# Hepatic glycogen storage diseases: pathogenesis, clinical symptoms and therapeutic management

Edyta Szymańska<sup>1</sup>, Dominika A. Jóźwiak-Dzięcielewska<sup>2</sup>, Joanna Gronek<sup>2</sup>, Marta Niewczas<sup>3</sup>, Wojciech Czarny<sup>4</sup>, Dariusz Rokicki<sup>1</sup>, Piotr Gronek<sup>2</sup>

<sup>1</sup>Department of Gastroenterology, Hepatology, Feeding Disorders and Pediatrics, The Children's Memorial Health Institute, Warsaw, Poland

<sup>2</sup>Laboratory of Genetics, Department of Gymnastics and Dance, University School of Physical Education, Poznan, Poland

<sup>3</sup>Department of Sport, Faculty of Physical Education, University of Rzeszow, Rzeszow, Poland

<sup>4</sup>Department of Human Sciences, Faculty of Physical Education, University of Rzeszow, Rzeszow, Poland

Submitted: 13 October 2017; Accepted: 8 December 2017; Online publication: 18 February 2019

Arch Med Sci 2021; 17 (2): 304-313 DOI: https://doi.org/10.5114/aoms.2019.83063 Copyright © 2019 Termedia & Banach

#### Abstract

Glycogen storage diseases (GSDs) are genetically determined metabolic diseases that cause disorders of glycogen metabolism in the body. Due to the enzymatic defect at some stage of glycogenolysis/glycogenesis, excess glycogen or its pathologic forms are stored in the body tissues. The first symptoms of the disease usually appear during the first months of life and are thus the domain of pediatricians. Due to the fairly wide access of the authors to unpublished materials and research, as well as direct contact with the GSD patients, the article addresses the problem of actual diagnostic procedures for patients with the suspected diseases. Knowledge and awareness of the problem among physicians seem insufficient, and research on the diagnosis and treatment of GSD is still ongoing, resulting in a heterogeneous GSD typology and a changing way of its diagnosis and treatment.

Key words: glycogen storage disease, hypoglycemia, hepatomegaly.

### Introduction

Glycogen is a complex/polymeric physiological glucose carbohydrate found mainly in the liver but also in skeletal muscles, kidneys and enterocytes. Its role is to store glucose, which is used as an endogenous source of energy following enzymatic digestion (glycogenolysis process) within 3–4 h after the meal in order to maintain normoglycemia. Glycogen is, therefore, a source of glucose obtained from hepatocytes during a short period of fasting or used for the needs of skeletal muscles in the first early phase of myocytes [1]. Glucose deficiency in the central nervous system (CNS) can lead to neuroglycopenia [2]. There might be symptoms similar to hypoxia, convulsions, irritability and cognitive decline. Permanent and prolonged hypoglycemia may lead to permanent CNS damage [3]. Systemic hypoglycemia is characterized by signs of sympathetic stimulation/activation, including tachycardia, excessive sweating, fatigue, muscle weakness, irritability, anger, hunger, seizures, headaches and dizziness [4].

#### **Corresponding author:**

Edyta Szymańska MD, PhD Department of Gastroenterology, Hepatology, Feeding Disorders and Pediatrics The Children's Memorial Health Institute Al. Dzieci Polskich 20 Warsaw, Poland Phone: +48 22 815 74 94 E-mail: edyta.szymanska@ ipczd.pl



Attribution-NonCommercial-ShareAlike 4.0 International (CC BY -NC -SA 4.0). License (http://creativecommons.org/licenses/by-nc-sa/4.0/).

Creative Commons licenses: This is an Open Access article distributed under the terms of the Creative Commons

Under physiological conditions, the liver plays a key role in the metabolism of consumed carbohydrates by storing them in the form of glycogen, thereby preventing postprandial hyperglycemia [5]. Synthesis of glycogen consists in combining glucose monomers with glycogen protein and  $\alpha$ 1,4- or  $\alpha$ 1,6-glycosidic bonds [6]. This process can be quickly reversed when the energy requirements in the body cells rise again, and the blood glucose levels fall (Figure 1).

Glycogen itself is a branched glucose polymer that is osmotically passive [7]. Due to the activity of many enzymes, hormone regulation and proper conditions in the cell, a healthy body "regulates" and "adjusts" glucose metabolism processes, sometimes "giving warning signals" when it is no longer able to use its own energy resources [3]. Irregularities in the above-mentioned metabolic processes cause problems for the basic functioning of the body [8]. Changes related to the disorder of glycogen metabolism are characterized by symptoms varying from developmental delay, especially of motor functions, to serious and immediate neuroglycopathy, which is a life-threatening condition. In addition, as a result of the accumulation of abnormal glycogen molecules, fatty liver occurs, and in extreme cases the storage of abnormal glycogen forms lead to fibrosis and cirrhosis (GSD type IV) [9].

To keep blood glucose levels within a narrow range (70–140 mg/dl) around the clock, hepatic absorption of postprandial glucose is counterbalanced by its "peripheral" (through other organs) consumption. When the blood glucose level drops below the critical lower limit (long after food intake or effort), the body is ready for a certain compensation and attempts to derive it from non-sugar precursors (for example alanine), but some processes cannot be replaced or stopped [10]. An example is metabolic acidosis with increased blood lactate level with no characteristic symptoms. It is associated with accumulation of lactic acid in cells above 5 mmol/l [11]. This condition results from acid-base balance disorders, and the symptoms are weakness, nausea, vomiting, diarrhea, and abdominal pain [11]. The glucose metabolism pathway in the body can be modulated by using, among other things, a diet rich in complex carbohydrates [12].

