

Protective effects of total glucosides of paeony and the underlying mechanisms in carbon tetrachloride-induced experimental liver injury

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Submitted: 16 September 2010

Accepted: 30 November 2010

Arch Med Sci 2011; 7, 4: 604-612

DOI: 10.5114/aoms.2011.24129

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Abstract

Introduction: We explored the protective effects of total glucosides of paeony (TGP) and the underlying mechanisms in carbon tetrachloride (CCl₄)-induced experimental liver injury in mice.

Material and methods: Chronic liver damage was induced by intraperitoneal injection of CCl₄ (0.5 µl/g) three times per week for 8 weeks. Mice also received 25, 50 or 100 mg/kg TGP. Liver sections were stained with haematoxylin/eosin. Serum amino transferases, lipid peroxidation and tumour necrosis factor-α (TNF-α) levels were determined using commercial assays. Quantitative real-time polymerase chain reaction was used to determine the changes in hepatic *TNF-α*, *COX-2*, *iNOS* and *HO-1* expression. Protein levels of nitric oxide synthase, cyclooxygenase-2, haem oxygenase-1 and cytochrome P450 2E1 were determined by western blotting.

Results: Histological results showed that TGP improved the CCl₄-induced changes in liver structure and alleviated lobular necrosis. The increases in serum protein and hepatic mRNA expression of TNF-α induced by CCl₄ treatment were suppressed by TGP. Total glucosides of paeony also attenuated the increase the expression in *iNOS* and *CYP2E1* but augmented the increase in *HO-1*. The mRNA and protein expression levels of inducible HO-1 increased significantly after CCl₄ treatment.

Conclusions: Total glucosides of paeony protects hepatocytes from oxidative damage induced by CCl₄. Total glucosides of paeony may achieve these effects by enhancing HO-1 expression and inhibiting the expression of proinflammatory mediators.

Key words: total glucosides of paeony, liver injury, carbon tetrachloride.

Introduction

Previous studies have shown that carbon tetrachloride (CCl₄) enhances oxidative stress and the recruitment of inflammatory cells and therefore leads to hepatic architectural and functional damage [1-7]. Carbon tetrachloride impairs hepatocytes directly by altering the permeability of the plasma, lysosomal, and mitochondrial membranes. A single dose of CCl₄ may lead to centrilobular necrosis and steatosis [8], while prolonged administration may induce liver fibrosis, cirrhosis, and even hepatocellular carcinoma (HCC). The lesions induced by carbon tetrachloride (CCl₄) replicate those seen in most cases of human liver diseases [9]. Therefore,

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the experimental model of CCl₄-induced hepatic injury has been used for many years to investigate the mechanisms involved in acute and chronic liver injury and to screen hepatoprotective drugs. Carbon tetrachloride-induced hepatic necrosis involves the bioactivation of a microsomal cytochrome P450-dependent mono-oxygenase system, causing the formation of trichloromethyl free radicals and reactive oxygen species (ROS) that initiate lipid peroxidation and protein oxidation, resulting in hepatocellular membrane damage [10]. This process is followed by the release of inflammatory mediators from the activated hepatic macrophages, which are thought to potentiate CCl₄-induced hepatic injury.

Paeonia lactiflora pall root, a Chinese traditional herbal medicine that has anti-inflammatory and antioxidative actions, has been used as a component of some hepatoprotective herb mixtures in China, Japan and Korea [11-13]. Total glucosides of paeony (TGP), extracted from *P. lactiflora*, contains paeoniflorin and other components such as hydroxyl-paeoniflorin, paeonin, albiflorin, and benzoylpaeoniflorin [14]. Paeoniflorin is the main active component of TGP, accounting for approximately 90% of the constituents of TGP. It has been shown that TGP has anti-inflammatory, antioxidative, hepatoprotective and immunoregulatory activities, without evident toxicity or side effects. Total glucosides of paeony has a long history of use in traditional Chinese medicine [15], particularly for the treatment of rheumatoid arthritis and systemic lupus erythematosus, and as a component of treatments for liver disease.

The present study was designed to evaluate whether TGP has any beneficial effect on liver histopathology or liver function in an experimental model of CCl₄-induced chronic liver injury. We also investigated the underlying mechanisms based on

the therapeutic role of TGP on the amelioration of inflammation and oxidative stress.

Material and methods

Drugs and animals

Total glucosides of paeony capsules (Ningbo Liwah Pharmaceutical CO., LTD, Zhejiang Province, China) were dissolved in saline to a concentration of 1.2 mg/ml. Sodium carboxymethyl cellulose (0.5 g; CMC-Na; Sigma, St. Louis, MO, USA) was dissolved in 100 ml of distilled water to provide a 0.5% (w/v) CMC-Na suspension. Biphenyldicarboxylate (BDP; Dezhou Deyao Pharmaceutical CO., LTD, Shandong Province, China) was dissolved in the 0.5% CMC-Na suspension to a concentration of 5.0 mg/ml. Figure 1 shows the chemical structures of paeoniflorin, the main component of TGP, and BDP.

