

# Causality analysis of plasma fatty acids with pneumonia: identifying diagnostic biomarkers through transcriptome-wide association study

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## Keywords

Plasma Fatty acid, Pneumonia, Mendelian randomization, Biomarkers, Machine learning algorithms

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## Abstract

### Introduction

Fatty acids mediate pulmonary inflammation through cytokine regulation, interactions with the tryptophan-kynurenine pathway, and mediators like 18-HEPE that boost interferon- $\lambda$ . To systematically dissect these mechanisms and their translational implications, we pioneered a novel framework integrating bidirectional two-sample mendelian randomization (MR) with multi-algorithm machine learning.

### Material and methods

We obtained Single nucleotide polymorphisms (SNPs) significantly associated with eight plasma fatty acids from genome-wide association study (GWAS) summary statistics and used them as instrumental variables in bidirectional two sample MR across six pneumonia phenotypes (Pneumonia, Asthma related pneumonia, Bacterial pneumonia, Critical pneumonia, Viral pneumonia, and Lobar pneumonia). Causal estimates were calculated using inverse variance weighting (IVW), which combines SNP-specific Wald ratios weighted by the inverse of their variance, with MR-Egger and weighted median approaches for sensitivity analysis. Transcriptomic data were then analyzed by Lasso regression, support vector machine recursive feature elimination and random forest to identify fatty acid metabolism-related biomarker candidates.

### Results

MR analysis suggests potential causal associations between omega-6 fatty acids and critical pneumonia (OR:1.28, CI:1.01-1.61,  $P=0.038$ ), Linoleic acids (LA) and bacterial pneumonia (OR:0.85, CI:0.73-0.99,  $P=0.047$ ), and Docosahexaenoic fatty acids (DHA) and pneumonia (OR:0.83, CI:0.74-0.93,  $P=0.002$ ). Moreover, ACAA1 and OLAH, which are genes involved in fatty acid metabolism, were identified as potential candidate biomarkers for pneumonia.

### Conclusions

Our study employed MR analysis to establish a causal link between omega-6, LA and DHA with pneumonia. Additionally, through transcriptomic analysis, we identified plasma fatty acid metabolism-associated biomarkers that may serve as diagnostic indicators for pneumonia.

# **Causality analysis of plasma fatty acids with pneumonia: identifying diagnostic biomarkers through transcriptome-wide association study**

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## Abstract

**Background:** Fatty acids mediate pulmonary inflammation through cytokine regulation, interactions with the tryptophan-kynurenine pathway, and mediators like 18-HEPE that boost interferon- $\lambda$ . To systematically dissect these mechanisms and their translational implications, we pioneered a novel framework integrating bidirectional two-sample mendelian randomization (MR) with multi-algorithm machine learning. This approach not only quantifies causal relationships between fatty acids and inflammatory outcomes but also identifies clinically actionable diagnostic biomarkers.

**Methods:** We obtained Single nucleotide polymorphisms (SNPs) significantly associated with eight plasma fatty acids from genome-wide association study (GWAS) summary statistics and used them as instrumental variables in bidirectional two sample MR across six pneumonia phenotypes (Pneumonia, Asthma related pneumonia, Bacterial pneumonia, Critical pneumonia, Viral pneumonia, and Lobar pneumonia). Causal estimates were calculated using inverse variance weighting (IVW), which combines SNP-specific Wald ratios weighted by the inverse of their variance, with MR-Egger and weighted median approaches for sensitivity analysis. Transcriptomic data were then analyzed by Lasso regression, support vector machine recursive feature elimination and random forest to identify fatty acid metabolism-related biomarker candidates.

**Results:** MR analysis suggests potential causal associations between omega-6 fatty acids and critical pneumonia (OR:1.28, CI:1.01-1.61, P=0.038), Linoleic acids (LA) and bacterial pneumonia (OR:0.85, CI:0.73-0.99, P=0.047), and Docosahexaenoic fatty acids (DHA) and pneumonia (OR:0.83, CI:0.74-0.93, P=0.002). Moreover, ACAA1 and OLAH, which are genes involved in fatty acid metabolism, were identified as potential candidate biomarkers for pneumonia.

**Conclusions:** Our study employed MR analysis to establish a causal link between omega-6, LA and DHA with pneumonia. Additionally, through transcriptomic analysis, we identified plasma fatty acid metabolism-associated biomarkers that may serve as

diagnostic indicators for pneumonia.

**Keywords:** Plasma Fatty acid, Pneumonia, Mendelian randomization, Biomarkers, Machine learning algorithms

## Highlights

**1. Significant Associations Found:** MR analysis reveals potential causal associations between specific fatty acids and pneumonia subtypes.

**2. Machine-Learning for Biomarker Discovery:** Applies machine-learning algorithms to identify diagnostic biomarkers among differentially expressed genes.

**Novel Biomarker Candidates:** Identifies ACAA1 and OLAH, genes in fatty acid metabolism, as potential pneumonia biomarkers.

**Table (1):** List of Abbreviations

| Abbreviation | Definition  |
|--------------|---|
| MR           | Mendelian randomization                                     |
| GWAS         | Genome-wide association study                               |
| SNP          | Single nucleotide polymorphism                              |
| IVW          | Inverse variance weighting                                  |
| OR           | Odds ratio  |
| LA           | Linoleic acids  |
| DHA          | Docosahexaenoic acid  |
| PUFA         | Polyunsaturated fatty acid                                  |
| FA           | Fatty acid  |
| SFA          | Saturated fatty acid  |
| MUFA         | Monounsaturated fatty acids                                 |
| T2D          | Type 2 diabetes   |
| COPD         | Chronic obstructive pulmonary disease                       |
| BMI          | Body mass index   |
| IEU          | Integrative epidemiology unit                               |
| IV           | Instrumental variable                                       |
| MR-PRESSO    | Mendelian randomization pleiotropy residual sum and outlier |
| FDR          | False discovery rate  |
| GO           | Gene ontology   |
| KEGG         | Kyoto encyclopedia of genes and genomes                     |
| LASSO        | Least absolute shrinkage and selection operator             |
| SVM-RFE      | Support vector machine recursive feature elimination        |
| CIBERSORT    | Cell-type identification by estimating relative subsets of  |

|      |                                   |
|------|-----------------------------------|
|      | RNA transcripts                   |
| GSEA | Gene set enrichment analysis      |
| ROC  | Receiver operating characteristic |
| PCA  | Principal component analysis      |
| Treg | Regulatory T cell                 |
| NK   | Neutral killer                    |