In this manuscript, the authors discuss GSDs in general in the first part of the paper, and then, later in the text, they focus only on the hepatic types.

## Glycogen storage diseases (glycogenoses)

#### Pathophysiology and epidemiology

Glycogen storage disease (GSD) is caused by a genetically determined metabolic block involving enzymes that regulate synthesis (glycogenesis) or glycogen breakdown (glycogenolysis) [13]. The nature of the individual types of glycogenosis is related to the impaired accumulation of abnormal molecules of this branched polysaccharide [14]. Moreover, GSD, although it is not a semantically correct name, also refers to a deficit in glycogen synthase (the so-called type 0 GSD), which consists in the lack of glycogen in tissues due to the inability to polymerize glucose molecules (lack



Figure 1. Cascade of glycogen breakdown

of enzyme catalyzing the glycogen synthesis reaction) [15].

In the International Statistical Classification of Diseases and Related Health Problems (ICD-10), glycogenosis is placed under the symbol E74.0 [icd10.pl].

Most genetically determined diseases are rare [16]. These are chronic conditions, usually progressive and mostly severe. The prevalence of metabolic blocks associated with glycogen is estimated at 1 : 25,000 (40,000) births [17].

The typical, biochemical characteristics of this type of GSD are postprandial hyperglycemia (blood glucose cannot be stored in the liver and therefore it is kept high in the blood) with subsequent hypoglycemia (no hepatic glycogen storage from which the body could 'boost' the reserves in case of a drop in the blood glucose).

Individuals affected by glycogenosis in Poland may apply for a disability ruling due to the disorder which, irrespective of the source of its occurrence, causes disorder of the body functionality or limitation in performing life and social activities.

### Inheritance

Glycogen storage diseases, like most metabolic diseases, are inherited in an autosomal recessive (AR) way. These are the innate defects of carbohydrate metabolism that affect the liver, muscles or both of these organs (the so-called mixed types) [18, 19]. The degree of consanguinity determines the AR inherited diseases in a given area or population – where the risk of mutation is high, which does not exclude *de novo* mutations. The IX-al-pha glycogenosis is the only GSD type which is X-linked and inherited recessively [20].

## Typology of GSDs

It is still an unsettled matter. There are categorizations according to different evaluation criteria: author [6], enzyme [21], occupied tissues [17] or mutation location. The classification by author in the order of discovering/describing individual GSDs is generally accepted and applied. The nomenclature comes from the author of the pioneering study, the method of treatment, or the diagnosis of the case of glycogen storage, since the time when this disorder was not classified as a separate disease unit. This division can also be found in all sorts of general papers on medical biochemistry, genetics, and in pediatrics or neonatology.

Another attempt to classify GSDs is the Shin [21] study, where the glycogenosis classification is precisely categorized according to the enzymatic substrate of glycogen metabolism and the sub-types associated with certain organs, giving a to-tal of 12 GSDs (Table I).

Özen [17] classifies GSDs on the basis of the latest knowledge in the field, their enzymatic deficits or involved tissues, mainly on the basis of available case studies. Case studies and data collection during patient health monitoring prove an increase in the number of GSDs studies [18, 22, 23]. The knowledge and awareness of these complex metabolic pathologies are increasing. The author divided the aforementioned disorders into hepatic, mixed, and muscular GSDs (Table II) [17].

## **Clinical symptoms**

The basic clinical symptoms common for every type of hepatic GSD are hypoglycemia and hepatomegaly [24]. A characteristic feature for muscle GSDs is progressive muscle pathology, including exercise-induced muscle weakness [25]. Metabolic acidosis with hypercholesterolemia and hyperlipidemia are found in blood biochemistry [26]. In both types of GSDs, there is also hypertransaminasemia (elevated activity of liver enzymes – aminotransferases). Physical development delay, mostly in the form of short stature or motor latency, is a typical clinical symptom. GSDs are a group of heterogeneous genetic diseases; therefore each type has its distinct, specific clinical presentation [27].

## Diagnosis

The diagnosis of GSD is based on clinical signs and/or symptoms related to hypoglycemia and hepatomegaly. Laboratory parameters such as increased lactate level (in GSD type I), elevated level of serum cholesterol and triglycerides and hypertransaminasemia are helpful in diagnosis establishment. The key point is to differentiate between GSD type I and so-called ketotic types (III/VI/ IX). Symptoms in GSD I typically present earlier (in the first few months of life) with severe fasting hypoglycemia within 3–4 h after feeding. Hypoglycemia is usually less severe in patients with GSD III/ VI/IX due to the intact process of gluconeogenesis [28]. Blood lactate levels increase rapidly in GSD I as blood glucose (BG) concentrations decrease to levels that normally trigger a counter-regulatory response (< 70 mg/dl or 4 mmol/l) and are markedly increased when BG levels decrease to < 40–50 mg/dl or 2.2–2.8 mmol/l). Blood  $\beta$ -hydroxybutyrate levels increase only modestly in GSD I, in contrast to marked hyperketonemia with fasting hypoglycemia characteristic of GSD 0, III, VI, and IX [29].

Other biochemical characteristics that help to distinguish between these disorders are elevated uric acid and lactate levels in GSD I, whereas these are typically normal in ketotic GSDs [30].