Male C57BL/6 mice weighing 25-30 g were obtained from the Institute of Laboratory Animal Sciences, Chinese Academy of Medical Sciences & Peking Union Medical College (Beijing, China). The research protocol was in accordance with the principles approved by the animal ethics committee of Chinese People's Liberation Army General Hospital. Animals were housed at 23 ± 2°C with a 12-h light/dark cycle and were allowed free access to standard laboratory chow and tap water. The mice were randomly divided into 6 groups: control, CCl₄, TGP25, TGP50, TGP100 and BDP100 groups. The control group was intraperitoneally administered with the appropriate vehicles. The experimental groups were intraperitoneally injected with CCl₄ (0.5 ml/kg, 1 : 3 dilution in corn oil; Sigma) 3 times per week for 8 weeks. The mice in groups TGP25, TGP50 and TGP100 were given 25 mg/kg, 50 mg/kg and 100 mg/kg TGP per day, respectively, by gavage, twice daily for 12 weeks. The mice in the

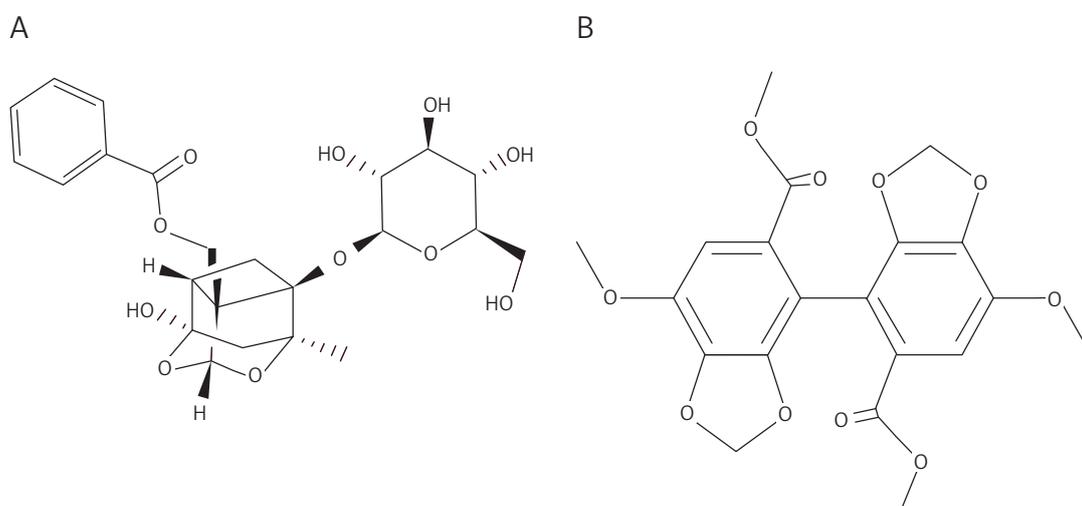


Figure 1. Chemical structures of paeoniflorin (A) and biphenyldicarboxylate (BDP) (B)
BDP – biphenyldicarboxylate

BDP100 group were given BDP, a positive hepato-protective control, at 100 mg/kg per day, once daily by gavage for 12 weeks. In the TGP and BDP groups, CCl₄ administration was started in week 5 of administration and was continued to week 12 (i.e., for 8 weeks). At the end of the treatment period, the mice were fasted overnight and were sacrificed. Blood was collected from the abdominal aorta and the liver was removed and stored at -80°C for later analysis.

Mouse hepatic lipid peroxidation and serum assays

Hepatic malondialdehyde (MDA) was measured as previously described [16-18]. In brief, 25 mg of liver tissue was added to 250 µl of radioimmuno-precipitation assay (RIPA) buffer containing protease inhibitors. This mixture was sonicated for 15 s at 40 V over ice and centrifuged at 1600 × g for 10 min at 4°C. The supernatant was used for analysis. MDA was quantified using the thiobarbituric acid reaction as described by Ohkawa [16, 17], and measured using a thiobarbituric acid reactive substances (TBARS) assay (Cayman Chemical Co. Inc., MI, USA). Superoxide dismutase (SOD), nitric oxide synthase (iNOS), catalase (CAT) and glutathione peroxidase (GSH) were also quantified using appropriate assays (Cayman Chemical Co. Inc.) [16-18].

Serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) activities were determined using standard spectrophotometric procedures. Serum tumour necrosis factor α (TNF-α) was measured using an enzyme-linked immunosorbent assay (R&D Systems, Minneapolis, MN, USA) according to the manufacturer's protocol.