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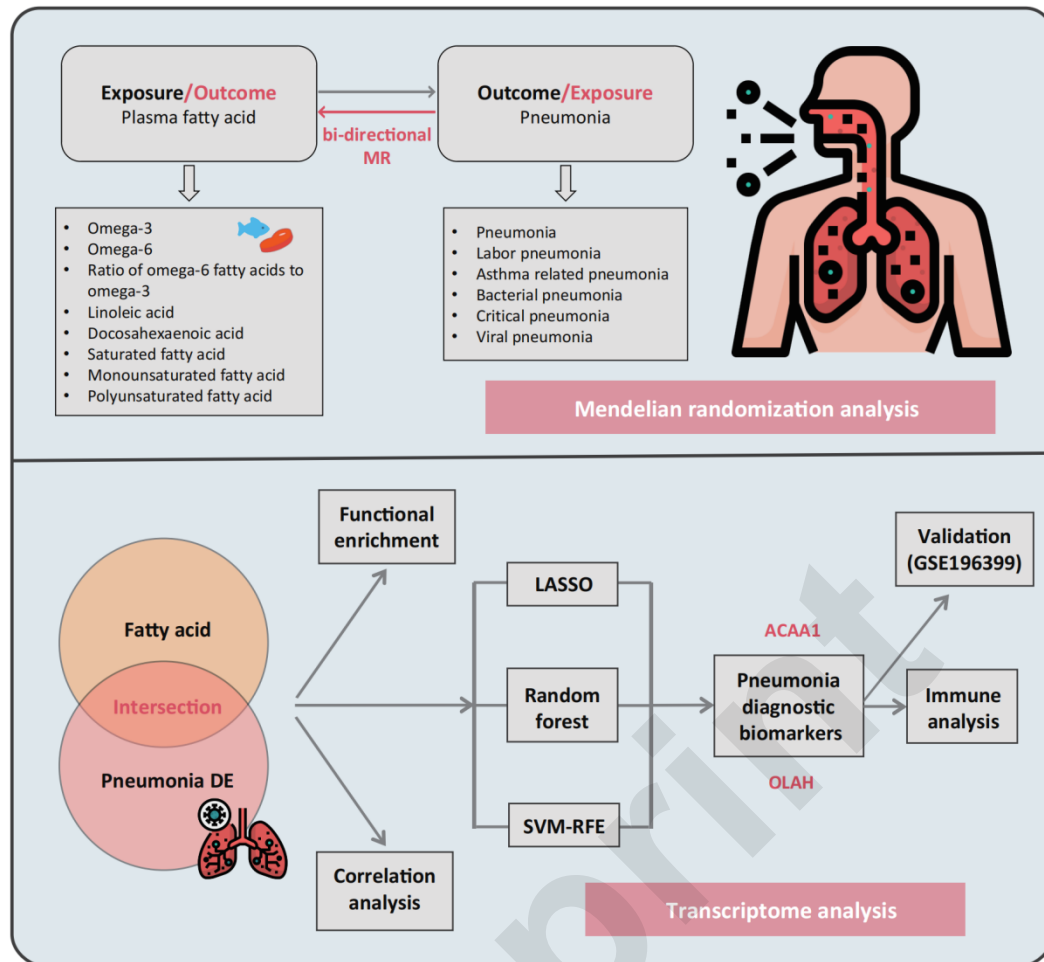
## Introduction

Pneumonia, a global health challenge driven by infectious pathogens, is aggravated by antibiotic resistance and chronic respiratory comorbidities [1]. This study focuses on pathogen-mediated mechanisms, distinguishing pneumonia from non-infectious pneumonitis caused by autoimmune disorders or chemical exposures [2]. Severe cases often progress to systemic complications, including coagulation abnormalities such as disseminated intravascular coagulation, distinct from coagulation necrosis [3]. These abnormalities synergize with cytokine storms, elevating cardiovascular mortality risk through endothelial dysfunction and myocardial suppression [4, 5]. Despite advances in therapy, pneumonia remains a leading global cause of morbidity and mortality, underscoring the urgency to elucidate pathogenic mechanisms and advance novel therapies [6].

Fatty acids are essential membrane phospholipids and bioactive mediators that regulate immune cell function and inflammatory signaling [7, 8]. Dysregulated lipid profiles, particularly imbalances in circulating polyunsaturated fatty acids (PUFAs), influence lung pathology through membrane-mediated immune cell dysfunction and the production of lipid signaling molecules [9]. This mechanistic duality is exemplified in chronic obstructive pulmonary disease, where skewed omega-3 to omega-6 PUFA ratios correlate with sustained airway inflammation [10]. The interplay extends to acute infectious contexts, as observed in severe pneumonia patients exhibiting activation of the tryptophan-kynurenine axis [11]. During this process, indoleamine 2,3-dioxygenase catalyzes tryptophan conversion into immunomodulatory kynurenines that suppress T-cell activity, establishing a regulatory network with PUFA-derived inflammatory mediators, a phenomenon inversely associated with protective PUFA ratios [11-13].

Experimental evidence underscores therapeutic opportunities, with murine models demonstrating omega-3 fatty acids attenuate bacterial pneumonia severity through coordinated cytokine modulation, reducing pro-inflammatory IL-6 while elevating anti-inflammatory IL-10 [14-16]. Parallel studies reveal butyrate-induced omega-3 metabolites exert antiviral effects via interferon- $\lambda$  induction in viral pneumonia models [17]. Despite these mechanistic insights, translational challenges persist in distinguishing observational associations from causal relationships, particularly regarding pneumonia-specific lipid signatures and targeted therapeutic strategies.

To address these gaps, this study employed bidirectional two-sample Mendelian randomization (MR) to causally link eight plasma fatty acids to six distinct pneumonia phenotypes. The workflow of this study was illustrated in Fig. (1). Building on MR-derived causal evidence, we further applied three complementary machine-learning algorithms to transcriptomic data, identifying diagnostic biomarkers of fatty acid metabolism. This integrated methodology synergistically combined genetic causality analysis with multi-algorithm biomarker identification, establishing unified framework in pneumonia research that bridged mechanistic discovery and clinical translation.



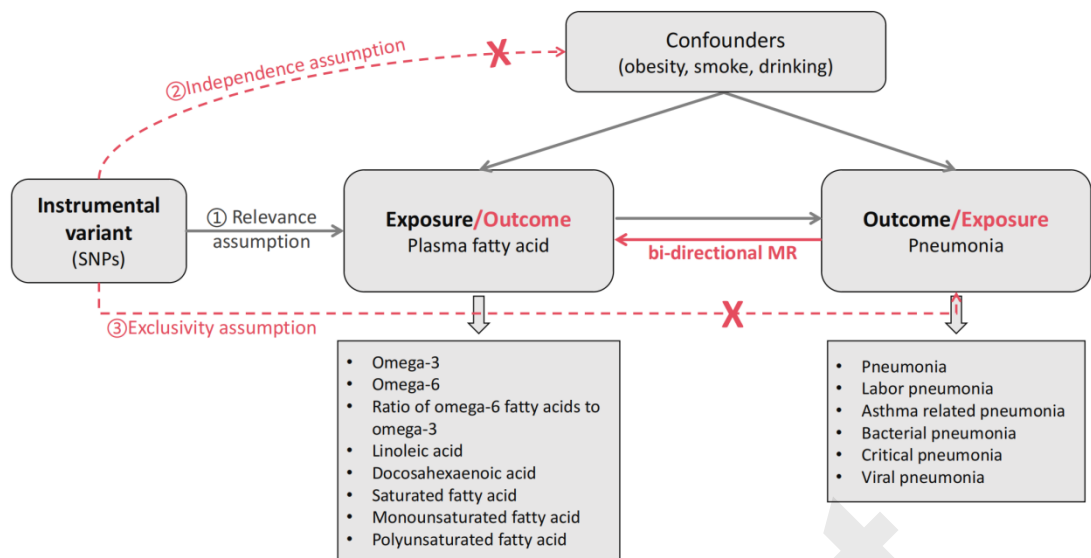
**Fig. (1): Work flow of this study.**

## Material and methods

### Study design

We first applied a two-sample MR framework to evaluate the causal effect of circulating plasma fatty acids on pneumonia risk. To address potential reverse causation, we then performed a reciprocal MR analysis [18], treating pneumonia liability as the exposure and plasma fatty acid levels as the outcome. In every MR analysis, we ensured that instrumental variables (IVs) satisfied the three core assumptions: strong association with the exposure, independence from confounders, and influence on the outcome exclusively through the exposure pathway [19]. To minimize bias from population stratification, all summary statistics were drawn from cohorts of European ancestry (Fig.

2).



**Fig. (2):** Flow chart of the Mendelian randomization study design.

## Data source

Genome-wide association study (GWAS) summary statistics for eight plasma fatty acid (FA) traits — omega-3, omega-6, omega-6/omega-3 ratio, linoleic acid (LA), docosahexaenoic acid (DHA), saturated fatty acids (SFA), monounsaturated fatty acids (MUFA), and PUFA—were obtained from the OpenGWAS platform. These data derive from 115,006 European ancestry participants in the UK Biobank. Pneumonia-related GWAS data were accessed via the IEU OpenGWAS repository and included overall pneumonia, severe pneumonia, lobar pneumonia, asthma-related pneumonia, viral pneumonia, and bacterial pneumonia. All summary statistics were drawn from cohorts of European ancestry, with detailed population descriptors provided in Table (2).