The final diagnosis is always established or confirmed with molecular tests. Liver biopsy, in

уре	Deficient enzyme	Gene symbol
arious types of glycogen storage disease types I–I	IV:	
l (von Gierke):		
la	Glucose 6-phosphatase	G6PC
lb	G6P translocase (T1)	SLC37A4
lc	Phosphate translocase (T2)	NPT4(?)
Id	Glucose translocase (T3)	Not specified
Infantile (Pompe disease)	Lysosomal $\alpha$ -glucosidase	GAA
Childhood	Lysosomal $\alpha$ -glucosidase	GAA
Juvenile	Lysosomal $\alpha$ -glucosidase	GAA
Adult	Lysosomal $\alpha$ -glucosidase	GAA
III (Cori disease):		
IIIa (Liver and muscle form)	Amylo-1,6-glucosidase	AGL
IIIb (Liver form)	Amylo-1,6-glucosidase	AGL
IIIc (Muscle form)	Amylo-1,6-glucosidase	AGL
IV (Andersen disease):		
Juvenile form	Branching enzyme	GBE1
(Liver)	Branching enzyme	GBE1
(Neuromuscular)	Branching enzyme	GBE1
Juvenile or adult form (liver, muscle)	Branching enzyme	GBE1
Polyglucosan body disease (APBD)	Branching enzyme	GBE1
arious types of glycogen storage disease types V-	-IX and 0:	
V (McArdle disease):		
Adult form	Muscle phosphorylase	PYGM
Infantile form	Muscle phosphorylase	PYGM
VI (Hers disease)	Liver phosphorylase	PYGL
VII (Tarui disease):		
Severe form	Phosphofructokinase	PFKM
Mild form	Phosphofructokinase	PFKM
Phosphorylase activation system defects		
VIII (Via/IXA):	Phosphorylase kinase (liver PBK)	
(XLG I/II)	$\alpha$ -subunit of PBK	PHKA2
Autosomal recessive	β-subunit of PBK	РНКВ
IXB	$\gamma$ or $\delta$ subunit of PBK (?)	PHKG2
IXC	Cardiac muscle PBK	Not specified
IXD (adult form)	Muscle PBK	PHKA1
(Severe muscle form)		PHKA1(?), PHKG1(?
X (multisystem)	Protein kinase (?)	Not specified
GSD 0	Glycogen synthase (liver)	GYS2
	Glycogen synthase (muscle)	GYS1

 Table I. Various types of GSDs (types of GSDs according to tissue-specific enzymatic deficiency)

Hepatic GSD (most dysfunction in liver)	Phosphorylase kinase system defects	Muscle GSD
<ul> <li>GSD type 0/glycogen synthase deficiency</li> <li>GSD type I/von Gierke disease/ glucose-6-phosphatase deficiency/ hepatorenal glycogenosis.</li> <li>Subtypes: Ia, Ib, Ic, Id</li> <li>GSD type III/Cori disease/Forbes disease/amylo-1,6-glucosidase deficiency/glycogen/debrancher deficiency</li> <li>Subtypes: IIIa, IIIb, IIIc, IIId</li> <li>GSD type IV/Andersen disease/ amylopectinosis/brancher deficiency/glycogen branching enzyme deficiency</li> <li>GSD type VI/Hers disease/liver glycogen phosphorylase deficiency</li> <li>GSD type XI/ Fanconi-Bickel syndrome (FBS)</li> </ul>	<ul> <li>Mutation of subunit α/X-linked liver phosphorylase kinase deficiency/XLG</li> <li>Subtypes: XLG I, XLG II (formerly GSD VIII or GSD VIa)</li> <li>Mutation of subunit β/autosomal liver and muscle phosphorylase kinase (β subunit) deficiency</li> <li>Mutation of subunit γ/autosomal liver phosphorylase kinase (γ subunit) deficiency (testis/ liver; cirrhosis due to liver phosphorylase kinase deficiency)</li> </ul>	<ul> <li>GSD type II/Pompe disease/ acid α-glucosidase deficiency/ acid maltase deficiency/α1,4- glucosidase deficiency</li> <li>Subtypes:</li> <li>Classic</li> <li>Infantile</li> <li>Juvenile</li> <li>Adult</li> <li>GSD type V/McArdle disease/ myophosphorylase deficiency/ muscle glycogen phosphorylase deficiency</li> <li>GSD type VII/Tarui disease/muscle phosphofructokinase deficiency/ glycogen storage disease of muscle</li> </ul>

Table II. Classification of GSDs into hepatic and muscle types with separate classification of PhK deficiency (type IX)

order to assess enzymatic activity in hepatocytes, is no longer the gold standard – it has been replaced by genetic examination, which is non-invasive and gives a certain and final diagnosis [29, 30]. The disadvantages of molecular testing are its still high costs, limited availability and long time of waiting for the results.

The Children's Memorial Health Institute in Warsaw (Department of Pediatrics, Nutrition and Metabolic Diseases) is the reference center for metabolic diseases.

#### Glycogen pathway

Decomposition of glycogen molecules is directly related to energy production, and its regulation takes place by activation of the relevant substances and enzymes. In a healthy body, glycogen metabolism is effective through the balance between glycogen synthase and phosphorylase activity (Figure 2). Both enzymes can be reversibly



Figure 2. Enzymatic activity of phosphorylase and synthase

phosphorylated in more than one site by separate kinases [8].