Histological and immunohistochemical analyses

The liver tissues were immediately fixed in 4% paraformaldehyde, dehydrated, embedded in paraffin and sectioned. Formalin-fixed, paraffin-embedded sections were cut and mounted on glass slides. Sections (5 µm thick) were deparaffinised in xylene and stained with haematoxylin and eosin (H&E) as described by Mihailovic *et al.* [19]. The immunohistochemical evaluation method was performed as described elsewhere [20]. Briefly, sections (5 µm thick) were cut from paraffin-embedded testis tissues and mounted on slides. After deparaffinization and rehydration, they were placed in a 10-mM citrate buffer (pH 6.0), followed by heating in a microwave oven for antigen retrieval. For this, three periods of 5 min each were used, after which the sections were treated with 3% H₂O₂ in PBS (pH 7.6) for 20 min. The sections were then incubated overnight at 4°C in PBS

containing 3% BSA and antibodies. Antibodies against TNF-α and nuclear factor κ light chain enhancer in B cells p65 (NF-κB p65) (Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA) were used at 1 : 100 and 1 : 200 dilution, respectively. The bound primary antibody was detected using biotinylated rabbit anti-mouse antibody (Zhongshan Golden Bridge, Beijing, China), and visualised using 3',3'-diaminobenzidine tetrahydrochloride. Sections were slightly counterstained with Mayer's haematoxylin and mounted.

Real-time reverse transcription-polymerase chain reaction (qRT-PCR)

Total RNA was extracted from the livers, isolated and purified with TRIzol reagent (Invitrogen, Carlsbad, CA, USA) and NucleoSpin® RNA clean-up kit (Macherey-Nagel, Duren, Germany). Messenger RNA (mRNA) analysis was carried out by qRT-PCR using LightCycler technology (Roche Diagnostics, Indianapolis, IN, USA) for continuous fluorescence detection. Complementary DNA (cDNA) was prepared for qPCR analyses using the M-MLV reverse transcriptase enzyme (Invitrogen-Gibco, Carlsbad, CA, USA) and random hexamers as primers, as described [21-23]. The primers for mouse *TNF-α*, inducible nitric oxide synthase (*iNOS*), cyclooxygenase-2 (*COX-2*), and haem oxygenase-1 (*HO-1*) mRNA were used as previously reported [21-23]. The following sense and anti-sense primers were used for amplification: *TNF-α*, 5'-CTACTCCCAGGTTCTCTCAA-3' and 5'-GCAGAGAGGAGGTTGACTTTC-3' [21]; *iNOS*, 5'-CAGCTGGGCTGTACAAACCTT-3' and 5'-CATTGG-AAGTGAAGCGTTTCG-3' [21]; *COX-2*, 5'-TTCAATGA-GATTGTGGAAAAAT-3' and 5'-AGATCATCTCTGCCTGA-GTATCTT-3' [22]; *HO-1*, 5'-CGCCTTCCTGCTCAACATT-3' and 5'-TGTGTTCTCTGTGTCAGCATCAC-3' [23], respectively. The reactions were performed using a LightCycler FastStart DNA Master SYBR Green I Kit (Roche Applied Science) according to the manufacturer's instructions. The PCR reaction conditions and the annealing temperature were as previously described [21-23]. β-Actin was used as a reference control gene to normalize the expression value of each gene [24]. The experiment was performed in triplicate for each gene, and the average expression value was computed for subsequent analysis. The results were compared using the 2^{-ΔΔCT} method [25] and are expressed as log₂ means ± SD.

Western blotting

Liver whole protein was extracted and used for western blotting, which was performed as previously described [25]. Appropriate amounts of the protein extracts were fractionated by electrophoresis in 6-12% SDS-polyacrylamide gels

and transferred to nylon membranes. The nylon membranes containing the transferred proteins were pretreated with 1.0% non-fat dried milk in 50 mM Tris (pH 8.0) and then incubated overnight with primary mouse monoclonal antibodies (Santa Cruz Biotechnology Inc.) against iNOS, COX-2, HO-1 and cytochrome P450 2E1 (CYP2E1), at dilutions 1 : 300, 1 : 200, 1 : 500 and 1 : 400, respectively. The membranes were washed and incubated with anti-mouse secondary antibodies (Zhongshan Golden Bridge Biotechnology, Beijing, China) and immunoreactivity was visualised by exposing X-ray film to blots incubated with ECL reagent (Amersham Biosciences, Piscataway, NJ, USA). Results were quantified using Image-Pro Plus software version 6.0 (Media Cybernetics Incorporated, Silver Spring, MD, USA).

Statistical analysis

Each experiment was performed at least 3 times. Data are means \pm standard deviation. Statistical analyses were performed by SPSS software (version 13.0, SPSS Inc., Chicago, IL, USA). One-way ANOVA with post hoc analysis using Bonferroni's method or Kruskal-Wallis H test on ranks was used for analysing results from different groups. Value of $p < 0.05$ was considered statistically significant.