**Table (2):** Detailed information of the genome-wide association studies in our analysis.

| Phenotype | PMID<br>(ID) | Sample size<br>(case/control) | Population |
|-----------|--------------|-------------------------------|------------|
|-----------|--------------|-------------------------------|------------|



|  |                                  |                          |          |
|--|----------------------------------|--------------------------|----------|
| Omega-3 fatty acid levels                              | 35213538<br>(ebi-a-GCST90092931) | 115006<br>/              | European |
| Omega-6 fatty acid levels                              | 35213538<br>(ebi-a-GCST90092933) | 115006<br>/              | European |
| Ratio of omega-6 fatty acids to<br>omega-3 fatty acids | 35213538<br>(ebi-a-GCST90092934) | 115006<br>/              | European |
| Linoleic acid levels                                   | 35213538<br>(ebi-a-GCST90092880) | 115006<br>/              | European |
| Docosahexaenoic acid levels                            | 35213538<br>(ebi-a-GCST90092816) | 115006<br>/              | European |
| Saturated fatty acid levels                            | 35213538<br>(ebi-a-GCST90092980) | 115006<br>/              | European |
| Monounsaturated fatty acid<br>levels                   | 35213538<br>(ebi-a-GCST90092928) | 115006<br>/              | European |
| Polyunsaturated fatty acid<br>levels                   | 35213538<br>(ebi-a-GCST90092939) | 115006<br>/              | European |
| Pneumonia  | 34594039<br>(ebi-a-GCST90018901) | 480299<br>(16887/463412) | European |
| Asthma related pneumonia                               | /<br>(ukb-d-ASTHMA_PNEUMONIA)    | 361194<br>(5900/355294)  | European |
| Bacterial pneumonia                                    | /<br>(finn-b-J10_PNEUMOBACT)     | 196855<br>(7987/188868)  | European |
| Critical pneumonia                                     | /<br>(ieu-b-4978)                | 431365<br>(2758/428607)  | European |
| Viral pneumonia  | /<br>(finn-b-J10_VIRALPNEUMO)    | 189568<br>(700/188868)   | European |
| Lobar pneumonia  | /<br>(ukb-b-6576)                | 463010<br>(2359/460651)  | European |

## **Instrumental variants selection**

The criteria for selecting IVs were as follows: SNP Selection: SNPs were used as IVs, with a genome-wide significance threshold of  $P < 5e-08$ . However, for some pneumonitis-related traits, the number of SNPs meeting this threshold was limited. To increase the number of available SNPs, we relaxed the threshold to  $P < 5e-06$ , and for lobar pneumonia, a threshold of  $P < 5e-05$  was applied. Clumping: SNPs were clumped to remove the effect of linkage disequilibrium ( $r^2 < 0.001$ , with a region length of 10,000 kb). F-Statistic: The F-statistic was used to assess the strength of IVs. SNPs with an F-statistic  $> 10$  were retained, indicating strong instrument strength. Weak instruments were excluded. Allelic Consistency: SNPs associated with both exposure and outcome were aligned in terms of allelic direction. Palindromic SNPs and SNPs with inconsistent allelic directions were removed. MR-PRESSO Outliers: Outlier SNPs identified by the MR-PRESSO test were excluded from the analysis.

## **MR analysis**

The primary causal estimates were obtained using the inverse variance weighted (IVW) method under a random effects model, supplemented by MR-Egger regression, weighted median, weighted mode, and simple mode analyses to assess robustness. Prior to MR, any SNPs associated with known confounders (BMI, waist measures, smoking, alcohol) were excluded. For reverse MR, the same procedure was followed with pneumonia as the exposure and individual fatty acid traits as outcomes.

## **Sensitivity Analysis**

To assess the robustness of the identified associations, we conducted sensitivity analyses using three methods: heterogeneity test, horizontal pleiotropy test, and leave-one-out test. Cochran's Q test was used to assess SNP heterogeneity. If heterogeneity was detected ( $P < 0.05$ ), a random-effects IVW model was applied. The presence of horizontal pleiotropy was evaluated using MR-Egger regression, with statistical significance of the intercept ( $P < 0.05$ ) indicating pleiotropy. Additionally, MR-PRESSO was used to identify and remove outliers. A "leave-one-out" test was

performed to evaluate the impact of individual SNPs on the causal associations by sequentially excluding each SNP.

### **Transcriptomic Data Acquisition and Differential Expression**

We retrieved two gene expression datasets from the Gene Expression Omnibus (GEO): GSE40012 (training set: 16 pneumonia, 18 controls) and GSE196399 (validation set: 56 pneumonia, 21 controls). A curated list of 308 fatty acid metabolism-related genes was compiled from the MSigDB Hallmark, KEGG, and Reactome collections (version 2024.1). Differential expression between pneumonia and control samples was determined using the limma package in R, with thresholds of  $|\log_2 \text{fold change}| > 1$  and adjusted  $P < 0.05$  (Benjamini–Hochberg correction).

### **Functional enrichment Analysis**

Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses were conducted in R with the clusterProfiler package. We controlled for multiple testing using the Benjamini-Hochberg method and considered pathways with adjusted  $P < 0.05$  to be significantly enriched.

### **Feature Selection and Predictive Modeling**

To identify key fatty acid-related genes predictive of pneumonia, we applied three machine learning approaches: least absolute shrinkage and selection operator (LASSO) regression for initial feature retention, support vector machine recursive feature elimination (SVM-RFE) with ten-fold cross-validation for iterative refinement, and random forest for ranking variable importance. The intersection of genes selected by all three methods constituted the final feature set used in model construction.

### **Immune infiltration analysis**

Immune cell proportions were estimated using the CIBERSORT algorithm implemented in the IOBR R package, and single sample gene set enrichment analysis (ssGSEA) of immune-related functions was performed via the GSVA package.

Differences between pneumonia and control groups were tested by Wilcoxon rank sum test with  $P < 0.05$  denoting significance.

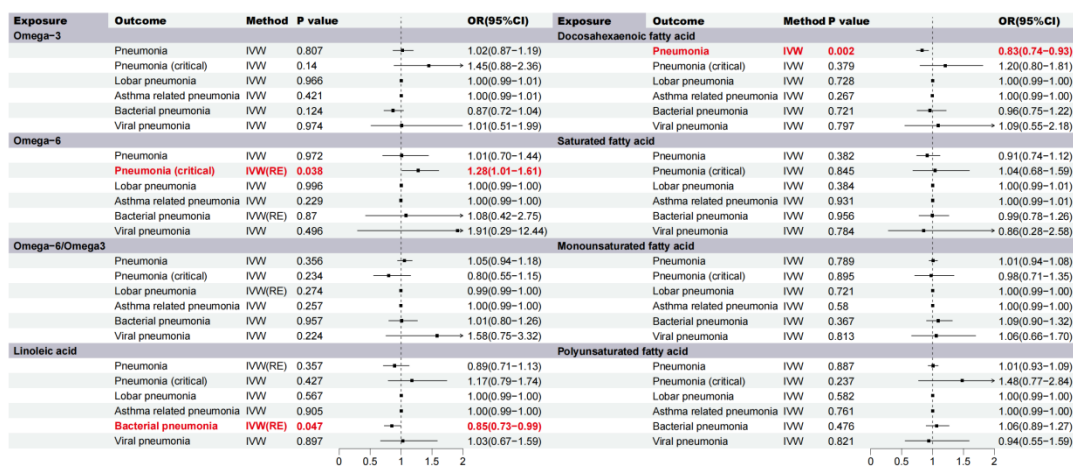
## Statistical Analysis

All analyses were executed in R version 4.4.1, utilizing the TwoSampleMR (0.6.8) [20], MRPRESSO (1.0) [21], limma [22], clusterProfiler [23] and IOBR [24] packages. Statistical significance was defined as two-sided  $P < 0.05$  throughout.

