscher M, Roos RS, et al. B cell-attracting chemokine 1, a human CXC chemokine expressed in lymphoid tissues, selectively attracts B lymphocytes via BLR1/CXCR5. *J Exp Med* 1998; 187: 655-60.
34. Carlsen HS, Baekkevold ES, Morton HC, et al. Monocyte-like and mature macrophages produce CXCL13 (B cell-attracting chemokine 1) in inflammatory lesions with lymphoid neogenesis. *Blood* 2004; 104: 3021-7.
35. Harrer C, Otto F, Radlberger RF, et al. The CXCL13/CXCR5 immune axis in health and disease-implications for in-

- trathecal B cell activities in neuroinflammation. *Cells* 2022; 11: 2649.
36. Forster R, Mattis AE, Kremmer E, et al. A putative chemokine receptor, BLR1, directs B cell migration to defined lymphoid organs and specific anatomic compartments of the spleen. *Cell* 1996; 87: 1037-47.
 37. Schmidt C, Plate A, Angele B, et al. A prospective study on the role of CXCL13 in Lyme neuroborreliosis. *Neurology* 2011; 76: 1051-8.
 38. Erhart DK, Klose V, Schaper T, et al. CXCL13 in cerebrospinal fluid: clinical value in a large cross-sectional study. *Int J Mol Sci* 2023; 25: 425.
 39. Remy MM, Schöbi N, Kottanattu L, et al. Cerebrospinal fluid CXCL13 as a diagnostic marker of neuroborreliosis in children: a retrospective case-control study. *J Neuroinflammation* 2017; 14: 173.
 40. Henningsson AJ, Gyllemark P, Lager M, et al. Evaluation of two assays for CXCL13 analysis in cerebrospinal fluid for laboratory diagnosis of Lyme neuroborreliosis. *APMIS* 2016; 124: 985-90.
 41. Wagner JN, Weis S, Kubasta C, et al. CXCL13 as a diagnostic marker of neuroborreliosis and other neuroinflammatory disorders in an unselected group of patients. *J Neurol* 2018; 265: 74-81.
 42. Pilz G, Steger R, Wipfler P, et al. Beyond LNB: real life data on occurrence and extent of CSF CXCL13 in neuroinflammatory diseases. *J Neuroimmunol* 2020; 338: 577087.
 43. Smiskova D, Dzupova O, Moravcova L, Picha D. Cerebrospinal fluid CXCL13 in non-borreliac central nervous system infections: contribution of CXCL13 to the differential diagnosis. *Infect Dis (Lond)* 2023; 55: 551-8.
 44. Hytönen J, Kortela E, Waris M, et al. CXCL13 and neopterin concentrations in cerebrospinal fluid of patients with Lyme neuroborreliosis and other diseases that cause neuroinflammation. *J Neuroinflammation* 2014; 11: 103.
 45. Leypoldt F, Hoftberger R, Titulaer MJ, et al. Investigations on CXCL13 in anti-N-methyl-D-aspartate receptor encephalitis: a potential biomarker of treatment response. *JAMA Neurol* 2015; 72: 180-6.
 46. Kothur K, Wienholt L, Mohammad SS, et al. Utility of CSF cytokine/chemokines as markers of active intrathecal inflammation: comparison of demyelinating, anti-NMDAR and enteroviral encephalitis. *PLoS One* 2016; 11: e016156.
 47. Liba Z, Kayserova J, Elisak M, et al. Anti-N-methyl-D-aspartate receptor encephalitis: the clinical course in light of the chemokine and cytokine levels in cerebrospinal fluid. *J Neuroinflammation* 2016; 13: 55.
 48. Tilea B, Voidazan S, Balasa R, et al. CXCL13 levels are more increased in cerebrospinal fluid and plasma of patients with acute infectious than in non-infectious diseases of the central nervous system. *Rev Rom Med Labor* 2017; 25: 63-73.
 49. Lin YT, Yang X, Lv JW, et al. CXCL13 is a biomarker of anti-leucine-rich glioma-inactivated protein 1 encephalitis patients. *Neuropsychiatr Dis Treat* 2019; 15: 2909-15.
 50. Zajkowska J, Moniuszko-Malinowska A, Pancewicz SA, et al. Evaluation of CXCL10, CXCL11, CXCL12 and CXCL13 chemokines in serum and cerebrospinal fluid in patients with tick borne encephalitis (TBE). *Adv Med Sci* 2011; 56: 311-7.
 51. Pilz G, Wipfler P, Otto F, et al. Cerebrospinal fluid CXCL13 indicates disease course in neuroinfection: an observational study. *J Neuroinflammation* 2019; 16: 13.
 52. Bremell D, Mattsson N, Edsbacke M, et al. Cerebrospinal fluid CXCL13 in Lyme neuroborreliosis and asymptomatic HIV infection. *Bmc Neurology* 2013; 13: 2.