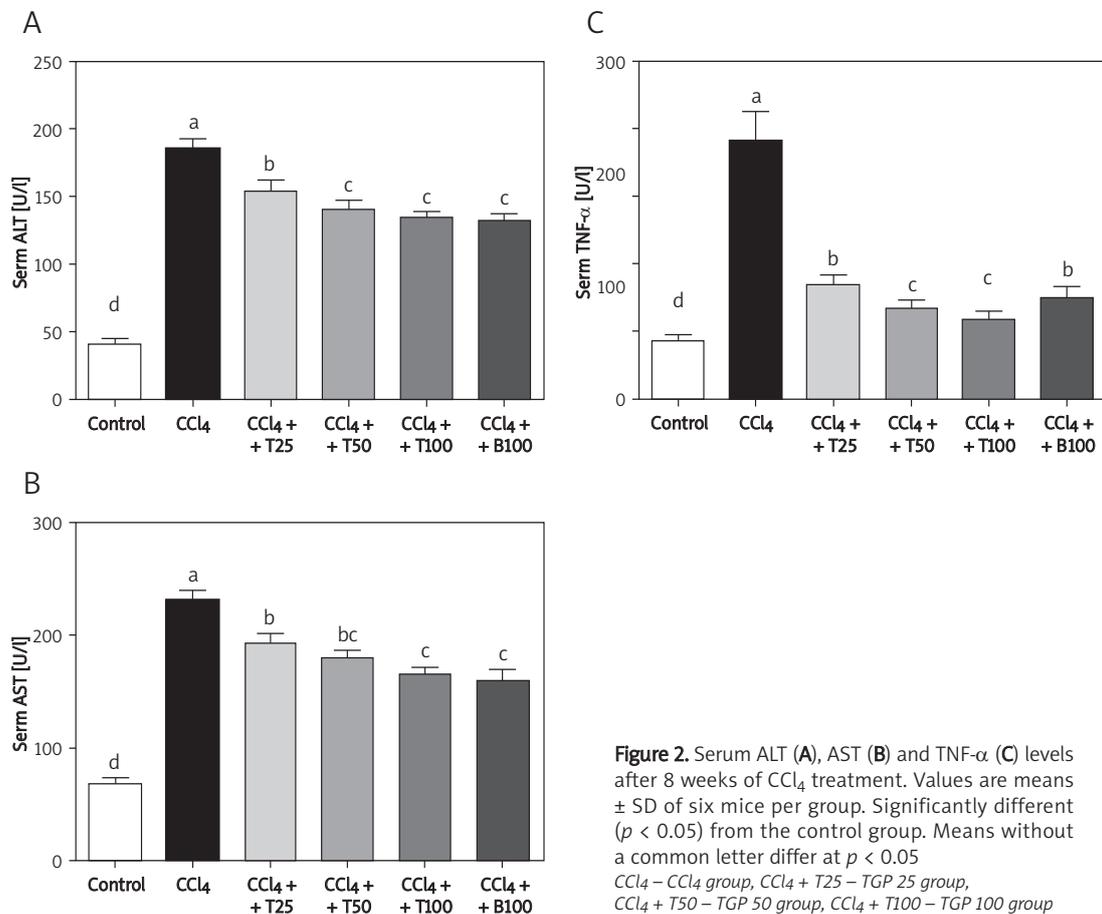
Results

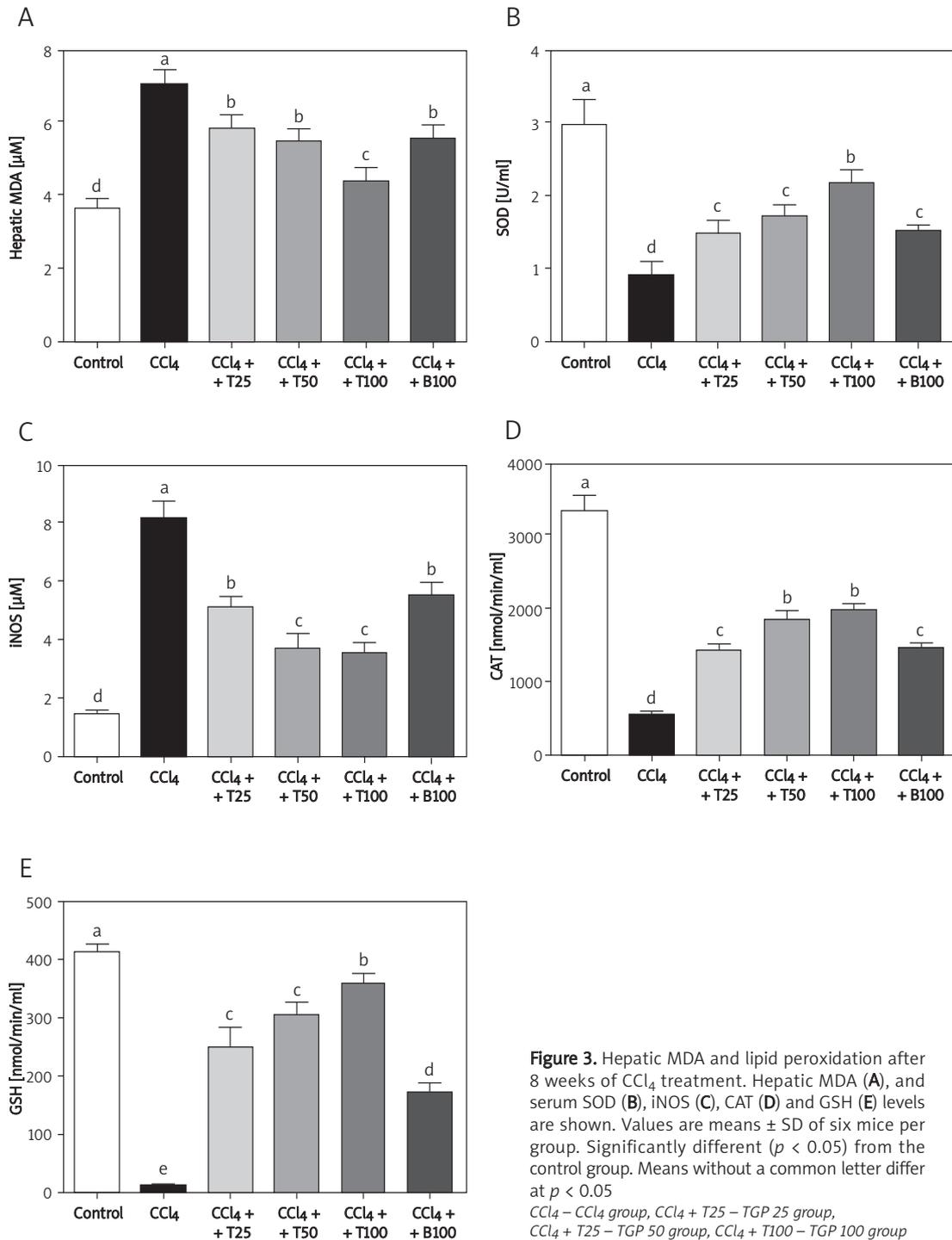
Serum TNF- α level and aminotransferase activities