## Results

### Causal relationship between fatty acids and pneumonia

Following rigorous selection and harmonization of genetic IVs, along with MR-PRESSO-mediated removal of pleiotropic outliers, we employed IVW MR to estimate causal effects of eight plasma fatty acids on pneumonia risk. We observed that higher omega-6 levels were associated with increased risk of severe pneumonia (OR=1.28, 95% CI: 1.01-1.61,  $P=0.038$ ), whereas greater LA levels conferred protection against bacterial pneumonia (OR=0.85, 95% CI: 0.73-0.99,  $P=0.047$ ). Similarly, elevated DHA was inversely related to overall pneumonia incidence (OR=0.83, 95% CI: 0.74-0.93,  $P=0.002$ ). No statistically significant causal links were detected for the remaining fatty acids (Fig. 3). Consistent direction and magnitude across MR-Egger, weighted median, weighted and simple mode analyses reinforced these findings (Supplementary Table 1), and the complete lists of SNP instruments appear in Supplementary Tables (2-7) with corresponding scatter and forest plots in Supplementary Figures (1-2).



**Fig. (3): Univariable Mendelian randomization results of Plasma fatty acid on Pneumonia.**

### Sensitivity analysis

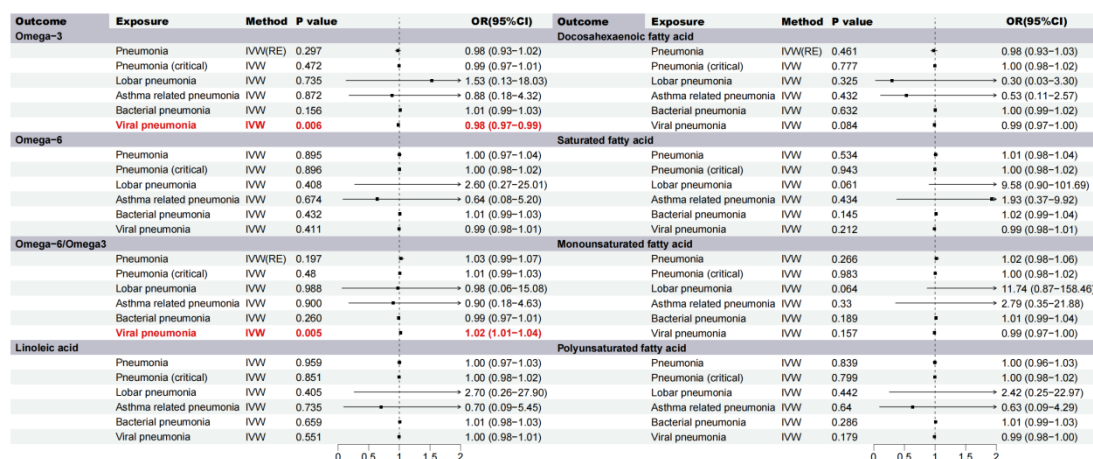
To verify robustness, we assessed heterogeneity and pleiotropy across instruments. For DHA, Cochran's Q test indicated no heterogeneity ( $P>0.05$ ), whereas omega-6 and LA instruments displayed modest heterogeneity ( $P<0.05$ ) but no evidence of horizontal pleiotropy by MR-Egger intercept (Table 3). All instruments demonstrated adequate strength ( $F>30$ ; Supplementary Tables 8-10). Applying a random effects IVW model for heterogeneous traits did not materially alter effect estimates ( $P<0.05$ ), and MR-PRESSO detected no additional outliers. Symmetrical funnel plots (Supplementary Figure 3) and stable leave-one-out analyses (Supplementary Figure 4) further support the reliability of these causal inferences (Supplementary Table 11).

**Table (3): Sensitivity analysis of causal relationship between plasma fatty acid and pneumonia subtypes.**

| Exposure                    | Outcome             | Heterogeneity         |                  |                  | Pleiotropy  |         |
|-----------------------------|---------------------|-----------------------|------------------|------------------|-------------|---------|
|                             |                     | MR-Egger Statistics Q | MR-Egger P-Value | IVW Statistics Q | IVW P-Value | P-Value |
| Docosahexaenoic acid levels | Pneumonia           | 11.74                 | 0.698            | 14.78            | 0.541       | 0.102   |
| Omega-6 fatty acid levels   | Critical pneumonia  | 62.52                 | 0.022            | 63.00            | 0.025       | 0.573   |
| Linoleic acid levels        | Bacterial pneumonia | 48.62                 | 0.030            | 49.33            | 0.034       | 0.499   |

### Reverse causality between fatty acids and pneumonia

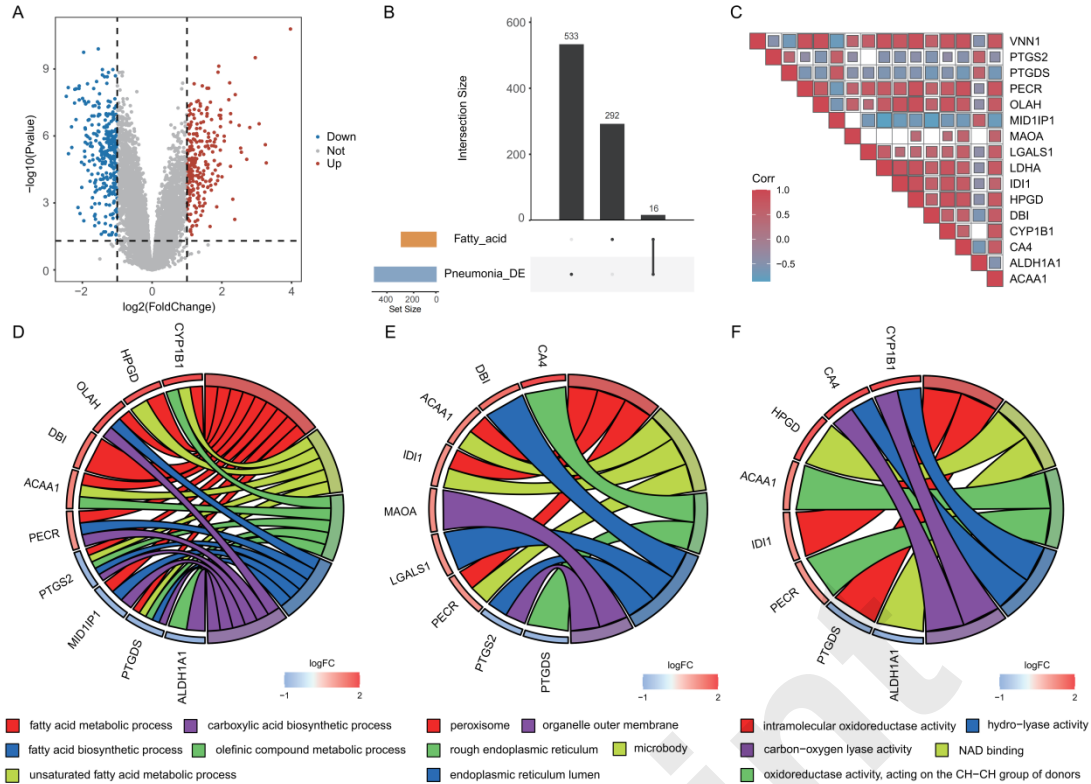
In reverse MR, we tested whether genetic liability to pneumonia subtypes influences fatty acid levels. Notably, higher genetic risk of viral pneumonia was causally linked to marginally lower omega-3 levels (OR=0.98, 95% CI: 0.97-0.99,  $P=0.006$ ) and to an elevated omega-6/omega-3 ratio (OR=1.02, 95% CI: 1.01-1.04,  $P=0.005$ ) (Fig. 4). A random effects IVW framework accounted for heterogeneity ( $P<0.05$ ), and sensitivity plots (Supplementary Figures. 5-8) confirmed absence of bias or influential SNPs.