Glycogenolysis (branched glucose polymer phosphorolysis) is carried out in three steps:

- 1. Release of glucose-1-phosphate-kinase phosphorylase (PhK).
- 2. Conversion of glucose-1-phosphate to glucose-6-phosphate – phosphoglucomutase.
- 3. Glucose-6-phosphate metabolism glucose 6-phosphatase-6-phosphatase 1 (G6PC).

Glycogenolysis takes place in the liver, during hunger or in the muscles during intensive exercise, to compensate for a decrease in blood glucose levels or a lack of chemical energy carriers in cells – ATP [31].

In the early phase of glycogenolysis, the released glucose is already phosphorylated and can undergo further transformation. In this state it cannot freely leave the cell, which makes it a very energetically beneficial reaction for the working muscle. However, there are circumstances in which glycogen metabolism is inadequate.

## GSD type IX

The GSD type IX accounts for about 25% of all glycogenosis and consists of a lack of phosphorylase kinase (PhK) enzyme, which plays a role in the process of glycogen degradation. The enzyme occurs in many tissues, thus its subunits also have their specific isoforms. The activity of PhK has been studied in such organs as the liver, muscles, kidney, testes, heart and also in erythrocytes, leukocytes and nerve cells [32, 33].

The PhK enzyme consists of 4 subunits:  $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$  subunits, which are encoded by the following genes:

– PHKA1 and PHKA2 (α1 and α2 subunits) expressed in the liver;

– PHKB (subunit β), PHKG1 and PHKG2 (subunit  $\gamma$ ) expressed in the liver [34, 35],

– CALM1, CALM2 and CALM3 (subunit  $\delta$ ) [32].

The  $\alpha$  and  $\beta$  subunits occupy 4/5 of the total weight of the holoenzyme and have a superior function that regulates phosphorylation. The cAMP-dependent protein kinase regulates the phosphorylation of the Ser residues in the  $\alpha$  and  $\beta$  subunits. The  $\gamma$  subunit contains a catalytic site, and the calcium-binding subunit  $\delta$  has an affinity for calmodulin [36].

Type IX is the only GSD that is X-chromosome linked recessively inherited: the  $\alpha 1$  subunit is X-linked inherited, while the remaining subunit units are AR inherited [37]. Mutation in the PHKA2 gene is the most common cause of GSD IX [38]. This type of GSD is characterized by a relatively mild course. Tissues affected by the disorder, depending on the subtype, are liver, erythrocytes, kidneys and muscles [39, 40]. The symptoms occur during infancy or early childhood, and include growth delay and liver enlargement [41]. Episodes of hypoglycemia or ketonuria without a reasonable cause are rare. If present, they are associated with a prolonged state of starvation or increased physical activity. Lactic acid and urinary acid concentrations are usually normal, and metabolic acidosis with hypocalcemia is rare. Hypertriglyceridemia and hypercholesterolemia, as well as various levels of hypertransaminases, are also observed: from slightly to several times elevated transaminases. It causes delays in motor development due to the possibility of spreading in the muscle tissue. These disorders normalize during adolescence [13]. Occasionally, these GSDs are associated with more serious complications such as cirrhosis of the subtype IX  $\gamma$  or cardiomyopathy/ megalocardial dysfunction [42-44].

## GSD type III and VI

Type III of GSD is caused by a reduced enzyme degradation enzyme (amylo-1,6 glucosidase + transferase – GDE), which is a key enzyme in the glycogenolysis process [45]. Based on the additional involvement of the skeletal muscles and the heart, there are 2 subtypes of GSD III: hepatic-muscle type (subtype a) and a hepatic one (subtype b). Like all other GSDs but type IX- $\alpha$ , it is inherited in an autosomal recessive manner; the mutation is in the *AGL* gene located on chromosome 1p21 [46].

The first signs of the disease appear in early childhood and include enlarged liver, delayed growth and physical development, whereas hypoglycemia (neurohyglycemia) is not that common. As age progresses, the hepatomegaly reverses, and muscle weakness (subtype IIIa) progresses slowly. Other symptoms common to this type of GSD include muscle hypotension and cardiomyopathy. Frequently, the symptoms regress during adolescence, except in rare cases when cirrhosis of the liver or myopathy occurs [47].

Type VI is a mild form of GSD. The enzyme block consists of decreased liver enzyme activity – phosphorylase involved in the glycogenolysis process. The disease is inherited in an autosomal recessive manner, with a mutation in the *PYGL* gene on chromosome 14q21-q22 [48]. Symptoms are the same as in type III. In this type of GSD the heart and skeletal muscles are never involved, and liver adenomas are very rare. The disease does not carry the risk of organ failure [49].

## GSD type IV

Type IV is a rare and severe form of GSD, accounting for approximately 3% of all types of glycogenosis. The enzyme defect consists of reduced glycogen brancher enzyme (GBE) activity, mutation in the *GBE1* gene on chromosome 3p12, and autosomal recessive inheritance. GBE deficiency results in the accumulation of abnormal forms of glycogen that resemble the amylopectin (polyglucosan body) structure. Hence, its other name is amylopectin or adult polyglucosan body disease (APBD) [50].

The symptoms are very heterogeneous and include both the liver and the neuromuscular system. Children are generally born healthy but, as early as in the first months of life, they develop hepatomegaly and hypotonia and their psycho-motor development is delayed. The disease progresses rapidly. It leads to liver fibrosis and portal hypertension which manifests as ascites and eventually leads to death. However, several cases of non-available hepatic GSD IV have also been reported in the literature [51].