The serum TNF- α level was 8.5 ± 2.2 pg/ml in the control group. In contrast, the serum TNF- α level was increased by almost 4-fold by CCl₄, which was significantly attenuated by TGP. The serum ALT and AST activities in the control group were 40.5 ± 10.9 U/l and 68.0 ± 12.9 U/l, respectively, and were markedly increased by CCl₄ treatment. These increases were inhibited by treatment with 25 mg/kg, 50 mg/kg and 100 mg/kg TGP (Figure 2).

Serum lipid peroxidation

The serum MDA level was significantly increased in the CCl₄-treated mice compared with the control group, and was attenuated by TGP or BDP. Similar to the MDA activities, the iNOS level was significantly increased in the CCl₄-treated mice, and was attenuated by all doses of TGP and by BDP. The serum SOD activity in the CCl₄-injected mice decreased to about 60% of the control value but this decrease was attenuated by all three doses of TGP. Changes in iNOS, CAT and GSH activities showed similar trends to that of SOD (Figure 3).





Histological and immunohistochemical analyses

Mice became lethargic and lean, and their fur became lustreless in the CCl₄-treated group by 8 weeks of treatment. Liver samples were stained with H&E to evaluate histopathological changes. The liver of mice in the control group showed a normal hepatic lobular architecture and cellular

structure. In contrast, the histopathological analysis of the liver of mice from the CCl₄-treated group showed extensive hepatocellular damage, with the presence of portal inflammation, marked neutrophilic infiltration, extensive fatty changes, centrilobular necrosis, Kupffer cell hyperplasia and hepatic fibrosis. These histopathological changes were ameliorated by daily administration of TGP or BDP for 12 weeks (Figure 4).