**Fig. (4): Reverse Mendelian randomization estimates of Pneumonia on Plasma fatty acid.**

## Identification of Differential Genes Associated with Fatty Acid Metabolism

Analyzing GSE40012, we identified 549 genes differentially expressed between pneumonia patients and controls ( $|\log_2 FC| > 1$ ,  $P < 0.05$ ), including 251 up-regulated and 298 down-regulated (Fig. 5A). Intersection with our curated set of 308 fatty acid metabolism genes yielded 16 overlapping targets (Fig. 5B). Correlation analysis revealed strong positive co-expression among most of these 16 genes (Fig. 5C). GO enrichment highlighted their roles in fatty acid and unsaturated fatty acid metabolism, carboxylic acid biosynthesis, and localization to peroxisomes, outer mitochondrial membranes, and rough endoplasmic reticulum (Fig. 5D-F).



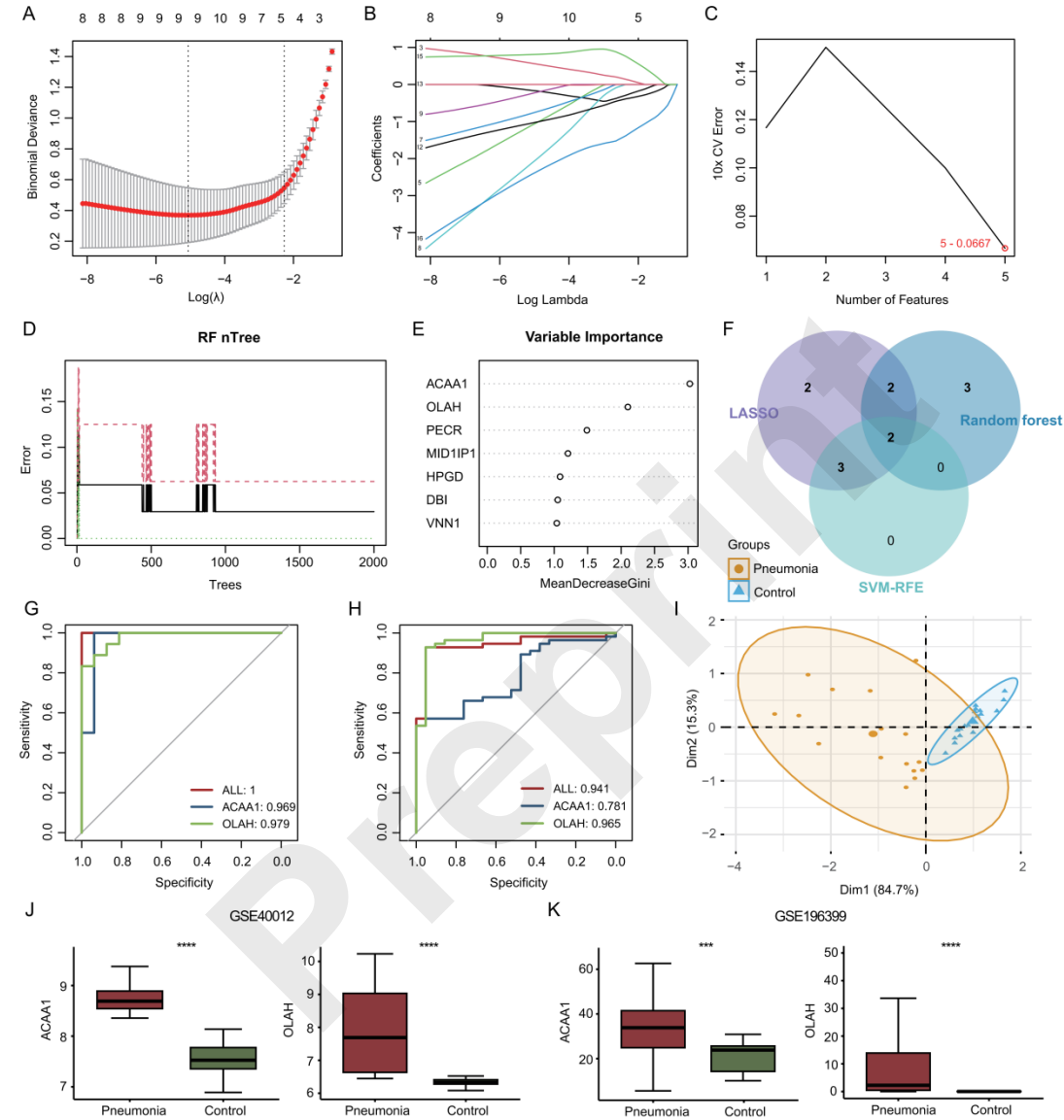
**Fig. (5): Differential Gene Expression and Functional Enrichment Analysis of Fatty Acid Metabolism-Related Genes in Pneumonia.** (A) Volcano plot depicting differential gene expression in pneumonia. (B) Intersection of fatty acid metabolism-related genes and pneumonia differential genes. (C) Correlation heatmap of differential genes associated with fatty acid metabolism. (D-F) GO enrichment map of 16 fatty acid metabolism differential genes enriched in BP, CC and MF.

BP: Biological Process, CC: Cellular Component, CC: Cellular Component

### Identification of key fatty acid metabolism genes in pneumonia

In order to exclude unimportant genes and identify key genes associated with pneumonia, we employed three machine learning algorithms for gene selection. Starting with 16 fatty acid metabolism-related differential genes, we applied Lasso regression, which selected 9 genes (Fig. 6A-B), SVM-RFE selected 5 genes (Fig. 6C), and the random forest algorithm identified 7 genes (Fig. 6D-E). Only ACAA1 and OLAH were consistently selected by all three methods (Fig. 6F). In the training cohort (GSE40012), ROC AUCs were 0.969 for ACAA1 and 0.979 for OLAH (Fig. 6G); in

validation (GSE196399), AUCs were 0.781 and 0.965, respectively (Fig. 6H). PCA based on these two genes demonstrated clear separation of cases versus controls (Fig. 6I), and expression differences were highly significant in both datasets (Fig. 6J-K).