## GSD type I

Unlike muscles, liver contains the glucose-6-phosphatase membrane enzyme, which removes the phosphate residue to allow the glucose to enter the bloodstream and regulates its concentration [52].

Glucose-6-phosphatase 1 (G6PC) enzyme catalyzes the hydrolysis of glucose-6-phosphate to glucose, thus forming the last step of glycogenolysis and gluconeogenesis [53–56]. The G6PC enzyme is encoded by the G6pc gene, which is expressed in the liver, kidneys and pancreas, and its mutation is inherited in an AR pattern [52, 57]. The function of the enzyme requires its translocation through the membrane of the endoplasmic reticulum. Another enzyme, G6PC translocase encoded by the SLC37A4 gene, is also involved in this process, which can also be mutated (AR inheritance) to impair the function of neutrophils, which is inscribed in the GSD type Ib.

After eating a meal, blood glucose rises. Hormone levels regulating these metabolic pathways are in inverse proportions, i.e. the concentration of glucagon decreases, and the concentration of insulin increases. There is no breakdown of glycogen then. The phosphorylation pathway of inactivated phosphorylase (b) to active (a) is off. On the other hand, the pathway of phosphorylation of active synthase (a) to inactive (b) is activated. We are dealing with the postprandial state of glycogen synthesis stored as a backup material in tissues, primarily in the liver. Before meals, when blood glucose levels fall, insulin levels drop, and another hormone, glucagon (adrenalin-like), shows an upward trend. The glycogen degradation reaction to glucose is started. We obtain an active form of glycogen phosphorylase (a) and an inactive form of glycogen synthase (b).

#### **Dietary management**

The basic treatment for GSDs is dietary management, which raises many controversies. Dietary recommendations differ slightly between the United States and Poland, but they are different in Europe, for example, comparing the UK and Poland.

The aim of dietary treatment is to avoid hypoglycemia. Therefore, it is necessary for patients to consume frequent meals during the day (every 3–4 h) with the addition of raw cornstarch (RS) and to shorten the night break by an additional portion of RS in the middle of the night [58–60]. Starch, like glycogen, is a polysaccharide – the process of releasing pure glucose into the bloodstream, in this case, is extended in time. This results in constant access of substrates to the biochemical reactions of energy synthesis pathways, limiting its accumulation in tissues and thereby involving glycogen [61, 62]. In the UK, overnight glucose infusions through a pump or probe are still used.

Studies have confirmed that a snack before bed time is a sufficient energy/glucose supply for patients with GSD type III/VI/IX. However, for patients with an unstable glucose level or ketosis, 2 g of unprocessed RS per kilogram of body weight is prepared to prevent morning hypoglycemia [63]. The situation is different for patients with type I GSD, who present with the most severe hypoglycemia and have most unstable glucose levels both glycogenolysis and gluconeogenesis are impaired in this type and, additionally, ketogenesis is ineffective and there is a risk of neuroglycopenia. Therefore, in GSD type I starch is a drug and not just a dietary supplement; it must be given regularly during the day every 3–4 h with a night break not exceeding 7-8 h [30]. Patients most often need to take an extra dose of starch at night while maintaining a constant supply of carbohydrates in a liquid form, especially during sleep, thus protecting patients with decompensated glycemia from dangerous complications of hypoglycemia.

When intolerance or reluctance to receive RS is observed, either a nasogastric tube or percutaneous endoscopic gastrostomy (PEG) is required - if the enteral supply is to be maintained for more than 6-8 weeks (ESPEGHAN guidelines). The disadvantage of starch is that it is hard to digest with an infection associated with gastrointestinal irritation in children with glycogenosis there are often problems with starch supplementation. In addition to child monitoring, an indispensable element in hypoglycemia and RS treatment is blood glucose measurement with a glucometer, and in the case of GSD III/VI/IX ketones in the urine are to be measured. In these types and in type Ia, it is possible to provide a modified, artificially produced starch called "Glycosade", which, due to its prolonged release, allows normoglycemia to be maintained for an 8-hour nighttime rest. It is not recommended for patients with type Ib due to their common inflammatory bowel disease and, therefore, additionally impaired absorption. The formula is used in patients older than 5 years old. Currently, glycosade is to be studied in the United States for its application during the day in adult patients, in whom, due to the slower metabolism than children, the extended-release preparation might be effective not only at night. According to current US dietary recommendations, the diet should be restricted to simple sugars (less than 5–10 g in each meal) in all types of GSDs, while it is most restrictive in type I, with RS supplementation (in type I every 3-4 h daily plus an extra overnight dose or only 1 night dose in other types) [64].

Type III/VI/IX is recommended to use the socalled high-protein diet (GSD type III; 3-4 g/kg BW, VI and IX: 2–3 g protein per kilogram BW per day) [65]. Because of the increased risk of micronutrient deficiencies (vitamin D<sub>3</sub>, calcium-phosphate disturbances, and the risk of osteopenia), multivitamins and vitamin D<sub>3</sub> supplements are also recommended [30, 64]. The basic principle of treatment is therefore to limit the simple sugars in a diet rich in complex carbohydrates. However, the diet is always individually selected for the patient based on its glycemic status, current biochemical findings (metabolic equilibrium) and anthropometric parameters, and requires close collaboration between the physician, metabolic dietitian, the patient and his family.

Table III summarizes different dietary strategies in ketotic and non-ketotic (type I) GSDs.