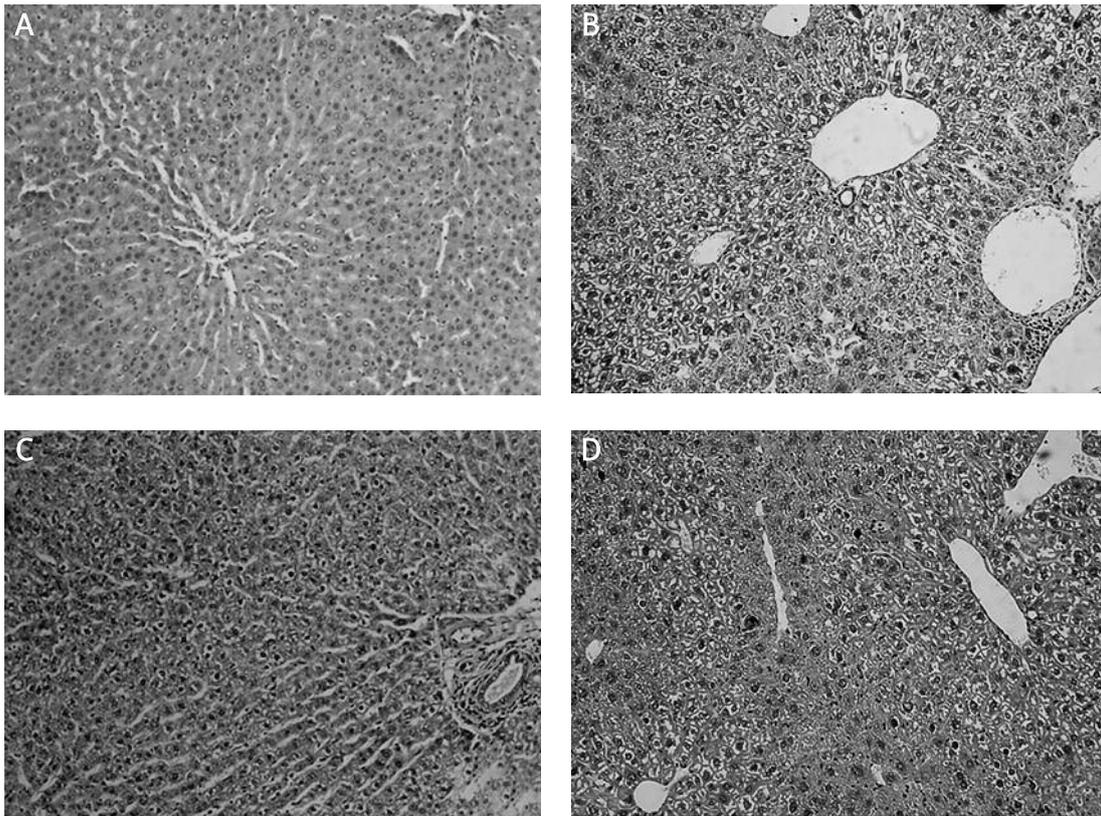


Figure 4. Effects of TGP on histological changes in the liver induced by CCl₄ treatment for 8 weeks. **A** – the control group shows normal lobular architecture and cell structure. **B** – the CCl₄-treated group shows extensive hepatocellular damage with the presence of portal inflammation, centrilobular necrosis, and Kupffer cell hyperplasia. **C** – mild portal inflammation, little hepatocellular necrosis and Kupffer cell hyperplasia are present in the 100 mg/kg TGP group. **D** – there were no marked improvements in CCl₄-induced liver injury in the 100 mg/kg BDP group. Bars, 100 μm

Immunohistochemistry was performed to determine the changes in intrahepatic TNF- α and NF- κ B p65 expression in the liver after CCl₄ inducement. We found that TGP significantly reduced the hepatic expression of both proteins compared with the CCl₄-treated mice. As shown in Figure 5, daily administration of 100 mg/kg TGP for 12 weeks reduced the expression of TNF- α and NF- κ B p65 in the liver and improved the histopathological changes and tissue fibrosis compared with mice treated with CCl₄ alone.

TNF- α , iNOS, COX-2 and HO-1 mRNA expression

Hepatic TNF- α , iNOS, COX-2, and HO-1 mRNA expression were quite low in the control group. However, the mRNA expression levels of these genes were greatly increased by CCl₄ administration. The increase in the expression of TNF- α and iNOS mRNA was suppressed by all three doses of TGP, whereas the mRNA expression of COX-2 was not affected by TGP. The mRNA expression of HO-1 was increased by TGP (Figure 6).

iNOS, COX-2, HO-1 and CYP2E1 protein expression

The protein expression of iNOS and COX-2 was significantly higher in the CCl₄-treated mice than in the control mice. The increase in the protein expression of iNOS and CYP2E1 was significantly suppressed by 100 mg/kg TGP; however, TGP had little effect on COX-2 protein expression, consistent with the changes in mRNA expression. The protein expression of HO-1 was significantly higher in the CCl₄-treated group than in the control group, and this increase was augmented by 100 mg/kg TGP (Figure 7).

Discussion

In this study, we investigated the protective effects of TGP on CCl₄-induced experimental liver injury in mice, and the underlying mechanism. It is generally accepted that CCl₄-induced hepatotoxicity involves the metabolism of CCl₄ by cytochrome P450 and the activation of Kupffer cells. The former leads to the generation of free radicals and lipid peroxidation, which may actuate the later.