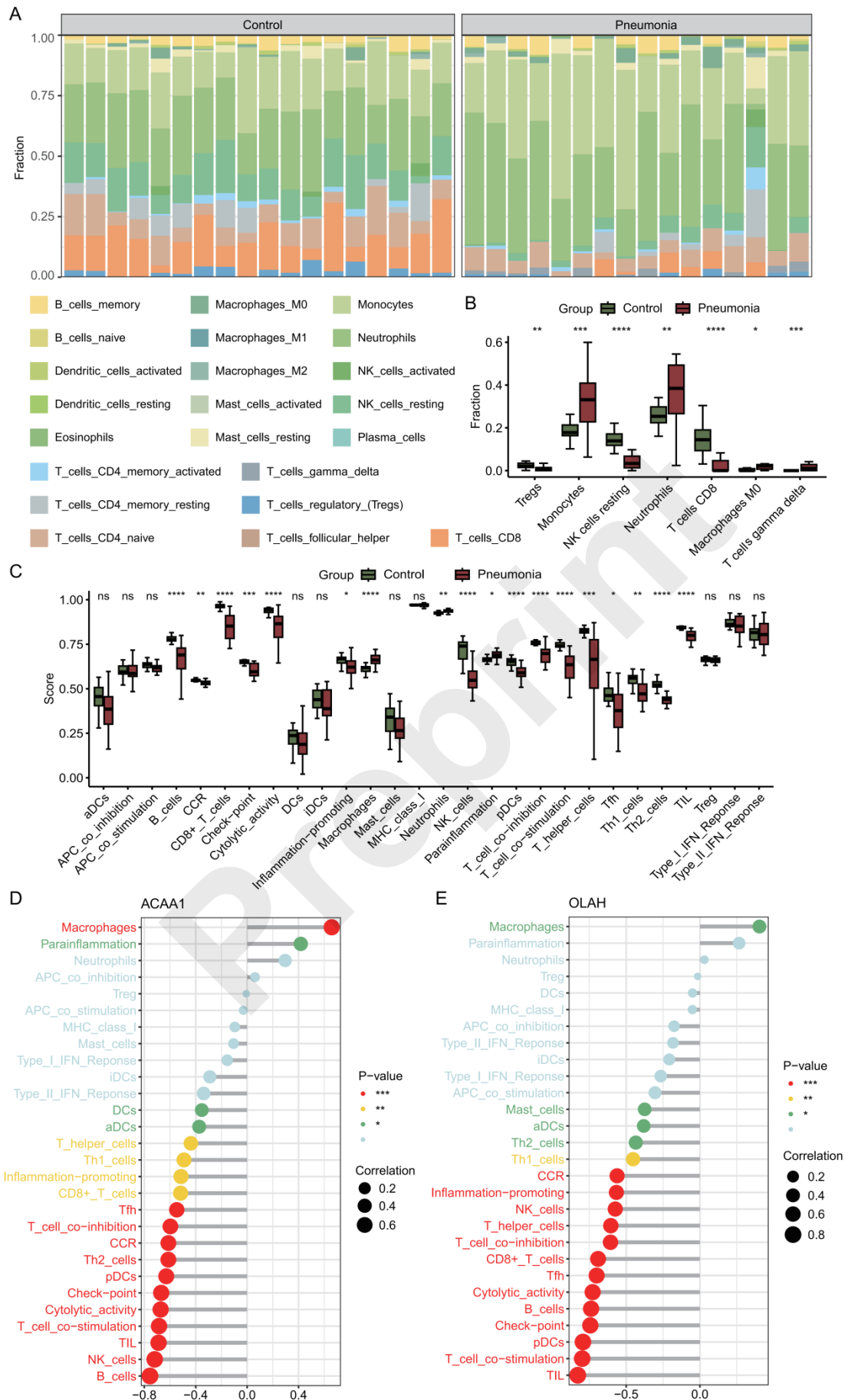
**Fig. (6): Identification and validation of pneumonia diagnostic markers.** (A) Lasso regression and (B) 10 fold cross validation diagram. (C) Error rate curve based on SVM-RFE algorithm with 10 fold cross validation. (D) The relationship between random forest error rate and the number of classification trees and (E) gene importance. (F) Genes shared by three machine learning models. (G) ROC curves of each signature gene in the training set (GSE40012) and (H) validation set (GSE196399) of the



diagnostic markers. (I) PCA plot of training set (GSE40012). (J) Expression of signature genes in the training set (GSE40012) and (K) validation set (GSE196399) in pneumonia and normal samples.

### **Immune Cell Infiltration and Immune-Related Function in Pneumonia**

To characterize immune microenvironment alterations in pneumonia, we first applied CIBERSORT to quantify immune cell proportions in pneumonia versus healthy lung tissues (Fig. 7A-B). Compared to controls, pneumonia tissues showed significant increases in neutrophils, monocytes, M0 macrophages, and gamma-delta T cells, accompanied by reduced regulatory T cells (Tregs), resting NK cells, and CD8+ T cells. Next, to explore broader functional implications, we performed single-sample gene set enrichment analysis (ssGSEA), which revealed distinct immune-related functional signatures and infiltration patterns between pneumonia patients and controls (Fig. 7C). Notably, ssGSEA further demonstrated that elevated expression of ACAA1 and OLAH correlated positively with macrophage infiltration but inversely with cytotoxic immune functions (Fig. 7D-E), suggesting these fatty acid metabolism-related genes may drive macrophage polarization while suppressing adaptive immunity during pneumonia pathogenesis.



**Fig. (7): Analysis of immune cell infiltration in patients with pneumonia and normal controls.** (A) Distribution and (B) differences in levels of immune cell infiltration between normal samples and pneumonia patients. (C) Differences in immune cell infiltration between normal and pneumonia patients using ssGSEA. (D) Association between the ACAA1, (E) OLAH and immune cell infiltration levels. The circle size indicates the correlation strength, with larger circles representing stronger correlations. The color reflects the p-value significance: red for  $p < 0.001$ , yellow for  $p < 0.01$ , and green for  $p < 0.05$ .

## Discussion

In this study, we investigated the causal relationships between plasma fatty acids and pneumonia, with a specific emphasis on identifying key genes involved in fatty acid metabolism and their potential roles in disease pathogenesis. Our MR analysis revealed that LA and DHA are causally associated with a reduced risk of pneumonia, suggesting their protective effects. These findings support and extend earlier observational studies, which have reported anti-inflammatory and immunomodulatory effects of omega-3 and omega-6 fatty acids in various inflammatory conditions, including pneumonia [25, 26]. For instance, a large prospective cohort study demonstrated that every 1-gram increase in LA intake was associated with a 4% reduction in pneumonia risk [27], which is consistent with our MR-derived protective estimate for LA.

Our study further confirms the anti-inflammatory role of omega-3 fatty acids, particularly DHA, which are known to reduce the severity of inflammatory diseases by modulating immune responses [14]. While previous observational studies have suggested a beneficial association, our MR analysis strengthens the causal interpretation, overcoming potential confounding and reverse causation. On the other hand, omega-6 fatty acids are generally considered pro-inflammatory and have been associated with adverse outcomes in certain pneumonia subtypes [28, 29]. Our results support this view, as genetically predicted higher omega-6 levels were associated with

increased risk of severe pneumonia, thereby aligning with earlier evidence and reinforcing the dualistic role of fatty acids in inflammation.

In animal models, omega-3 supplementation has been shown to mitigate pneumonia severity by suppressing microglial activation and inflammatory cytokine release through HMGB1 and TLR4/NF- $\kappa$ B signaling pathways [14, 30]. Furthermore, omega-3 and its derivatives have been shown to alleviate intestinal inflammation and modulate systemic immune responses in diseases such as ulcerative colitis [31, 32]. Short-chain fatty acids, another important class of fatty acids, exert anti-inflammatory effects by lowering colonic pH and limiting the growth of pathogenic bacteria [33]. These consistent findings across different disease models underscore the importance of fatty acid metabolism in modulating immune function and inflammatory responses.

We further explored potential molecular mediators linking fatty acids to pneumonia by applying machine learning algorithms. Through this approach, we identified ACAA1 and OLAH as key genes involved in fatty acid metabolism. ACAA1 plays a crucial role in mitochondrial  $\beta$ -oxidation, while OLAH is involved in fatty acid biosynthesis and homeostasis [34, 35]. Previous studies have linked ACAA1 variants with asthma and other inflammatory airway diseases, suggesting its involvement in immune regulation [36]. The identification of these genes complements and expands previous research by offering specific molecular targets through which fatty acids may exert their protective effects in pneumonia.

Infiltration analysis of immune cells revealed significant differences between pneumonia patients and healthy individuals. In line with previous reports, healthy controls exhibited higher infiltration of Tregs, resting NK cells, and CD8<sup>+</sup> T cells, whereas pneumonia patients showed increased infiltration of neutrophils, monocytes, M0 macrophages, and gamma-delta T cells. These findings are consistent with the established role of neutrophils and macrophages in pneumonia pathogenesis [37, 38]. In *Streptococcus pneumoniae*-induced pneumonia, for example, neutrophil accumulation is a hallmark of the early immune response, contributing to bacterial clearance but also to lung tissue damage and systemic spread [39, 40]. Macrophages act as first responders to pathogens and contribute to both host defense and

inflammation by releasing cytokines and orchestrating immune responses [41-44]. The elevated levels of gamma-delta T cells in pneumonia samples also suggest a role in early immune activation, consistent with their known function in bridging innate and adaptive immunity [45].

In addition to lipid mediators, various acute-phase plasma proteins, including C-reactive protein, procalcitonin, surfactant proteins, and complement components, are key players in the pathophysiology of pneumonia [46-49]. These proteins are closely correlated with disease severity and outcomes [50]. Notably, their interplay with fatty-acid-driven cytokine modulation implies that integrating lipid and protein biomarkers into combined panels could significantly enhance the accuracy of pneumonia diagnosis and improve prognostication[51, 52].