As mentioned above, a properly applied specific diet is a treatment in GSDs. It leads not only to stable normoglycemia, but also to decreased hepatomegaly (reduction of glycogen storage), improvement of growth and biochemical meta-

Table III. Summary of different	dietary strategies in ketotic and	non-ketotic (type I) GSDs
---------------------------------	-----------------------------------	---------------------------

Dietary management		
All GSDs – Restriction of simple sugars – Frequent meals during the day – "around the clock"		
GSD I: – CS every 3–4 h during the day plus extra dose at night	GSD III/VI/IX: – Usually only 1 dose of CS at night – High-protein diet	

bolic control (normalization of transaminases, triglycerides, in type I – reduction of lactate and uric acid in blood) [13].

The problem of diet, as an important factor in treatment, may lead to eating disorders in children with GSDs. In the course of metabolic diseases, some eating disorders related strictly to the (elimination) diet can be observed. For example, people consuming frequent meals with high carbohydrate content that slows down the release of glucose into the blood may experience a lack of hunger. Since a lot of attention is paid to eating, it may cause a lack of pleasure from eating a meal, not to mention the lack of taste of non-sweet foods. As far as children are concerned, following the heavy-starch diet is often associated with digestive problems. Children who have previously consumed sweet products being deprived of candy may feel alienated among their peers, in a particular environment, for example, in a pre-school class where the group's diet contains sweet supplements, unlike meals prepared for a child with GSDs. Establishing the right diagnosis and setting a proper glycogenic diet will make the patients enjoy eating and discovering new flavors, changing the attitude to a meal as a necessity. In the treatment of eating disorders, therapies of these disorders adapted to the requirements of GSDs are helpful [28].

The GSDs, like all genetic diseases, is incurable. Currently, clinical trials on gene therapy for type I glycogenosis (clinical phase of GSDI adult safety assessment) are ongoing at the Connecticut facility in the US [66].

#### Gene therapy

Currently, in the USA, the development of adeno-associated virus (AAV) vector-mediated gene therapy is being carried out for GSD type I based on the success of early-stage clinical trials of gene therapy in hemophilia [67]. So far, AAV vectors containing a human G6Pase regulatory cassette/ promoter have proven to be efficacious in animal models of GSD Ia, and these vectors contain sequence elements that regulate G6Pase expression appropriately [68]. At present, gene therapy for GSD type I is at the stage of a safety clinical trial on adult patients with this type of GSD, and is taking place in the Connecticut Hospital within the GSD program of Prof. Weinstein.

#### Conclusions

The GSD is a congenital defect of carbohydrate metabolism characterized by hypoglycemia, hepatomegaly, and growth disorders (short stature). Basic therapeutic treatment consists in maintaining a proper diet with RS supplementation.

Due to the fact that awareness and knowledge about rare diseases are still insufficient, it is important to popularize them among pediatricians, hepatologists and geneticists. Knowledge of the biochemical basics and glucose metabolism in the human body facilitates proper treatment of GSD.

#### **Conflict of interest**

The authors declare no conflict of interest.

References

- 1. Zhai L, Feng L, Xia L, Yin H, Xiang S. Crystal structure of glycogen debranching enzyme and insights into its catalysis and disease – causing mutations. Nat Commun 2016; 7: 11229.
- Wang HF, Wu KH, Tsai CL. Neuroglycopenia in an euglycaemic patient under intensive insulin therapy. Anaesth Intensive Care 2010; 38: 1137-8.
- 3. Joung-Hwan J, Streamson CC Jr. The brain liver connection between BDNF and glucose control. Diabetes 2013; 62: 1367-8.
- 4. Douillard C, Menton K, Dobbelaere D, Wemeau JL, Saudubray JM, Vantyghem MC. Hypoglycaemia related to inherited metabolic disease in adults. Orphanet J Rare Dis 2012; 7: 26.
- 5. Moore MC, Katie C, Coate J, et al. Regulation of hepatic glucose uptake and storage in vivo. Adv Nutr 2012; 3: 286-94.
- 6. Koolman J, Roehm KH. Color Atlas of Biochemistry. 2<sup>nd</sup> edn. Thieme Stuttgart, New York 2005.
- 7. Lee PJ, Bhattacharya K. Glycogen Storage Diseases. Oxford University Press, Oxford 2013.
- Murray RK, Granner DK, Mayes PA, Rodwell VW. Harper's Illustrated Biochemistry. 26<sup>th</sup> edn. Lange Medical Books/McGraw – Hill Medical Publishing Division, 2003.
- 9. Kneeman JM, Kneeman JM, Misdraji J, Corey KE. Secondary causes of nonalcoholic fatty liver disease. Therap Adv Gastroenterol 2012; 5: 199-207.
- 10. Chung ST, Chacko SK, Sunehag AL, Haymond MW. Measurements of gluconeogenesis and glycogenolysis: a methodological review. Diabetes 2015; 64: 3996-4010.