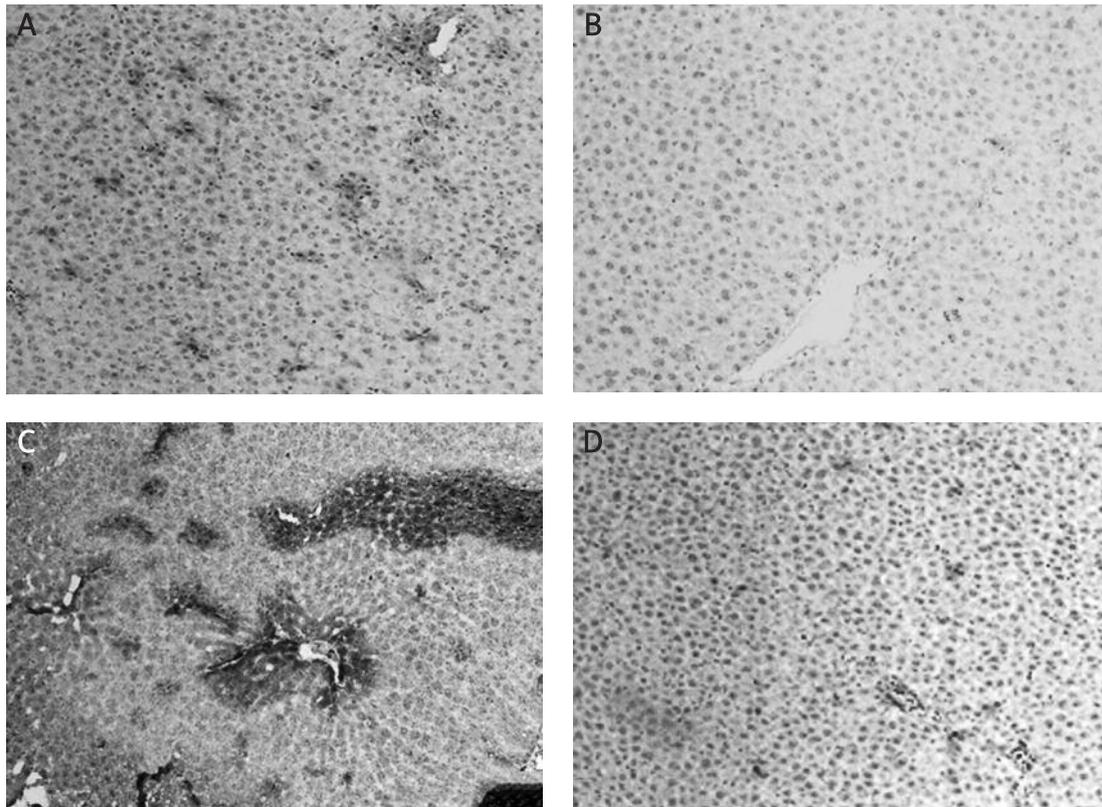


Figure 5. Immunohistochemical analyses showed that TGP suppressed the increased TNF- α and NF- κ B 65 expression induced by 8 weeks of CCl₄ treatment. **A** – TNF- α expression in the CCl₄-treated group, **B** – the 100 mg/kg TGP-treated group, **C** – NF- κ B 65 expression in the CCl₄-treated group, **D** – the 100 mg/kg TGP-treated group. Bars, 100 μ m

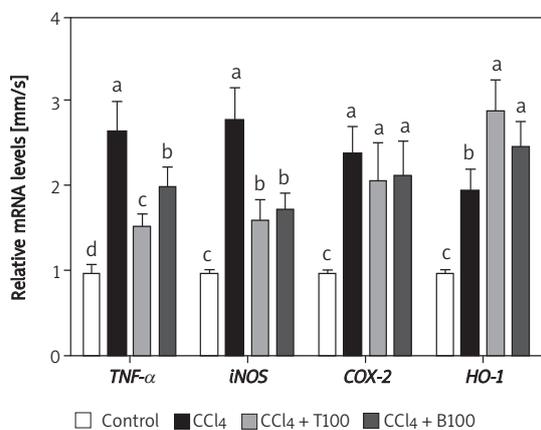


Figure 6. Effects of 100 mg/kg TGP on the mRNA expression of *iNOS*, *COX-2* and *HO-1* in the liver of CCl₄-treated mice. Values are means \pm SD of five mice per group. Significantly different ($p < 0.05$) from the control group. Means without a common letter differ at $p < 0.05$
T100 – TGP 100 group, *BDP100* – BDP 100 group

Meanwhile, the activation of Kupffer cells is accompanied by the production of proinflammatory mediators [27, 28].

Total glucosides of peony is relatively inexpensive, with negligible side effects, and shows great

potential in the treatment of liver diseases. Previous studies have demonstrated that the antioxidative effects of TGP are mediated by the inhibition of thiobarbituric acid (TBA) reactive substance, an index of endogenous lipid peroxidation, and enhanced activities of antioxidant enzymes such as superoxide dismutase, catalase and glutathione peroxidase following cerebral ischaemia and immunological liver injury [29].

During hepatic injury, deteriorated integrity of the cellular membrane causes release of enzymes into the circulation, reflecting damage to the hepatic cells [30]. Thus, abnormally high levels of serum hepatospecific enzymes such as ALT and AST are regarded as good biomarkers for liver injury. We found that TGP inhibited the elevations of ALT and AST in CCl₄-induced liver injury. The attenuated increase in serum ALT and AST levels suggests that TGP improved the structural integrity of the hepatocellular membrane, thus protecting mice against CCl₄-induced hepatotoxicity. Histological analyses showed that CCl₄ caused a variety of histological changes in the liver, including centrilobular necrosis, portal inflammation, Kupffer cell hyperplasia, extensive fatty changes, and even hepatic fibrosis and cirrhosis. These histo-

pathological changes were significantly attenuated by TGP. Our findings suggest that the hepatoprotective effect of TGP was better or at least the same as that of BDP, a potent hepatoprotective agent. Although BDP has a marked ALT-lowering effect [31], this is transient and relapse is likely. Hence, its clinical application is limited. The present results suggest that TGP could be used to treat liver diseases.