While our MR framework reduces confounding and reverse causation, it is limited by reliance on peripheral blood-derived GWAS and transcriptomic data, which lack direct validation in lung tissue. Future studies should combine clinical phenotyping with analyses of lung biopsy or bronchoalveolar lavage specimens to validate these findings. Additionally, functional studies of ACAA1 and OLAH in animal models will be critical to translate these genetic insights into targeted therapies.

## Conclusion

In summary, our bidirectional MR study establishes that genetically higher levels of LA and DHA reduce pneumonia risk, while elevated omega-6 increases susceptibility to severe disease. Integrating machine learning driven transcriptomic analysis, we pinpoint ACAA1 and OLAH as novel biomarkers linking fatty acid metabolism to pneumonia pathogenesis. Immune cell deconvolution further reveals a shift toward neutrophil and macrophage driven inflammation alongside reduced regulatory and cytotoxic T-cell subsets in patients. Together, these findings deepen our understanding of lipid-immune crosstalk in pneumonia and highlight promising targets for future diagnostic and therapeutic strategies.

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389   Supplementary Tables 1-3: Univariable MR estimates for plasma fatty acids across  
390   pneumonia phenotypes

391   Supplementary Tables 4-6: Genetic instruments used in MR analyses

392   Supplementary Table 7: Heterogeneity and pleiotropy test results (Cochran's Q, MR-  
393   Egger intercept)

394   Supplementary Tables 8-9: Instrument strength (F-statistics) and outlier removal details

395   Supplementary Table 10: Scatter plots and forest plots summary for key SNPs

396   Supplementary Table 11: Leave-one-out sensitivity analyses

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Preprint

**Declaration**

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Availability of data and materials**

The data and materials in the current study are available from the corresponding author on reasonable request.

**Competing interests**

The authors declare that they have no potential conflicts of interest.

**Funding**

None.

**Authors' contributions**

XBC, XYW, PAM and ZJG contributed to the study design. XBC conducted the literature search. PAM and ZJG acquired the data. XBC wrote the article and performed data analysis. XMW revised the article and gave the final approval of the version to be submitted. All authors read and approved the final manuscript.

**Acknowledgements**

Not applicable.

## Reference

- [1] A. Torres, C. Cilloniz, M.S. Niederman, R. Menéndez, J.D. Chalmers, R.G. Wunderink, et al., Pneumonia, *Nature reviews. Disease primers*. 2021;7(1): 25.
- [2] S. Neill, N. Dean, Aspiration pneumonia and pneumonitis: a spectrum of infectious/noninfectious diseases affecting the lung, *Current opinion in infectious diseases*. 2019;32(2): 152-157.
- [3] H. Zhang, D. Zhan, D. Chen, W. Huang, M. Yu, Q. Li, et al., Next-generation sequencing diagnosis of severe pneumonia from fulminant psittacosis with multiple organ failure: a case report and literature review, *Annals of translational medicine*. 2020;8(6): 401.
- [4] S.R. Savla, K.S. Prabhavalkar, L.K. Bhatt, Cytokine storm associated coagulation complications in COVID-19 patients: Pathogenesis and Management, *Expert review of anti-infective therapy*. 2021;19(11): 1397-1413.
- [5] N.Y. Kusumawardhani, I.C.S. Putra, W. Kamarullah, R. Afrianti, M. Pramudyo, M. Iqbal, et al., Cardiovascular Disease in Post-Acute COVID-19 Syndrome: A Comprehensive Review of Pathophysiology and Diagnosis Approach, *Reviews in cardiovascular medicine*. 2023;24(1): 28.
- [6] C. Cilloniz, C.S. Dela Cruz, G. Dy-Agra, R.S. Pagcatipunan, Jr., World Pneumonia Day 2024: Fighting Pneumonia and Antimicrobial Resistance, *American journal of respiratory and critical care medicine*. 2024;210(11): 1283-1285.
- [7] C. Garcia, C.J. Andersen, C.N. Blesso, The Role of Lipids in the Regulation of Immune Responses, *Nutrients*. 2023;15(18).
- [8] R.M. Torres, J. Cyster, Lipid mediators in the regulation of innate and adaptive immunity, *Immunological reviews*. 2023;317(1): 4-7.
- [9] J.F.J. Bogie, M. Haidar, G. Kooij, J.J.A. Hendriks, Fatty acid metabolism in the progression and resolution of CNS disorders, *Advanced drug delivery reviews*. 2020;159: 198-213.
- [10] S. Kotlyarov, A. Kotlyarova, Anti-Inflammatory Function of Fatty Acids and Involvement of Their Metabolites in the Resolution of Inflammation in Chronic Obstructive Pulmonary Disease, *International journal of molecular sciences*.



2021;22(23).

[11] B. Guo, M. Xue, T. Zhang, H. Gan, R. Lin, M. Liu, et al., Correlation between immune-related Tryptophan-Kynurenine pathway and severity of severe pneumonia and inflammation-related polyunsaturated fatty acids, *Immunity, inflammation and disease*. 2023;11(11): e1088.

[12] A. Krupa, I. Kowalska, The Kynurenine Pathway-New Linkage between Innate and Adaptive Immunity in Autoimmune Endocrinopathies, *International journal of molecular sciences*. 2021;22(18).

[13] P.J. Siska, J. Jiao, C. Matos, K. Singer, R.S. Berger, K. Dettmer, et al., Kynurenine induces T cell fat catabolism and has limited suppressive effects in vivo, *EBioMedicine*. 2021;74: 103734.

[14] C.A. Hinojosa, N. Gonzalez-Juarbe, M.M. Rahman, G. Fernandes, C.J. Orihuela, M.I. Restrepo, Omega-3 fatty acids in contrast to omega-6 protect against pneumococcal pneumonia, *Microbial pathogenesis*. 2020;141: 103979.

[15] H. Zhao, P.A. Dennerly, H. Yao, Metabolic reprogramming in the pathogenesis of chronic lung diseases, including BPD, COPD, and pulmonary fibrosis, *American journal of physiology. Lung cellular and molecular physiology*. 2018;314(4): L544-L554.

[16] C. Kocherlakota, B. Nagaraju, N. Arjun, A. Srinath, K.S.D. Kothapalli, J.T. Brenna, Inhalation of nebulized omega-3 fatty acids mitigate LPS-induced acute lung inflammation in rats: Implications for treatment of COPD and COVID-19, *Prostaglandins, leukotrienes, and essential fatty acids*. 2022;179: 102426.

[17] M. Hagihara, M. Yamashita, T. Ariyoshi, S. Eguchi, A. Minemura, D. Miura, et al., Clostridium butyricum-induced  $\omega$ -3 fatty acid 18-HEPE elicits anti-influenza virus pneumonia effects through interferon- $\lambda$  upregulation, *Cell reports*. 2022;41(11): 111755.

[18] E. Sanderson, M.M. Glymour, M.V. Holmes, H. Kang, J. Morrison, M.R. Munafò, et al., Mendelian randomization, *Nature reviews. Methods primers*. 2022;2.

[19] S. Burgess, D.S. Small, S.G. Thompson, A review of instrumental variable estimators for Mendelian randomization, *Statistical methods in medical research*.

2017;26(5): 2333-2355.

[20] F.P. Hartwig, N.M. Davies, G. Hemani, G. Davey Smith, Two-sample Mendelian randomization: avoiding the downsides of a powerful, widely applicable but potentially fallible technique, *International journal of epidemiology*. 2016;45(6): 1717-1726.

[21] M. Verbanck, C.Y. Chen, B. Neale, R. Do, Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases, *Nature genetics*. 2018;50(5): 693-698.

[22] M.E. Ritchie, B. Phipson, D. Wu, Y. Hu, C.W. Law, W. Shi, et al., limma powers differential expression analyses for RNA-sequencing and microarray studies, *Nucleic acids research*. 2015;43(7): e47.