- 11. Morris CG, Low J. Metabolic acidosis in the critically ill: part 1. Classification and pathophysiology. Anaesthesia 2008; 63: 294-301.
- Hochuli M, Christ E, Meienberg F, et al. Alternative nighttime nutrition regimens in glycogen storage disease type I: a controlled crossover study. J Inherit Metab Dis 2015; 38: 1093-8.
- 13. Wolfsdorf JI, Weinstein DA. Glycogen storage diseases. Rev Endocr Metab Disord 2003; 4: 95-102.
- 14. Chou JY, Jun HS, Mansfield BC. Glycogen storage disease type I and G6Pase-beta deficiency: etiology and therapy. Nat Rev Endocrinol 2010; 6: 676-88.
- Soggia AP, Correa-Giannella ML, Fortes MAH, et al. A novel mutation in the glycogen synthase 2 gene in a child with glycogen storage disease type 0. BMC Med Genet 2010; 11: 3.
- 16. Chial H. Rare genetic disorders: learning about genetic disease through gene mapping, snps, and microarray data. Nature Education 2008; 1: 192.
- Özen H. Glycogen storage diseases: new perspectives, World J Gastroenterol 2007; 13: 2541-53.
- Kim J, Parikh P, Mahboob M, et al. Asymptomatic young man with Danon disease. Tex Heart Inst J 2014; 41: 332-4.
- 19. Orho M, Bosshard NU, Buist NR, et al. Mutations in the liver glycogen synthase gene in children with hypoglycemia due to glycogen storage disease type 0. J Clin Invest 1998; 102: 507-15.
- Brown LM, Corrado MM, van der Ende RM, et al. Evaluation of glycogen storage disease as a cause of ketotic hypoglycemia in children. J Inherit Metab Dis 2015; 38: 489-93.
- 21. Shin YJ. Glycogen storage desease: clinical, biochemical, and molecular heterogeneity. Semin Peditar Neurol 2006; 13: 115-20.
- 22. Shirazi N, Kalra BP, Bhat NK, et al. Embryonal hepatoblastoma with co-existent glycogen storage disease in a seven-month-old child. J Clin Diagn Res 2016; 10: ED09-10.
- Lee KJ, Choi SJ, Kim WS, et al. Esophageal stricture secondary to candidiasis in a child with glycogen storage disease 1b. Pediatr Gastroenterol Hepatol Nutr 2016; 19: 71-5.
- 24. Lu C, Qiu Z, Sun M, et al. Spectrum of AGL mutations in Chinese patients with glycogen storage disease type III: identification of 31 novel mutations. J Hum Genet 2016; 61: 641-5.
- 25. Raben N, Sherman JB. Mutations in muscle phosphofructokinase gene. Hum Mutat 1995; 6: 1-6.
- Fernandes J, Pikaar NA. Hyperlipemia in children with liver glycogen disease. Am J Clin Nutr 1969; 22: 617-27.
- Hoffmann GF, Smit PA, Schoser B. Glycogen storage diseases of all types. J Inherit Metab Dis 2015; 38: 389-90.
- 28. Clayton PT. Diagnosis of inherited disorders of liver metabolism. J Inherit Metab Dis 2003; 26: 135-46.
- 29. Fernandes J, Pikaar NA. Ketosis in hepatic glycogenosis. Arch Dis Child 1972; 47: 41-6.
- 30. Kishnani PS, Austin SL, Abdenur JE, et al. Diagnosis and management of glycogen storage disease type I: a practice guideline of the American College of Medical Genetics and Genomics. American College of Medical Genetics and Genomics. Genet Med 2014; 16: e1.
- Kim H, Zheng Z, Walker PD, Kapatos G, Zhang K. CREBH maintains circadian glucose homeostasis by regulating hepatic glycogenolysis and gluconeogenesis. Mol Cell Biol 2017; 37: pii: e00048-17.
- 32. Brushia RJ, Walsh DA. Phosphorylase kinase: the complexity of its regulation is reflected in the complexity of its structure. Front Biosci 1999; 4: D618-41.

- Hendrickx J, Willems PJ. Genetic deficiences of glycogen phosphorylase system. Hum Genet 1996; 97: 551-6.
- 34. Burwinkel B, Shiomi S, Al Zaben A, et al. Liver glycogenosis due to phosphorylase kinase deficiency: PHKG2 gene structure and mutations associated with cirrhosis. Hum Mol Genet 1998; 7: 149-54.
- Burwinkel B, Tanner MS, Kilimann MW. Phosphorylase kinase deficient liver glycogenosis: progression to cirrhosis in infancy associated with PHKG2 mutations (H144Y and L225R). J Med Genet 2000; 37: 376-7.
- 36. Francke U, Darras TB, Zander NF, Kilimann MW. Assignment of human genes for phosphorylase kinase subunits alpha (PHKA) to Xq12-q13 and beta (PHKB) to 16q12-q13. Am J Hum Genet 1989; 45: 276-82.
- 37. Choi R, Park HD, Kang B, et al. PHKA2 mutation spectrum in Korean patients with glycogen storage disease type IX: prevalence of deletion mutations BMC Med Genet 2016; 17: 33.
- Carrière C, Jonic S, Mornon JP, et al. 3D mapping of glycogenosis-causing mutations in the large regulatory alpha subunit of phosphorylase kinase. Biochim Biophys Acta 2008; 1782: 664-70.
- Tsilianidis LA, Fiske LM, Siegel S, et al. Aggressive therapy improves cirrhosis in glycogen storage disease type IX. Mol Genet Metab 2013; 109: 179-82.
- 40. Bali DS, Goldstein JL, Fredrickson K, et al. Variability of disease spectrum in children with liver phosphorylase kinase deficiency caused by mutations in the PHKG2 gene. Mol Genet Metab 2014; 111: 309-13.
- 41. Davit-Spraul A, Piraud M, Dobbelaere D, et al. Liver glycogen storage diseases due to phosphorylase system deficiencies: diagnosis thanks to non-invasive blood enzymatic and molecular studies. Mol Genet Metab 2011; 104: 137-43.
- 42. Kagalwalla AF, Kagalwalla YA, Al Ajaj S, et al. Phosphorylase b kinase deficiency glycogenosis with cirrhosis of the liver. J Pediatr 1995; 127: 602-5.
- 43. Moses SW, Parvari R. The variable presentations of glycogen storage disease type IV: a review of clinical, enzymatic and molecular studies. Curr Mol Med 2002; 2: 177-88.
- 44. Rudolfová J, Slovácková R, Trbusek M, et al. Identification of three novel mutations in the PHKA2 gene in Czech patients with X-linked liver glycogenosis. J Inherit Metab Dis 2001; 24: 85-7.
- Shen JJ, Chen YT. Molecular characterization of glycogen storage disease type III. Curr Mol Med 2002; 2: 167-75.
- 46. Lucchiari S, Fogh I, Prelle A, et al. Clinical and genetic variability of glycogen storage disease type IIIa: seven novel AGL gene mutations in the Mediterranean area. Am J Med Genet 2002; 109: 183-9.
- 47. Demo E, Frush D, Gottfried M, et al. Glycogen storage disease type III-hepatocellular carcinoma a long-term complication? J Hepatol 2007; 46: 492-8.
- Dagli AI, Weinstein DA. In: Adam MP, Ardinger HH, Pagon RA, et al. (eds.). GeneReviews<sup>®</sup> [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2019. 2009 Apr 23 [updated 2011 May 17].
- 49. Roscher A, Patel J, Hewson S, et al. The natural history of glycogen storage disease types VI and IX: long-term outcome from the largest metabolic center in Canada. Mol Genet Metab 2014; 113: 171-6.
- Magoulas PL, El-Hattab AW. In: Adam MP, Ardinger HH, Pagon RA, et al. (eds.). GeneReviews<sup>®</sup> [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2019. 2013 Jan 3.