Previous studies have indicated that the induction of CYP2E1 by CCl₄ *in vivo* leads to the generation of trichloromethyl free radicals and ROS, which initiate lipid peroxidation [32]. In turn, lipid peroxidation stimulates the expression of TNF- α and inflammatory mediators (e.g., iNOS and COX-2), and activates the signalling pathways that are classically associated with inflammation, such as NF- κ B, which might in turn further trigger the production of toxic compounds such as ROS or nitric oxide (NO), inflammatory cytokines and glutamate, thus causing liver damage [33]. Enhanced antioxidant activity may be one mechanism through which drugs may protect against CCl₄-induced liver injury.

A recent report suggested that TGP ameliorated immunological hepatic fibrosis in rats caused by inhibitory oxidation and nitration [34]. In the present study, TGP inhibited CYP2E1 expression and consequently suppressed the increase in gene expression of TNF- α and iNOS, suggesting that TGP regulates CCl₄-induced production of TNF- α and iNOS at the transcriptional level. Total glucosides of paeony blocked the CCl₄-mediated oxidative stress and lipid peroxidation by significantly reducing the levels of by-products of lipid oxidation (i.e., hepatic MDA) and serum CAT levels, suppressing the activities of serum SOD and GSH, and inhibiting the increase in TNF- α levels. Concomitantly, the increased protein level of NF- κ B was reduced by TGP treatment. These results indicate that TGP exerted its hepatoprotective effects by improving *de novo* protein synthesis, particularly of HO-1, and counteracting the increased formation of free radicals. Moreover, HO-1 is believed to play an important protective role against oxidative injury [35]. Overexpression of HO-1 might be closely associated with the protective mechanism of some molecules against CCl₄-induced hepatic injury. Our findings suggest that the gene expression of HO-1 increased markedly within 24 h after CCl₄ administration, and was maintained to the end of the study, comprising an important mechanism in the protective effects of TGP in CCl₄-induced liver injury. However, in this study, the gene and protein expression of COX-2 increased significantly in response to CCl₄ and was not affected by TGP. More studies are needed to examine this effect in more detail and explore the underlying mechanisms. Meanwhile, the physiology

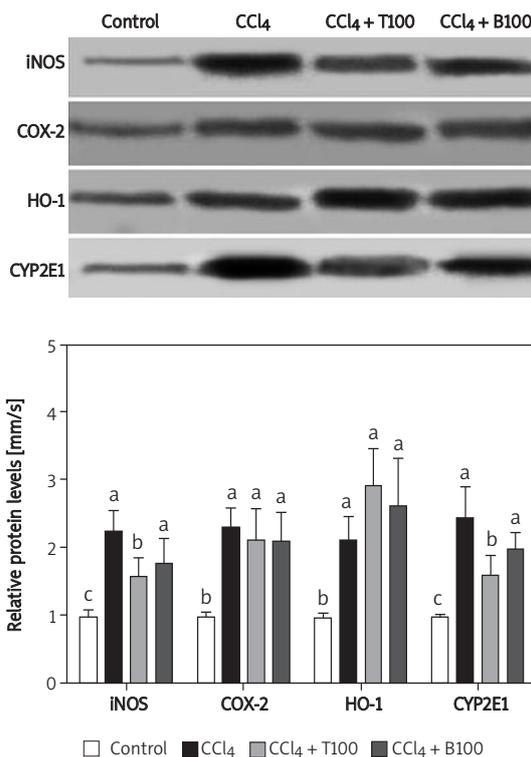


Figure 7. Effects of TGP (100 mg/kg) on the protein expression of iNOS, COX-2, HO-1 and CYP2E1 in the liver of CCl₄-treated mice. Values are means \pm SD of five mice per group. Significantly different ($p < 0.05$) from the control group. Means without a common letter differ at $p < 0.05$

T100 – TGP 100 group, BDP100 – BDP 100 group

and biochemistry of rodents are somewhat different from those observed in people. Further experimental studies in both the rodents and humans and subsequently all phases of clinical trials in humans are also required before TGP is used in clinical study.

In conclusion, TGP protects the liver from CCl₄-induced oxidative stress and the inflammatory response, without any known side effects. While the applicability of the findings of this work is very far from any conceivable clinical application in humans, the possible role of TGP as a promising therapeutic in human oxidative stress and inflammatory liver disease deserves further consideration. The potential for TGP in experimental and practical applications should be examined in future studies.

Acknowledgments

This study was supported partly by research grants from the National Science Foundation for Post-doctoral Scientists of China (No. 20080431363) and the National Natural Science Foundation of China (No. 20635002).

The authors declare that they have no competing interests.

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