[23] T. Wu, E. Hu, S. Xu, M. Chen, P. Guo, Z. Dai, et al., clusterProfiler 4.0: A universal enrichment tool for interpreting omics data, *Innovation (Cambridge (Mass.))*. 2021;2(3): 100141.

[24] D. Zeng, Z. Ye, R. Shen, G. Yu, J. Wu, Y. Xiong, et al., IOBR: Multi-Omics Immuno-Oncology Biological Research to Decode Tumor Microenvironment and Signatures, *Frontiers in immunology*. 2021;12: 687975.

[25] U. Radzikowska, A.O. Rinaldi, Z. Çelebi Sözen, D. Karaguzel, M. Wojcik, K. Cypryk, et al., The Influence of Dietary Fatty Acids on Immune Responses, *Nutrients*. 2019;11(12).

[26] I. Djuricic, P.C. Calder, Beneficial Outcomes of Omega-6 and Omega-3 Polyunsaturated Fatty Acids on Human Health: An Update for 2021, *Nutrients*. 2021;13(7).

[27] A.T. Merchant, G.C. Curhan, E.B. Rimm, W.C. Willett, W.W. Fawzi, Intake of n-6 and n-3 fatty acids and fish and risk of community-acquired pneumonia in US men, *The American journal of clinical nutrition*. 2005;82(3): 668-74.

[28] R. Poggioli, K. Hirani, V.G. Jogani, C. Ricordi, Modulation of inflammation and immunity by omega-3 fatty acids: a possible role for prevention and to halt disease progression in autoimmune, viral, and age-related disorders, *European review for medical and pharmacological sciences*. 2023;27(15): 7380-7400.

[29] C. Lemoine, E. Brigham, H. Woo, A. Koch, C. Hanson, K. Romero, et al.,

Relationship between Omega-3 and Omega-6 Fatty Acid Intake and Chronic Obstructive Pulmonary Disease Morbidity, *Annals of the American Thoracic Society*. 2020;17(3): 378-381.

[30] X. Chen, S. Wu, C. Chen, B. Xie, Z. Fang, W. Hu, et al., Omega-3 polyunsaturated fatty acid supplementation attenuates microglial-induced inflammation by inhibiting the HMGB1/TLR4/NF- $\kappa$ B pathway following experimental traumatic brain injury, *Journal of neuroinflammation*. 2017;14(1): 143.

[31] L.T. Marton, R.A. Goulart, A.C.A. Carvalho, S.M. Barbalho, Omega Fatty Acids and Inflammatory Bowel Diseases: An Overview, *International journal of molecular sciences*. 2019;20(19).

[32] R. Reifen, A. Karlinsky, A.H. Stark, Z. Berkovich, A. Nyska,  $\alpha$ -Linolenic acid (ALA) is an anti-inflammatory agent in inflammatory bowel disease, *J Nutr Biochem*. 2015;26(12): 1632-40.

[33] Y. Shin, S. Han, J. Kwon, S. Ju, T.G. Choi, I. Kang, et al., Roles of Short-Chain Fatty Acids in Inflammatory Bowel Disease, *Nutrients*. 2023;15(20).

[34] X. Huang, Y. Li, P. Zhuang, X. Liu, Y. Zhang, P. Zhang, et al., Habitual Fish Oil Supplementation and Risk of Incident Inflammatory Bowel Diseases: A Prospective Population-Based Study, *Frontiers in nutrition*. 2022;9: 905162.

[35] R. Palanki, H. Yamagata, M.J. Mitchell, OLAH connects fatty acid metabolism to the severity of respiratory viral disease, *Cell*. 2024;187(17): 4549-4551.

[36] J.E. Sordillo, S. Sharma, A. Poon, J. Lasky-Su, K. Belanger, D.K. Milton, et al., Effects of endotoxin exposure on childhood asthma risk are modified by a genetic polymorphism in ACAA1, *BMC medical genetics*. 2011;12: 158.

[37] R.D. Pechous, With Friends Like These: The Complex Role of Neutrophils in the Progression of Severe Pneumonia, *Frontiers in cellular and infection microbiology*. 2017;7: 160.

[38] A. Zhou, K. Chen, Y. Gao, X. Zhou, Z. Tian, W. Chen, et al., Bioengineered Neutrophil Extinguisher Targets Cascade Immune Pathways of Macrophages for Alleviating Cytokine Storm in Pneumonia, *ACS nano*. 2023;17(17): 16461-16477.

[39] H. Domon, Y. Terao, The Role of Neutrophils and Neutrophil Elastase in

Pneumococcal Pneumonia, *Frontiers in cellular and infection microbiology*. 2021;11: 615959.

[40] C.S. Palmer, J.M. Kimmey, Neutrophil Recruitment in Pneumococcal Pneumonia, *Frontiers in cellular and infection microbiology*. 2022;12: 894644.

[41] A. Guillon, E.I. Arafa, K.A. Barker, A.C. Belkina, I. Martin, A.T. Shenoy, et al., Pneumonia recovery reprograms the alveolar macrophage pool, *JCI insight*. 2020;5(4).

[42] M. Miyagi, K. Uchida, S. Takano, H. Fujimaki, J. Aikawa, H. Sekiguchi, et al., Macrophage-derived inflammatory cytokines regulate growth factors and pain-related molecules in mice with intervertebral disc injury, *Journal of orthopaedic research : official publication of the Orthopaedic Research Society*. 2018.

[43] S. Turner, M.A. Khan, D. Putrino, A. Woodcock, D.B. Kell, E. Pretorius, Long COVID: pathophysiological factors and abnormalities of coagulation, *Trends in endocrinology and metabolism: TEM*. 2023;34(6): 321-344.

[44] H. Wei, H. Zhao, R. Li, F. Yang, Y. Wu, Rhinovirus impairs the immune response of alveolar macrophages to facilitate Streptococcus pneumonia infection, *Pathogens and disease*. 2020;78(3).

[45] Y. Hu, Q. Hu, Y. Li, L. Lu, Z. Xiang, Z. Yin, et al.,  $\gamma\delta$  T cells: origin and fate, subsets, diseases and immunotherapy, *Signal transduction and targeted therapy*. 2023;8(1): 434.

[46] T. Güleç, S. Yılmaz, R. Ak, A.C. Tatlıparmak, Ö. Karcıoğlu, Can we recognize severe community-acquired pneumonia without pneumonia severity index? Use of modified qSOFA with procalcitonin, *Heliyon*. 2023;9(9): e19937.

[47] N.Y. Saleh, R.A.L. Ibrahim, A.A.H. Saleh, S.E.S. Soliman, A.A.S. Mahmoud, Surfactant protein D: a predictor for severity of community-acquired pneumonia in children, *Pediatric research*. 2022;91(3): 665-671.

[48] S. Bhattacharya, C. Munshi, Biological significance of C-reactive protein, the ancient acute phase functionary, *Frontiers in immunology*. 2023;14: 1238411.

[49] Z. Xu, X.F. Hou, C.M. Feng, L. Zheng, D.X. Xu, H. Zhao, et al., The association between serum complement C3a and severity in patients with community-acquired pneumonia, *Frontiers in immunology*. 2023;14: 1034233.

- [50] A. Zinellu, A.A. Mangoni, Serum Complement C3 and C4 and COVID-19 Severity and Mortality: A Systematic Review and Meta-Analysis With Meta-Regression, *Frontiers in immunology*. 2021;12: 696085.
- [51] S. Ozbay, M. Ayan, O. Ozsoy, C. Akman, O. Karcioglu, Diagnostic and Prognostic Roles of Procalcitonin and Other Tools in Community-Acquired Pneumonia: A Narrative Review, *Diagnostics (Basel, Switzerland)*. 2023;13(11).
- [52] A. Elmore, A. Almuntashiri, X. Wang, S. Almuntashiri, D. Zhang, Circulating Surfactant Protein D: A Biomarker for Acute Lung Injury?, *Biomedicines*. 2023;11(9).