- Paradas C, Akman HO, Ionete C, et al. Branching enzyme deficiency: expanding the clinical spectrum. JAMA Neurol 2014; 71: 41-7.
- 52. van Schaftingen E, Gerin I. The glucose-6-phosphatase system. Biochem J 2002; 362: 513-32.
- 53. Marandel L, Panserat S, Plagnes-Juan E, et al. Evolutionary history of glucose-6-phosphatase encoding genes in verte brate lineages: towards a better understanding of the functions of multiple duplicates. BMC Genomics 2017; 18: 342.
- 54. Mithieux G. New knowledge regarding glucose-6 phosphatase gene and protein and their roles in the regulation of glucose metabolism. Eur J Endocrinol 1997; 136: 137-45.
- 55. van de Werve G, Lange AJ, Newgard CB, et al. New lessons in the regulation of glucose metabolism taught by the glucose-6-phosphatase system. Eur J Biochem 2000; 267: 1533-49.
- Foster JD, Pederson BA, Nordlie RC. Glucose-6-phosphatase structure, regulation, and function: an update. Proc Soc Exp Biol Med 1997; 203: 274-85.
- 57. Froissart R, Piraud M, Boudjemline AM, et al. Glucose-6-phosphatase deficiency. Orphanet J Rare Dis 2011; 6: 27.
- Wolfsdorf JI, Crigler JF Jr. Cornstarch regimens for nocturnal treatment of young adults with type I glycogen storage disease. Am J Clin Nutr 1997; 65: 1507-11.
- 59. Weinstein DA, Wolfsdorf JI. Effect of continuous glucose therapy with uncooked cornstarch on the long-term clinical course of type 1a glycogen storage disease. Eur J Pediatr 2002; 161: 35-9.
- 60. Shah KK, O'Dell SD. Effect of dietary interventions in the maintenance of normoglycaemia in glycogen storage disease type 1a: a systematic review and meta-analysis. J Hum Nutr Diet Off J Br Diet Assoc 2013; 26: 329-39.
- Bhattacharya K, Orton RC, Qi X, et al. A novel starch for the treatment of glycogen storage diseases. J Inherit Metab Dis 2007; 30: 350-7.
- 62. Correia CE, Bhattacharya K, Lee PJ, et al. Use of modified cornstarch therapy to extend fasting in glycogen storage disease types Ia and Ib. Am J Clin Nutr 2008; 88: 1272-6.
- Nakai A, Shigematsu Y, Takano T, et al. Uncooked cornstarch treatment for hepatic phosphorylase kinase deficiency. Eur J Pediatr 1994; 153: 581-3.
- Kishnani PS, Austin SL, Arn P, et al. ACMG. Glycogen storage disease type III diagnosis and management guidelines. Genet Med 2010; 12: 446-63.
- 65. Weinstein DA, Correia CE, Saunders AC, et al. Hepatic glycogen synthase deficiency: an infrequently recognized cause of ketotic hypoglycemia. Mol Genet Metab 2006; 87: 284-8.
- Sliwinska A, Kasinska MA, Drzewoski J. MicroRNAs and metabolic disorders – where are we heading? Arch Med Sci 2017; 13: 885-96.
- 67. Nathwani AC, Tuddenham EG, Rangarajan S, et al. Adenovirus-associated virus vector-mediated gene transfer in hemophilia B. N Engl J Med 2011; 365: 2357-65.
- Yiu WH, Lee YM, Peng WT, et al. Complete normalization of hepatic G6PC deficiency in murine glycogen storage disease type Ia using gene therapy. Mol Ther 2010; 18: 1076-84.